

CONTENTS

LEGAL NOTICES

GENERAL NOTICES

PREFACE

INTRODUCTION

MONOGRAPHS

Arka

General Description	1
1. AJAMODĀ ARKA	2
2. BRĀHMĪ ARKA	4
3. GULĀBA ARKA	6
4. JAṬĀMĀMSĪ ARKA	8
5. KĀKAMĀCĪ ARKA.....	10
6. MUṆḌĪTIKĀ ARKA.....	12
7. NĪLOḌUPUṢPA ARKA.....	14
8. PARPAṬA ARKA.....	16
9. PUDĪNĀ ARKA	18
10. PUNARNAVĀ ARKA.....	20
11. ŚATĀHVĀ ARKA	22
12. YAVĀNĪ ARKA	24

Avaleha

General Description	26
13. AŚVAGANDHĀDI LEHYA	27
14. HARIDRĀ KHAṆḌA	29
15. NĀRIKELA KHAṆḌA.....	32

Cūrṇa

General Description	34
16. CITRAKĀDI CŪRṆA.....	35

Ghṛta	38
General Description	
17. SUKUMĀRA GHṚTA	40
Guggulu	
General Description	43
18. SAPTĀṄGA GUGGULU.....	44
19. VARĀDI GUGGULU.....	47
20. VIḌAṄGĀDI GUGGULU	50
Taila	
General Description	53
21. AṆU TAILA.....	55
22. APĀMĀRGA KṢĀRA TAILA.....	58
23. ARIMEDĀDI TAILA	60
24. ASANABILVĀDI TAILA.....	64
25. BALĀ TAILA	67
26. BALĀHAṬHĀDI TAILA	71
27. BHRṆGARĀJA TAILA.....	73
28. BRĤAT SAINDHAVĀDYA TAILA.....	75
29. CITRAKĀDI TAILA	78
30. HIṄGVĀDI TAILA.....	80
31. JYOTIṢMATĪ TAILA	82
32. KANAKA TAILA.....	84
33. MAHĀNĀRĀYAṆA TAILA	86
34. NĀLPĀMARĀDI TAILA.....	90
35. NĪLĪBHRṆGĀDI TAILA	92
36. PAṆCAGUṆA TAILA.....	94
37. PRABHAṆJANA VIMARDANA TAILA.....	96
38. PRASĀRIṆĪ TAILA	99

39. TUVARAKA TAILA.....	102
40. YAṢṢĪMADHUKA TAILA.....	104

Vaṭī

General Description	106
41. ARKA VAṢĪ.....	107
42. CITRAKĀDI GUṢĪKĀ.....	109
43. ELĀDI GUṢĪKĀ.....	112
44. LAŚUNĀDI VAṢĪ.....	115
45. LAVANĠĀDI VAṢĪ.....	118
46. PLĪHĀRI VAṢĪKĀ.....	120
47. PRABHĀKARA VAṢĪ.....	123
48. RAJAḠPRAVARTINĪ VAṢĪ.....	125
49. SAÑĠĪVANĪ VAṢĪ.....	128
50. ŚAÑKHA VAṢĪ.....	131
51. PUNARNAVĀDI MAṢḠŪRA (TABLET).....	135

Appendix-1. Apparatus for Tests & Assays

1.1. Nessler Cylinders.....	
1.2. Sieves.....	
1.3. Thermometers.....	
1.4. Ultraviolet Lamp (For general purposes and for chromatography work).....	
1.5. Volumetric Glassware.....	
1.6. Weights and Balances.....	

Appendix-2. Tests and Determinations

1.7. Muslin Cloth.....	
2.1. Microscopic identification:.....	
2.2. Determination of Quantitative Data:.....	
2.2.1. Net Content.....	

2.2.2.	Foreign Matter
2.2.3.	Determination of Total Ash
2.2.4.	Determination of Acid-Insoluble Ash
2.2.5.	Determination of Water Soluble Ash
2.2.6.	Determination of Sulphated Ash.....
2.2.7.	Determination of Alcohol Soluble Extractive.....
2.2.8.	Determination of Water Soluble Extractive
2.2.9.	Determination of Ether Soluble Extractive (Fixed Oil Content).....
2.2.10.	Determination of Moisture Content (Loss on Drying)
2.2.11.	Determination of Volatile Oil in Drugs
2.2.12.	Special Processes Used in Alkaloidal Assays
2.2.13.	Thin-Layer Chromatography (TLC).....
2.2.14.	Fatty oil estimation
2.3.	Limit Tests:
2.3.1.	Limit Test for Arsenic.....
2.3.2.	Limit Test for Chlorides:
2.3.3.	Limit Test for Heavy metals:
2.3.4.	Limit Test for Iron
2.3.5.	Limit Test for Lead.....
2.3.6.	Limit Test for Sulphates:
2.3.7.	Heavy Metals by Atomic absorption spectrophotometry:
2.4.	Microbial Limit Tests:
2.4.1.	Total Aerobic Microbial Count:
2.4.2.	Tests for Specified Micro-organisms:
2.5.	Pesticide Residue:
2.5.1.	Qualitative and Quantitative Analysis of Pesticide Residues:
2.5.2.	Test for Pesticides:
2.5.3.	Quantitative Analysis:.....
2.6.	Test for Aflatoxins:
2.7.	Gas Chromatography:
2.8.	Test for the Absence of Methanol:

Appendix-3. Physical Tests and Determinations

3.1.	Refractive Index:
3.2.	Weight per Millilitre and Specific Gravity:
3.3.	Determination of pH Values:
3.4.	Determination of Melting Range and Congealing Range:
3.4.1.	Determination of Melting Range:
3.4.2.	Determination of Congealing Range:

- 3.5. Determination of Boiling Range:
- 3.6. Determination of Optical Rotation and Specific Optical Rotation:
- 3.7. Determination of Viscosity:
- 3.8. Determination of Total Solids:
- 3.9. Solubility in Water:
- 3.10. Determination of Saponification Value:
- 3.11. Determination of Iodine Value:
- 3.12. Determination of Acid Value:
- 3.13. Determination of Peroxide Value:
- 3.14. Determination of Unsaponifiable Matter:
- 3.15. Detection of Mineral Oil (Holde's Test):
- 3.16. Rancidity Test (Kreis Test):
- 3.17. Determination of Alcohol Content:
- 3.18. Disintegration test for tablets:
- 3.19. Uniformity of Weight of Single Dose Preparations

Appendix-4. Regents and Solutions

Appendix-5. Assays and Chemical Tests

- 5.1. Estimation of Sugars
- 5.2. Determination of Aluminum:
- 5.3. Determination of Borax:
- 5.4. Determination of Calcium:
- 5.5. Determination of Copper:
- 5.6. Determination of Iron (Fe).....
- 5.7. Determination of Magnesium:.....
- 5.8. Determination of Mercury:
- 5.9. Determination of Silica (SiO₂)
- 5.10. Estimation of Sodium and Potassium by Flame Photometer:.....
- 5.11. Determination of Sodium Chloride:
- 5.12. Determination of Sulphur:
- 5.13. Qualitative Reactions :
- 5.13.1. Sodium

5.13.2.	Potassium
5.13.3.	Magnesium
5.13.4.	Carbonates and Bicarbonates
5.13.5.	Sulphates
5.13.6.	Chlorides
5.13.7.	Calcium
5.13.8.	Sulphides
5.13.9.	Test for Mercury
5.13.10.	Test for Boron
5.13.11.	Test for Sulphur
5.13.12.	Test for Anthraquinones
5.13.13.	Test for Hingu

Appendix-6. Ayurvedic Definitions and Methods

6.1.Kalpanā Paribhāṣā:

6.1.1.	Kalka:
6.1.2.	Kvātha / Kaṣāya:
6.1.3.	Cūrṇa:
6.1.4.	Puṭapāka Svarasa:
6.1.5.	Svarasa:
6.1.6.	Hima Kaṣāya:

6.2.Sāmānya paribhāṣā:

6.2.1.	Kajjalī:
6.2.2.	Kāñjika:
6.2.3.	Kṣāra Preparation:
6.2.4.	Cūrṇodaka:
6.2.5.	Mastu Preparation:
6.2.6.	Prakṣepa:
6.2.7.	Bhāvanā:
6.2.8.	Śodhana:
6.2.8.1.	Godantī Śodhana:
6.2.8.2.	Gairika Śodhana:
6.2.8.3.	Gandhaka Śodhana:
6.2.8.4.	Guggulu Śodhana:
6.2.8.5.	Ṭaṅkaṇa Śodhana:
6.2.8.6.	Tuttha Śodhana:
6.2.8.7.	Bhallātaka Śodhana:

6.2.8.8.	Manahśilā Śodhana:
6.2.8.9.	Vatsanābha Śodhana:
6.2.8.10.	Karavīra Śodhana:.....
6.2.8.11.	Citraka Śodhana:
6.2.8.12.	Lāṅgalī Śodhana:
6.2.8.13.	Śilājatu Śodhana:.....
6.2.8.14.	Haritāla Śodhana:
6.2.8.15.	Hingu Śodhana:.....
6.2.8.16.	Vijayā Śodhana:
6.2.8.17.	Kāśīśa Śodhana:
6.2.8.18.	Sauvīrāñjana Śodhana:
6.2.8.19.	Narasāra Śodhana:.....
6.2.8.20.	Pārada Sāmānya Śodhana:
6.2.8.21.	Aṣṭasamskāra of Pārada.....
6.2.8.21.a.	Svedana:
6.2.8.21.b.	Mardana:
6.2.8.21.c.	Mūrcchana:
6.2.8.21.d.	Utthāpana:.....
6.2.8.21.e.	Pātana:
6.2.8.21.f.	Rodhana / Bodhana:
6.2.8.21.g.	Niyāmana:.....
6.2.8.21.h.	Dīpana / Sandīpana:.....
6.2.9.	Mūrcchanā:
6.2.9.1.	Mūrcchanā of Eraṇḍa Taila:.....
6.2.9.2.	Mūrcchanā of Ghṛta:
6.2.9.3.	Mūrcchana of Sarṣapa Taila:
6.2.9.4.	Mūrcchana of Tila Taila:
6.3. Yantra Paribhāṣā:	
6.3.1.	Khalva yantra:.....
6.3.2.	Tiryak pātana yantra:
6.3.4.	Dolāyantra:.....

Appendix-7. Weight and Measures

- 7.1. Metric Equivalents of Classical Weights and Measures
- 7.2. Metric System

Appendix-8. Classical Ayurvedic References

Appendix-9. List of Single Drugs used in Formulations

- 9.1. List of Single Drugs of Animal origin used in Formulations, with equivalent English name
- 9.2. List of Single Drugs of metal/mineral origin used in Formulations
- 9.3. List of Single Drugs of plant origin used in Formulation, with Latin Nomenclature

Appendix-10. Bibliography

ARKA

General Description:

Arka is a liquid preparation obtained by distillation of certain liquids or drugs soaked in water using the *Arkayantra* or any convenient modern distillation apparatus.

General Method of preparation:

The drugs are cleaned and coarsely powdered. Some quantity of water is added to the drugs for soaking and kept over-night. This makes the drugs soft and when boiled releases the essential volatile principles easily. The following morning it is poured into the *Arkayantra* and the remaining water is added and boiled. The vapour is condensed and collected in a receiver. In the beginning, the vapour consists of only steam and may not contain the essential principles of the drugs. It should therefore be discarded. The last portion also may not contain therapeutically essential substance and should be discarded. The aliquots collected in between contain the active ingredients and may be mixed together to ensure uniformity of the *Arka*.

Characteristics:

Arka is a suspension of the distillate in water having slight turbidity and colour according to the nature of the drugs used and smell of the predominant drug.

AJAMODĀ ARKA

(AFI Part III, 2:15)

Definition:

Ajamodā Arka is a liquid preparation obtained by hydro-distillation of fruits of *Trachyspermum roxburghianum*.

Formulation Composition:

1	Dvīpāntara ajamodā (API)	<i>Trachyspermum roxburghianum</i>	Fr.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	20.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through 355 µm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of Dvīpāntara ajamodā powder in a round bottom standard joint flask of 3.0 l capacity. Add 2.0 l of *Jala*.
- Attach the proper distillation assembly with distillation and receiving heads, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 1.5 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Ajamodā Arka is a cloudy milky turbid liquid having characteristic spicy odour with slightly pungent lingering bitter taste.

Identification:

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained under assay in 1 ml of *toluene* or in any suitable solvent. Dissolve separately 0.1 mg of each of *β-cycloavandulal* and *seslin* in 1 ml of *toluene* separately. Apply 2 µl of the solution of the oil and reference solutions on TLC plate precoated with silica gel 60 F₂₅₄ of 0.2 mm thickness. Develop the plate to a distance of 8 cm using *ethyl acetate: hexane* (2: 8) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.22 (greenish-blue corresponding to *sesline*), 0.44 (peach coloured, corresponding to *β-cycloavandulal*), 0.51 (pink), 0.58 (red-orange) and 0.67 (bluish-purple) in visible light.

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column walls coated with FFAP maintained at 90⁰ for 2 min then programmed at the rate of 7⁰/min to 220⁰ with injection port at 240⁰ and detector at 260⁰ and with a flow rate of carrier gas 1.5 ml/min.

Inject separately 0.1 µl each of oil obtained by hydro-distillation of the crude drug as well as of the *Arka* under assay along with reference.

Both the chromatograms show major peaks at R_t 7.25 corresponding to *limonene*, 12.60 corresponding to *seslin*, 15.65 corresponding to *β-cycloavandulal* and 16.69 corresponding to *cadinene*.

Physico- Chemical parameters:

Specific gravity (20⁰): 0.995 to 0.998, Appendix 3.2

ASSAY:

Ajamodā Arka contains 0.20 to 0.30 per cent of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2, 11).

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Agnimāndya (digestive impairment); Ajīrṇa (dyspepsia); Bastiroga (urinary bladder disorder); Vātakapharoga (diseases due to Vāta and Kapha doṣa)

Dose:

10 to 20 ml per day in divided doses

BRĀHMĪ ARKA

(AFI Part III, 2:11)

Definition:

Brāhmī Arka is a liquid preparation obtained by hydro-distillation of whole plant of *Bacopa monnieri*.

Formulation Composition:

1	Brāhmī API	<i>Bacopa monnieri</i>	Pl.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	30.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 μm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Brāhmī* powder in a round bottom standard joint flask of 5.0 l capacity. Add 3.0 l of *Jala*.
- Attach the proper distillation assembly, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 2.0 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Brāhmī Arka is a turbid pale yellow liquid with a faint odour and slightly astringent taste.

Identification

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation of the *Arka* in 1 ml of toluene or in any suitable solvent. In a separate setup extract *Brāhmī* oil from *Brāhmī* API. Apply 2 μl each of the solutions on TLC plate separately and develop the plate to a distance of 8 cm using *toluene: ethyl acetate* (97: 3) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. and examine under ultraviolet light at 254 nm. Both the chromatograms show major spots at R_f 0.17 (purple brown), 0.35 (red brown) and 0.63 (saffron red).

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column walls coated with BP-10 maintained at 150⁰ for 2 min. programmed at the rate of 7⁰/min to 200⁰, again 15⁰/17 min.

to 260⁰ and detector at 300⁰ with a flow rate of carrier gas 1.5 ml/ min. with injection port temperature 280⁰.

Inject separately 0.1µl of oil obtained by hydro-distillation of the crude drug as well as of *Arka* and programme the column as given in preceding paragraph.

Both the chromatograms show major peaks at R_t 14.12, 20.42, 14.23 and 21.78.

Physico-chemical parameters:

Specific gravity (20⁰): 0.998 to 1.0, Appendix 3.2

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Buddhimandatā (mental retardation), Smṛtibhrama (impaired memory)

Dose:

10 to 20 ml per day in divided doses

GULĀBA ARKA

(AFI Part III, 2:6)

Definition:

Gulāba Arka is a liquid preparation obtained by hydro-distillation of dried petals of *Rosa damascena*.

Formulation Composition

1	Gulāba (API)	<i>Rosa damascena</i>	Dry petals	1.0 kg
2	Jala API for soaking and for Preparation of Arka	Potable Water	–	12.5 l

Method of preparation:

- Take the raw material of pharmacopoeial quality and wash.
- Place 100 g of *Gulāba* petals in a round bottom standard joint flask of 3.0 l capacity. Add 1.25 l of *Jala*.
- Attach the proper distillation assembly with distillation and receiving heads, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 1.0 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Gulāba Arka is a hazy liquid with a pleasing odour of rose flower with sweetish to slightly bitter taste.

Identification:

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation in 1 ml of *toluene*. Dissolve separately 0.1 ml each of *citronellol*, *geraniol* and *phenyl ethanol* in 1 ml each of *toluene*. Apply separately 2 µl each of the solution of the oil and reference solution on TLC plate precoated with silica gel 60 F₂₅₄ of 0.2 mm thickness and develop the plate to a distance of 8 cm using *ethyl acetate: hexane* (20: 80) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.42 (orange-red) corresponding to *geraniol* and 0.46 (pinkish-purple) corresponding to *citronellol* in visible light.

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column walls coated with FFAP maintained at 90⁰ for 2 min. then programmed at the rate of 7⁰/ min. to 220⁰ and detector at 260⁰ and with a flow rate of carrier gas 1.5 ml/min.

Inject separately 0.1 µl of oil obtained by hydro-distillation of drug under assay and, programme the column as given in preceding paragraph.

The chromatogram shows major peak at R_t 12.52 (corresponding to linalool), 16.64 (corresponding to *citronellol*), 17.34 (corresponding to *nerol*), 18.25 (corresponding to *geraniol*) and 19.68 (corresponding to *phenyl ethanol*).

Physico- Chemical parameters:

Specific gravity (20⁰): 1.0 Appendix 3.2

ASSAY:

Gulāba Arka contains 0.035 to 0.08 per cent v/v of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2.11).

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Dāha (burning sensation), Tṛṣṇā (thirst), Hṛllāsa (nausea), Netraroga (eye diseases)

Dose:

10 to 20 ml per day in divided doses

External use : 2-3 drops in each eye, 2/3 times a day

JAṬĀMAṢĪ ARKA

(AFI Part 1, 2:3)

Definition:

Jaṭāmaṣī Arka is a liquid preparation obtained by hydro-distillation of rhizomes of *Nardostachys jatamansi*.

Formulation Composition:

1	Jaṭāmaṣī API	<i>Nardostachys jatamansi</i>	Rz.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	25.0 l

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 µm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Jaṭāmaṣī* powder in a round bottom standard joint flask of 5.0 l capacity. Add 2.5 l of *Jala*.
- Attach the proper distillation assembly with double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 1.8 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Jaṭāmaṣī Arka is a liquid having a slight turbidity with a spicy, slightly pungent and lingering bitter taste.

Identification:

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained from the *Arka* under assay in 1 ml of *toluene* or in any suitable solvent. In a separate setup extract *Jaṭāmaṣī* oil from *Jaṭāmaṣī* API. Apply 4 µl each of the oil solutions on TLC plate separately and develop the plate to a distance of 8 cm using *toluene* (double run) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. Both the chromatograms show major spots at R_f 0.25 (bluish purple), 0.33 (blue), 0.42 (pink), 0.69 and 0.79 (both pinkish purple) in visible light.

Gas Chromatography:

Carry out the gas chromatography analysis procedure using a 30 m fused silica capillary column walls coated with FFAP maintained at 90⁰ for 2 min, then programmed at the rate of 7⁰/min to 220⁰ and detector at 260⁰ and with a flow rate of carrier gas 1.5 ml/ min.

Inject separately 0.1 µl of oil obtained by hydro-distillation of the crude drug as well as of the oil of the *Arka* under assay and, after 2 min. increase the temperature of the column to 220⁰ at a rate of 7⁰/min.

Both the chromatograms show major peaks at R_t 20.26, 21.61, 22.51 (corresponding to patchouli alcohol), 23.01, 24.06 and 26.22.

Physico-chemical parameters:

Specific gravity (20⁰): 0.995 to 1.0, Appendix 3.2

ASSAY:

Jaṭāmāṃsī *Arka* contains 0.04 to 0.06 per cent v/v of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2.11).

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Agnimāndya (digestive impairment); Arocaka (tastelessness); Mukhadaurgandhya (halitosis); Unmāda (mania/psychosis); Apsmāra (epilepsy)

Dose:

10 to 20 ml per day in divided doses

KĀKAMĀCĪ ARKA

(AFI Part III, 2:1)

Definition:

Kākamācī Arka is a liquid preparation obtained by hydro-distillation of fruits of *Solanum nigrum*.

Formulation Composition:

1	Kākamācī API	<i>Solanum nigrum</i>	Fr.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	30.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 µm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Kākamācī* powder in a round bottom standard joint flask of 5.0 l capacity. Add 3.0 l of *Jala*.
- Attach the proper distillation assembly, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts control and continue the distillation to collect about 2 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Kākamācī Arka is a light greenish liquid with slight turbidity, taste slightly bitter.

Identification

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation of the *Arka* in 1 ml of *toluene*. In a separate setup extract *Kākamācī* oil from *Kākamācī* API and dissolve 0.1 ml of oil in 1 ml of *toluene* or in any suitable solvent. Apply 2 µl each of the solution of oils on TLC plate separately and develop the plate to a distance of 8 cm using *toluene: ethyl acetate (97: 3)* as mobile phase. After development, allow the plate to dry in air and spray the plate with *acetic anhydride sulphuric acid reagent* followed by heating at 105° for about 10 min. Both the chromatograms show major spots at R_f 0.17 (yellowish Grey) and 0.63 (sky blue) in visible light.

Gas Chromatography:

Carry out Gas chromatography using a 30 m fused silica capillary column walls coated with BP-10 maintained at 150° for 2 min programmed at the rate of 7°/min to 200° again 15°/17 min to 260° and detector at 300° and with a flow rate of carrier gas 1.5 ml/ min with injection port temperature 260°.

Inject separately 0.1µl each of the oil obtained by hydro-distillation of the crude drug as well as of the *Arka* and programme the column as given above.

Both the chromatograms show major peaks at R_t 22.01, 23.53 and 23.75.

Physico-chemical parameters:

Specific gravity (20⁰): 0.998 to 1.0, Appendix 3.2

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Hrd roga (heart diseases), Yakṛdroga (liver disorders), Śoṭha (anasarca)

Dose:

10 to 20 ml per day in divided doses

MUṄḌĪTIKĀ ARKA

(AFI Part III, 2:12)

Definition:

Muṅḍītikā *Arka* is a liquid preparation obtained by hydro-distillation of flowers of *Sphaeranthus indicus*.

Formulation Composition:

1	Muṅḍītikā API	<i>Sphaeranthus indicus</i>	Fl.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	35.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 µm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Muṅḍītikā* powder in a round bottom standard joint flask of 5.0 l capacity. Add 3.5 l of *Jala*.
- Attach the proper distillation assembly with double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 2.0 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Muṅḍītikā Arka is a slightly turbid dark brownish liquid.

Identification:

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation of *Arka* in 1 ml of toluene or in any suitable solvent, similarly dissolve separately 0.1 ml of *Muṅḍītikā* oil obtained by hydro-distillation of *Muṅḍītikā* API. Apply 2 µl each of the solutions of the oil on TLC plate separately and develop the plate to a distance of 8 cm using *toluene: ethyl acetate* (97: 3) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min and examine under ultraviolet light at 254 nm. Both the chromatograms show major spots at R_f 0.16 (orange-red), 0.35 (red- brown) and 0.63 (saffron-red).

Gas Chromatography:

Carry out the gas chromatography using 30 m fused silica capillary column having walls coated with BP-10 maintained at 90⁰ for 2 min. then programmed at the rate of 7⁰/min to 220⁰, injection port temperature 240⁰ and detector at 260⁰ and with a flow rate of carrier gas 1.5 ml/min.

Inject 0.1 µl of oil obtained by hydro-distillation of the *Arka* and programme the column as given in preceding paragraph. The chromatogram shows peaks at R_t 42.15 (all mixed), 42.46 and 42.75.

Physico-chemical parameters:

Specific gravity (20⁰): 0.998 to 1.0, Appendix 3.2

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Plīhārti (splenic disorders), Meha (increased frequency and turbidity of urine), Vātārti (disorders due to vitiation of Vāta doṣa), Tvakroga (skin diseases), Aruṃṣikā (dandruff)

Dose:

10 to 20 ml per day in divided dose

NĪLOḌUPUṢPA ARKA

(AFI Part III, 2:8)

Definition:

Nīloḍupuṣpa Arka is a liquid preparation obtained by hydro-distillation of whole plant of *Borago officinalis*.

Formulation Composition:

1	Nīloḍupuṣpa (API)	<i>Borago officinalis</i>	Pl.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	30.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through 355 μm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Nīloḍupuṣpa* powder in a round bottom standard joint flask of 3.0 l capacity. Add 2.0 l of *Jala*.
- Attach the proper distillation assembly with distillation and receiving heads, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 1.5 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Nīloḍupuṣpa Arka is a slightly turbid pale liquid with a slightly spicy and sour taste.

Identification:

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation of *Nīloḍupuṣpa Arka* in 1 ml of *toluene* and 0.1 ml of authentic *Nīloḍupuṣpa* oil in 1 ml of *toluene* separately in any suitable solvent. Apply 2 μl each of the oil solution on TLC plate precoated with silica gel 60 F₂₅₄ of 0.2 mm thickness and develop the plate to a distance of 8 cm using *toluene: ethyl acetate* (97: 3) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. and examine under ultraviolet light at 254 nm. It shows major spots at R_f 0.17 (orange-red), 0.35 (red-brown) and 0.63 (saffron-red).

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column coated with BP-10 maintained at 150⁰ for 2 min. then programmed at the rate of 7⁰/ min. to 200⁰ and again 15⁰/17 min. to 260⁰ and detector at 300⁰ keeping flow rate of carrier gas 1.5 ml/min. with injection port temperature 280⁰.

Inject separately 0.1 µl of oil obtained by hydro-distillation of the *Arka* and programme the column as given in preceding paragraph. The chromatogram shows major peaks at R_t 11.94, 13.07, 13.83, 14.11 and 23.14.

Physico- Chemical parameters:

Specific gravity (20⁰): 0.998 to 1.0, Appendix 3.2

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Kapharoga (diseases due to vitiation of Kapha Doṣa), Kāsa (cough), Śvāsa (dyspnoea/asthma), Kaṇṭharoga (throat diseases)

Dose:

10 to 20 ml per day in divided doses

PARPAȚA ARKA

(AFI Part III, 2:9)

Definition:

Parpața Arka is a liquid preparation obtained by hydro-distillation of whole plant of *Fumaria vaillantii*.

Formulation Composition:

1	Parpața API	<i>Fumaria vaillantii</i> (= <i>F. parviflora</i>)	Pl.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	30.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 μm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Parpața* powder in a round bottom standard joint flask of 3.0 l capacity. Add 3.0 l of *Jala*.
- Attach the proper distillation assembly with distillation and receiving heads, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 2.0 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Parpața Arka is a slightly turbid light yellow liquid with a sweet odour.

Identification:

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation of the *Arka* in 1 ml of *toluene* or in any suitable solvent. In a separate setup extract *Parpața* oil from *Parpața* API and dissolve 0.1 ml of the oil in 1 ml of *toluene* or in any suitable solvent. Apply 2 μl each of the solution on TLC plate separately and develop the plate to a distance of 8 cm using *toluene* as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.17 (orange-red.), 0.35 (red-brown) and 0.63 (saffron-red) in visible light.

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column, walls coated with BP-10 maintained at 150⁰ for 2 min then programmed at the rate of 7⁰/min. to 200⁰, again 15⁰/17 min. to 260⁰ and detector at 300⁰ and with a flow rate of carrier gas 1.5 ml/min. with injection port temperature 280⁰.

Inject 0.1 µl of oil obtained by hydro-distillation of the *Arka* under assay and programme the column as given in preceding paragraph. The chromatogram shows major peak at R_t 23.67, 14.14.

Physico-chemical parameters:

Specific gravity (20⁰): 0.998 to 1.0, Appendix 3.2

Assay:

Parpaṭa Arka contains not less than 0.02 per cent v/v of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2.11).

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Jvara (fever), Tṛṣṇā (thirst), Atisāra (diarrhoea), Dāha (burning sensation)

Dose:

10 to 20 ml per day in divided doses

PUDĪNĀ ARKA

(AFI Part II, 2:1)

Definition:

Pudīnā Arka is a liquid preparation obtained by hydro-distillation of areial parts of *Mentha viridis*.

Formulation Composition:

1	Pudīnā API	<i>Mentha viridis</i>	A. Pt.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	30.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 μm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Pudīnā* powder in a round bottom standard joint flask of 2.0 l capacity. Add 1.4 l of *Jala*.
- Attach the proper distillation assembly with distillation and receiving heads, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 700 ml of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Pudīnā Arka is a slightly turbid liquid with a pleasant mint odour; slightly bitter to taste, producing a cooling sensation.

Identification:

Thin Layer Chromatography:

Dissolve separately 0.1 ml of the oil obtained from the *Arka* under assay and 0.1 g each of 1-*menthol*, *menthone* and *menthyl acetate* in 1 ml of *toluene* each. Apply 4 μl of the solution of oil and 2 μl each of reference solutions on TLC plate serarately. Develop the plate to a distance of 8 cm using *methanol: toluene* (1: 19) as mobile phase. After development, allow the plate to dry in air and spray the plate with *vanillin sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.15 (maroon purple corresponding to *menthol*), 0.30 (bluish purple, corresponding to *menthone*) and 0.45 (dark maroon purple, corresponding to *menthyl acetate*) in visible light.

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column, walls coated with FFAP maintained at 90⁰ for 2 min then programmed at the rate of 7⁰/ min to 220⁰, and detector at 260⁰ and with a flow rate of carrier gas 1.5 ml/min with injection port temperature 220⁰.

Inject 0.1 µl of oil obtained by hydro-distillation of the *Arka* under assay and programme the column as given in preceding paragraph. The chromatogram shows major peaks at R_t 6.37 (corresponding to *limonene*), 10.81 (corresponding to *menthone*), 11.3 (corresponding to *iso-menthone*), 12.82 (corresponding to *menthyl acetate*) and 13.97 (corresponding to 1-*menthol*).

Physico-chemical parameters:

Specific gravity (32⁰): 0.9831 to 1.0, Appendix 3.2

ASSAY:

Pudīnā Arka contains 0.14 to 0.18 per cent v/v of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2.11).

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Chardi (vomiting); Ajīrṇa (indigestion); Udaraśūla (abdominal pain); Agnimāndya (impaired digestive fire)

Dose:

10 to 20 ml per day in divided doses

PUNARNAVĀ ARKA

(AFI Part III, 2:10)

Definition:

Punarnavā Arka is a liquid preparation obtained by hydro-distillation of roots of *Boerhaavia diffusa*.

Formulation Composition:

1	Punarnavā (Rakta Punarnavā API)	<i>Boerhaavia diffusa</i>	Rt.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	–	35.0 l

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 µm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Punarnavā* powder in a round bottom standard joint flask of 5.0 l capacity. Add 3.5 l of *Jala*.
- Attach the proper distillation assembly, double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 2.0 l of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Punarnavā Arka is a slightly milky turbid liquid.

Identification

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained by hydro-distillation of the *Arka* in 1 ml of *toluene*. In a separate setup extract *Punarnavā* oil from *Punarnavā* API and dissolve 0.1 ml of oil in 1 ml of *toluene* or in any suitable solvent. Apply 2 µl each of the solution of oils on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate* (97: 3) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min and examine under ultraviolet light at 254 nm. Both the chromatograms show major spot at R_f 0.63 (saffron red).

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column walls coated with BP-10 maintained at 150⁰ for 2 min programmed at the rate of 7⁰/min to 200⁰ again 15⁰/17 min to

260⁰ and detector at 300⁰ and with a flow rate of carrier gas 1.5 ml/min with injection port temperature 260⁰.

Inject 0.1µl of oil obtained by hydro-distillation of the crude drug as well as oil of *Arka* and programme the column as given in preceding paragraph.

The chromatograms show major peaks at R_t 13.93 and 22.89.

Physico-chemical parameters:

Specific gravity (20⁰): 0.999 to 1.0, Appendix 3.2

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Jalodara (ascites), Śoṭha (anasarca), Netraroga (disorders of eyes)

Dose:

10 to 20 ml per day in divided doses

External use : 2-3 drops in each eye, 2/3 times a day

ŚATĀHVĀ ARKA

(AFI Part III, 2:14)

Definition:

Śatāhvā Arka is a liquid preparation obtained by hydro-distillation of fruits of *Anethum sowa*.

Formulation Composition:

1	Śatāhvā API	<i>Anethum sowa</i>	Fr.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	12.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 µm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of Śatāhvā powder in a round bottom standard joint flask of 3.0 l capacity. Add 1.2 l of Jala.
- Attach the proper distillation assembly with double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. Arka.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 700 ml of Arka.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Śatāhvā Arka is a slightly turbid liquid with pleasing fruity odour of fresh sowa fruits and sweetish spicy, slightly bitter taste.

Identification

Thin layer chromatography:

Dissolve 0.1 ml of the oil obtained from the Arka under assay in 1 ml of *toluene* or in any suitable solvent. Dissolve separately 0.1 g of *carvone* in 1 ml of *toluene*. Apply 2 µl each of the solution of oil and reference solution on TLC plate. Develop the plate to a distance of 8 cm using *toluene* (double run) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105° for about 10 min. It shows major spots at R_f 0.10 (blue purple), 0.32 (pink corresponding to *carvone*), 0.46 (pinkish blue), 0.55 (dark coke coloured) in visible light.

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column walls coated with BP-10 maintained at 90° for 2 min then programmed at the rate of 7°/min to 230° for 5 min and detector at 260° and with a flow rate of carrier gas 1.5 ml/min.

Inject separately 0.1 µl each of oil obtained by hydro-distillation of drug under assay and, *carvone* reference standard and programme the column as given above.

The chromatogram shows major peaks at R_t 5.2, 8.7, 10.4 and 11.5 (corresponding to *carvone*).

ASSAY:

Śatāhvā Arka contains 0.20 to 0.50 per cent v/v of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2.11).

Physico- Chemical parameters:

Specific gravity (20^o): 0.991 to 0.998, Appendix 3.2

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Jvara (fever); Vāta Kaphaja roga (diseases due to Vāta and Kapha doṣa); Vraṇaśūla (pain due to wound); Akṣiroga (diseases of eye), Agnimāndya (impairment of digestive power), Atisāra (diarrhoea)

Dose:

10 to 20 ml per day in divided doses

YAVĀNĪ ARKA

(AFI Part II, 2:2)

Definition:

Yavānī Arka is a liquid preparation obtained by hydro-distillation of fruits of *Trachyspermum ammi*.

Formulation Composition:

1	Yavānī API	<i>Trachyspermum ammi</i>	Fr.	1.0 kg
2	Jala API for soaking and for preparation of Arka	Potable Water	—	12.0 l

Method of preparation:

- Take the raw materials of pharmacopoeial quality.
- Wash, dry and powder the ingredient number 1 of the Formulation Composition and pass through the 355 μm I. S. sieve (sieve number 44) to obtain coarse powder.
- Place 100 g of *Yavānī* powder in a round bottom standard joint flask of 2.0 l capacity. Add 1.2 l of *Jala*.
- Attach the proper distillation assembly with double surface condenser and receiving flask and enough circulating water to condense the distillate i.e. *Arka*.
- Place the flask on a heating mantle. Adjust the temperature control when boiling starts and continue the distillation to collect about 700 ml of *Arka*.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Yavānī Arka is a colourless slightly cloudy liquid with the characteristic odour of *Yavānī* with a bitter burning taste

Identification:

Thin layer chromatography:

Dissolve separately 0.1 ml of the oil obtained from the *Arka* under assay and 0.1 g of *thymol* in 1 ml of *toluene*. Apply 4 μl of the solution of oil and 2 μl of reference solution on TLC plate. Develop the plate to a distance of 8 cm using *ethyl acetate: hexane* (3: 17) as mobile phase. After development, allow the plate to dry in air and observe under ultraviolet light (256 nm). The plate shows one blue fluorescent spot at R_f 0.54 (corresponding to *thymol*). Spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105° for about 10 min. It shows major spots at R_f 0.32 (purple), 0.54 (maroon red, corresponding to *thymol*) and 0.64 (bluish Grey) in visible light.

Gas Chromatography:

Carry out the gas chromatography using a 30 m fused silica capillary column walls coated with FFAP maintained at 90⁰ for 2 min. then programmed at the rate of 7⁰/min to 220⁰ and detector at 260⁰ and with a flow rate of carrier gas 1.5 ml/ min and injection port temperature at 260⁰.

Inject separately 0.1 µl each of oil obtained by hydro-distillation of drug under assay and, *γ-terpinene*, *p-cymene* and *thymol* reference standards and programme the column as given above.

The chromatogram shows major peaks at R_t 7.96 (corresponding to *γ-terpinene*), 8.32 (corresponding to *p-cymene*) and 23.93 (corresponding to *thymol*).

Physico- Chemical parameters:

Specific gravity (20⁰): 0.995 to 0.999, Appendix 3.2

ASSAY:

Yavānī Ārka contains 0.20 to 0.60 per cent of essential oil, determined for a well stirred quantity of not less than 2.0 l. (Appendix 2.2.11).

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Agnimāndya (digestive impairment); Trikaśūla (pain in sacral region)

Dose:

10 to 20 ml per day in divided doses

AVALEHA

General Description:

Avaleha or *Lehya* is a semi-solid preparation of drugs, prepared with addition of jaggery, sugar or sugar-candy and boiled with prescribed juices or decoction.

These preparations generally have

- (1) *Kaṣāya* or other liquids,
- (2) Jaggery, sugar or sugar-candy,
- (3) Powders or pulps of certain drugs,
- (4) Ghee or oil and
- (5) Honey.

Jaggery, sugar or sugar-candy is dissolved in the liquid and strained to remove the foreign particles. This solution is boiled over a moderate fire. When pressed between two fingers if *pāka* becomes thready (*Tantuvat*), or when it sinks in water without getting easily dissolved, it should be removed from the fire. Fine powders of drugs are then added in small quantities and stirred continuously to form a homogenous mixture. Ghee or oil, if mentioned, is added while the preparation is still hot and mixed well. Honey, if mentioned is added when the preparation becomes cool and mixed well.

The *Lehya* should neither be hard nor a thick fluid. When pulp of the drugs is added and ghee or oil is present in the preparation, this can be rolled between the fingers. When metals are mentioned, the *bhasmas* of the metals are used. In case of drugs like *Bhallātaka*, purification process is to be followed.

The *Lehya* should be kept in glass or porcelain jars. It can also be kept in a metal container which does not react with it. Normally, *Lehyas* should be used within one year.

AŚVAGANDHĀDI LEHYA

(AFI Part-I, 3:2)

Definition:

Aśvagandhādi lehya is a semisolid preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Śarkarā API	Sugar	–	1.356 kg
2	Aśvagandhā API	<i>Withania somnifera</i>	Rt.	192 g
3	Sārivā (Śveta sārivā API)	<i>Hemidesmus indicus</i>	Rt.	192 g
4	Jīvaka (Śveta jīraka API)	<i>Cuminum cyminum</i>	Fr.	192 g
5	Madhusnuhī API	<i>Smilax china</i> [<i>Smilax glabra</i> (Official Substitute)]	Rt. Tr.	192 g
6	Drākṣā API	<i>Vitis vinifera</i>	Dr. Fr.	192 g
7	Ghṛta (Goghṛta API)	Clarified butter from cow milk	–	226 g
8	Madhu API	Honey	–	452 g
9	Elā (Sūkṣmailā API)	<i>Elettaria cardamomum</i>	Sd.	24 g
10	Jala API	Potable Water	–	452 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Wash, clean, dry the ingredient number 2 to 5 and 9 of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85) to obtain fine powder.
- Wash, clean *Drākṣā*, soak in water till fully swollen and crush to make paste.
- Take *Ghṛta* in a stainless steel vessel and heat till it becomes free from moisture.
- Add soaked *Drākṣā* to the *Ghṛta* and fry it to make moisture free, then add powdered ingredients and fry till it turns to a soft bolus.
- Add sugar to water and heat, maintaining the temperature between 80⁰ and 90⁰. After the sugar dissolves, filter the hot syrup through *muslin cloth*.
- Add the fried paste to the syrup, heat with constant stirring, maintaining the temperature between 90⁰ and 100⁰ and observe the mixture for formation of soft bolus, which does not disperse in water. Stop heating and allow to cool.
- Add honey when it comes to room temperature.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

A blackish brown, semisolid paste with a spicy pleasant odour and bitter astringent taste

Identification:

Thin-layer chromatography:

Extract 5 g of *Avaleha* with 75 ml *n-hexane* (25 ml x 3) under reflux on a water bath for 30 min. Pool the extracts, filter and concentrate the filtrate to 10 ml and carry out thin layer chromatography. Apply 10 µl on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate* (9: 1) as mobile phase. After developing the plate, allow it to dry. Spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.10, 0.15, 0.26, 0.42 (all bluish grey), 0.54 and 0.70 (both purple) in visible light.

Physicochemical parameters:

<i>Total Ash:</i>	Not more than 2 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 1 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 19 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 46 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 28 per cent,	Appendix 2.2.10
<i>pH (1% aqueous solution):</i>	4.7 to 5.0,	Appendix 3.3

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Raktavikāra (disorders of blood), Kṛśatva (cachexia), Arśa (piles), Unamāda (psychosis), used as Balya (tonic), Rasāyana (rejuvenating agents), Vājīkara (aphrodisiasis)

Dose:

6 to 12 g with milk

HARIDRĀ KHAṆḌA

(AFI Part I, 3:31)

Definition:

Haridrā Khaṇḍa consists of granular material made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Haridrā API	<i>Curcuma longa</i>	Rz.	384 g
2	Haviṣ (Goghṛta API)	Clarified butter from cow milk	—	288 g
3	Kṣīra (Godugdha API)*	Cow milk	—	
4	Khaṇḍa (Śarkarā API)	Sugar	Syrup	2.400 kg
5	Trikaṭu			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	48 g
	b. Marica API	<i>Piper nigrum</i>	Fr.	48 g
	c. Pippalī API	<i>Piper longum</i>	Fr.	48 g
6	Trijāta			
	a. Tvak API	<i>Cinnamomum zeylanicum</i>	St. Bk.	48 g
	b. Sūkṣmailā API	<i>Elettaria cardamomum</i>	Sd.	48 g
	c. Tvakpatra API	<i>Cinnamomum tamala</i>	Lf.	48 g
7	Kṛmighna (Viḍaṅga API)	<i>Embelia ribes</i>	Fr.	48 g
8	Trivṛtā (Trivṛt API)	<i>Operculina turpethum</i>	Rt.	48 g
9	Triphalā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	48 g
	b. Bibhītaka API	<i>Terminalia bellerica</i>	P.	48 g
	c. Āmalakī API.	<i>Emblica officinalis</i>	P.	48 g
10	Keśara (Nāgakeśara API)	<i>Mesua ferrea</i>	Stmn.	48 g
11	Mustā API	<i>Cyperus rotundus</i>	Rt. Tr.	48 g
12	Lauha (Lauha bhasma (API))	Calcined Lauha	—	48 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Lauha* to prepare *Lauha bhasma*.
- Wash, clean, dry the ingredients numbered 5 to 11 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder and mix them all to a homogeneous mixture along with *Lauha bhasma*.
- Wash, clean, dry *Haridrā*, powder and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder.

* To prevent spoilage of the preparation, it is advisable not to use milk in the Formulation; instead the Formulation should be taken along with *Godugdha* as *Sahapāna*.

- Fry *Haridrā* powder in *Ghṛta* maintaining the temperature between 80⁰ to 90⁰ till *Haridrā* turns brown and its typical smell emanates.
- Prepare sugar syrup and filter while hot through *muslin cloth*.
- Add the fried *Haridrā* to the syrup, heat with constant stirring, maintaining the temperature at about 90⁰.
- Add mixture of fine powders and mix thoroughly to prepare a homogeneous mixture.
- Pass the mixture through a granulator to obtain granules of suitable size. Allow the mixture to cool to room temperature.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Yellowish to brown granular material with taste and odour characteristic of turmeric along with pungency

Identification:

Thin-layer Chromatography:

Extract 5 g of the formulation in 75 ml *methanol* (25 ml x 3) under reflux on a water bath for 30 min. Combine the extracts, filter and concentrate to 10 ml, and carry out thin layer chromatography. Apply 10 µl on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: methanol: formic acid* (3: 3: 0.8: 0.2) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light. The plate shows major spots at R_f 0.48, 0.57, 0.66, 0.76, 0.96 under 254 nm and fluorescent spots at R_f 0.13, 0.20, 0.30, 0.37, 0.71 (all blue); 0.55, 0.60, 0.76 (all yellow) under 366 nm. Spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.65, 0.72, 0.80, 0.86, (all purple) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 3 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 1 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 14 per cent,	Appendix 2.2.7
<i>Loss on drying:</i>	Not more than 6 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	3.3 to 3.7,	Appendix 3.3
<i>Total Sugar estimated as</i>	80 to 85 per cent,	Appendix to be prepared and
<i>Reducing Sugars:</i>		added
<i>Iron content (% w/w):</i>	Not more than 0.05 per cent,	Appendix 5.6

Assay:

Haridrā Khaṇḍa granular material contains 1.0 to 1.4 per cent of *curcumin* when determined by the following procedure.

Dissolve 4 mg accurately weighed *curcumin* in *methanol* in a 25 ml-volumetric flask and make up the volume. Transfer the aliquots (0.4 -1.4 ml) of stock solution to 10- ml volumetric flasks and

make up the volume with *methanol* to obtain standard solutions containing 6.4 to 22.4 µg/ml *curcumin*, respectively.

Apply 10 µl each of the standard solutions prepared above on precoated TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: methanol* (5: 0.5: 1) as mobile phase. Scan the plate in the TLC scanner at a wave length 429 nm. Record the peak area under curve for a peak corresponding to *curcumin* and plot the calibration curve by plotting peak area vs concentration of *curcumin*.

Weigh about 5 g, accurately weighed Haridrā Khaṇḍa and extract with *methanol* (25 ml x 4). Filter, pool the filtrates, concentrate and make up the volume to 25 ml with *methanol* in a volumetric flask.

Apply 5 µl of the sample solution on TLC plate and carry out thin layer chromatography. Develop, dry and scan the plate as described in preceding paragraph for calibration curve of *curcumin*. Calculate the amount *curcumin* in the sample solution from the calibration of *curcumin*.

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed amber coloured glass containers, protected from light and moisture.

Therapeutic uses:

Śītapitta (urticaria); Kaṇḍū (itching); Visphoṭa (blister); Dadru (Taeniasis); Udarda (urticaria); Koṭha (urticaria)

Dose:

6 g with milk or water

NĀRIKELA KHAṆḌA

(AFI Part I, 3:16)

Definition:

Nārikela Khaṇḍa is a solid Avaleha preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Nārikela API	<i>Cocus nucifera</i>	Enm.	192 g
2	Sarpi (Goghṛta API)	Clarified butter from cow milk	–	48 g
3	Khaṇḍa (Śarkarā API)	Sugar candy	–	192 g
4	Nārikela paya (Nārikela API)	<i>Cocus nucifera</i>	Tender Coconut water	768 g
5	Dhanyāka (Dhānyaka API)	<i>Coriandrum sativum</i>	Fr.	3 g
6	Pippalī API	<i>Piper longum</i>	Fr.	3 g
7	Payoda (Mustā API)	<i>Cyperus rotundus</i>	Rz	3 g
8	Tugā (Vaṃśalocana API)	Bamboo manna	S. C.	3 g
9	Dvijīra			
	a. Śveta jīraka API	<i>Cuminum cyminum</i>	Fr.	3 g
	b. Kṛṣṇa jīraka API	<i>Carum carvi</i>	Fr.	3 g
10	Trijāta			
	a. Tvak API	<i>Cinnamomum zeylanicum</i>	St. Bk.	3 g
	b. Tvalpatra API	<i>Cinnamomum tamala</i>	Lf.	3 g
	c. Sūkṣmailā API	<i>Elettaria cardamomum</i>	Sd.	3 g
11	Ibhakeśara (Nāgakeśara API)	<i>Mesua ferrea</i>	Stmn.	3 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Wash, clean, dry the ingredients numbered 5 to 11 of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85) to obtain fine powder and mix them all to a homogeneous mixture.
- Cut ingredient number 1 of the Formulation Composition into small pieces and grind to a paste.
- Fry the paste in *Ghṛta* maintaining the temperature between 80⁰ to 90⁰ till it turns brown and its typical smell emanates.
- Strain *Nārikela paya* through a *muslin cloth*.
- Add sugar to *Nārikela paya* and heat, maintaining the temperature between 80⁰ and 90⁰. After the sugar dissolves, filter the hot syrup through *muslin cloth*.
- Add the fried paste to the syrup, heat with constant stirring, maintaining the temperature about 90⁰ and observe the mixture for formation of soft bolus, which does not disperse in water. Stop heating and allow to cool to 50⁰.
- Add mixture of fine powders and mix thoroughly to prepare a homogeneous blend.

- Spread the paste on a plate greased with *Ghṛta* and cut into small diamond shaped pieces. Allow to cool it to room temperature.
- Store in containers and pack them air-tight to protect from light and moisture.

Description:

Solid brown polygonal pieces of various shapes and sizes, sweet with smell characteristic of coconut

Identification:

Thin-layer chromatography:

Extract 20 g of the formulation powder with 50 ml of *methanol* by refluxing on a water bath. Filter the extract and concentrate to 10 ml and carry out thin layer chromatography. Apply 20 µl of the extract on a TLC plate and develop the plate to distance of 8 cm using *toluene: ethyl acetate: formic acid: methanol* (6: 6: 1.6: 1.6) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light. The plate shows major spots at R_f 0.24, 0.71, 0.78 under 254 nm and four light fluorescent spots at R_f 0.11, 0.75, 0.78 (all blue), 0.72 (red) under 366 nm. Spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105° for about 10 minutes. The plate shows major spots at R_f 0.19 (black), 0.66 (purple), 0.73 (brown), 0.78 (green) and 0.84 (purple) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 3.0 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 1.0 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 40 per cent,	Appendix 2.2.7
<i>Loss on drying:</i>	Not more than 8.0 per cent,	Appendix 2.2.10
<i>pH (5 % aqueous solution):</i>	5.0 to 5.2,	Appendix 3.3
<i>Total Sugar estimated as Reducing Sugars:</i>	46 to 52 per cent,	Appendix to be prepared and added

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed amber coloured glass containers, protected from light and moisture.

Therapeutic uses:

Aruci (tastelessness); Vami (vomiting); Śūla (pain/colic); Amlapitta (hyperacidity); Raktapitta (bleeding disorder); Kṣata (wound); Kṣaya (Pthisis); Daurbalya (weakness)

Dose:

6 to 12 g with milk

CŪRṆA

General Description:

Drugs according to the formulation composition of the particular *Cūrṇa* are collected, dried, powdered individually and passed through 180 μm I. S. sieve to prepare a fine powder. They are mixed in the specified proportion and stored in well closed container.

The term *Cūrṇa* may be applied to the powder prepared by a single drug or a combination of more drugs.

Raja and *Kṣoda* are the synonyms for *Cūrṇa*. *Cūrṇas* may be of plant origin, or mixed with other ingredients. The following points are to be noted.

If metals / minerals are used, prepare *bhasma* or *sindūra* of the minerals unless otherwise mentioned.

In cases where *Pārada* and *Gandhaka* are mentioned, prepare *Kajjalī* and add other drugs, one by one, according to the formula.

In general the aromatic drugs like *Hīṅgu* [Asafoetida] etc. should be fried before they are converted to fine powders.

Specific care should be taken in case of Salts and Sugars. Formulations with hygroscopic components should not usually be prepared during rainy seasons. If so, specific precautions should be taken during storage.

Cūrṇas should be stored in air tight containers. Polyethylene and foil packing also provides damp proof protection.

Special precaution for storage should be taken in cases of formulations with salts, sugars and *Kṣāras*.

CITRAKĀDI CŪRṆA

(AFI Part-I, 7:11)

Definition:

Citrakādi Cūrṇa is a powder preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Citraka API	<i>Plumbago zeylanica</i>	Rt.	12 g
2	Nāgara (Śuṅṭhī API)	<i>Zingiber officinale</i>	Rz.	12 g
3	Hīṅgu API	<i>Ferula foetida</i>	Exd.	12 g
4	Pippalī API	<i>Piper longum</i>	Fr.	12 g
5	Pippalījaṭā (Pippalī API)	<i>Piper longum</i>	St.	12 g
6	Cavya API	<i>Piper retrofractum</i>	St.	12 g
7	Ajamodā API	<i>Apium leptophyllum</i>	Fr.	12 g
8	Marica API	<i>Piper nigrum</i>	Fr.	12 g
9	Svarjikā (Svarkīkṣāra (API))	Crude alkaline earth	–	6 g
10	Yavakṣāra (API)	<i>Hordeum vulgare</i>	Water soluble Ash of Pl.	6 g
11	Sindhu (Saindhava Lavaṇa (API))	Rock salt	–	6 g
12	Sauvarcala (Sauvarcala Lavaṇa (API))	Black salt	–	6 g
13	Viḍa Lavaṇa			6 g
14	Sāmudraka (Sāmudra Lavaṇa API)	Sea salt	–	6 g
15	Romaka Lavaṇa			6 g
16	Mātuluṅga API - rasa	<i>Citrus medica</i>	Fr. juice	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Hīṅgu* to prepare *Hīṅgu - Śuddha* (Appendix 6.2.8.15).
- Roast *Svarjikā kṣāra* and *Yava kṣāra* in a stainless steel pan on low flame till free from moisture.
- Roast coarsely powdered *Sauvarcala*, *Saindhava*, *Sāmudra* and *Viḍa Lavaṇas* in a stainless steel pan on low flame till free from moisture, powder separately and pass through 180 µm I. S. sieve (sieve number 85).
- Wash, clean, dry the ingredients numbered 1, 2 and 4 to 8 of the Formulation Composition and powder separately. The powders should completely pass through 355 µm I. S. sieve (sieve number 44) and not less than 50 per cent pass through through 180 µm I. S. sieve (sieve number 85).
- Weigh each ingredient separately and mix together. Pass the *cūrṇa* through 355 µm I. S. sieve (sieve number 44) to obtain a homogenous blend.
- Cut and squeeze the *Mātuluṅga* fruits and filter the juice through muslin cloth to obtain *Mātuluṅga rasa*.

- Soak the powder mixture in *Mātuluṅga rasa* in a ceramic vessel and dry under sunlight till the powder absorbs all the juice.
- After complete drying pass the *cūrṇa* through 355 μm I. S. sieve (sieve number 44).
- Store in the container and pack it air-tight.

Description:

- Brown-coloured, smooth powder with pleasant odour, sour, spicy and pungent taste. The powder completely passes through 355 μm I. S. sieve (sieve number 44) and not less than 50 per cent pass through 180 μm I. S. sieve (sieve number 85).

Identification:

Microscopy:

Take about 2 g of *Curṇa*, and wash it with water thoroughly to remove salt without loss of *Curṇa*. Remove water and use the washed *Curṇa* for the following mounts: warm a few mg of material with chloral hydrate, wash and mount in glycerin; treat a few mg with iodine in potassium iodide solution and mount in glycerin; heat a few mg in 2 per cent aqueous potassium hydroxide, wash in water and mount in glycerin. Observe the following characters in the different mounts.

Tangentially elongated cork cells in surface view; tiers of ray parenchyma cells in tangential view; thin walled bifurcated fibres with sharp tip upto 500 μ in length (**Citraka**); abundant large simple oval shaped starch grains with eccentric hilum upto 60 μ in size, fragments of septate fibres (**Śuṅṭhī**); uniseriate multicellular trichome, stone cells with broad lumen (**Pippalī**); abundant simple and compound starch grains having 2-7 components round to oval with central hilum appearing like a point up to 28 μ in size (**Pippalīmūla**); parenchymatous tissue with prominent intercellular space; bordered barrel shaped pitted and scalariform vessels up to 350 μ in length and 140 μ in width; elongated sclereids with narrow lumen upto 600 μ in length (**Cavya**); epidermal tissue debris showing papillose and striated cells; fragments of epidermis with papillary outgrowth; fragment of yellowish brown vittae (**Ajamodā**); beaker shaped stone cells; spiral vessels; stone cells associated with parenchyma cells from hypodermis (**Marica**). In addition general characteristics such as abundant perisperm cells from *Marica* and *Pippalī*, vessels members and stone cells are also present.

Thin Layer Chromatography:

Extract 4 g of formulation powder in 75 ml alcohol (25 ml x 3) under reflux on a water bath for 30 min. Filter the extracts, pool the filtrates, concentrate to 10 ml and carry out the thin layer chromatography. Apply 10 μl of extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate* (5: 4) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light (366 nm). It shows major spots at R_f 0.11, 0.23, 0.35 (all pale blue), 0.50, 0.67, 0.85 (all fluorescent blue) 0.58 (dark blue) and 0.76 (blue). Spray the plate with *vanillin-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.19, 0.26, 0.73, 0.86 (all pink), 0.36 (dark grey), 0.47 (yellow), 0.50 (green), 0.60, 0.95 (both violet) and 0.82 (grey) in visible light.

Chemical tests:

Dissolve 1 g of sample in 10 ml of *water* and filter. The filtrate complies with *Tests for Sulphates* (Appendix 5.2.12.5.c) and *Sulphides* (Appendix 5.13.8).

Dissolve 1 g of sample in 10 ml of N *hydrochloric acid* and filter. The filtrate complies with *Test for Magnesium* (Appendix 5.13.3).

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 34 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 3.2 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 11 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 41 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 8 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	4 to 5,	Appendix 3.3

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Arocaka (tastlessness), Āmajaśūla (intestinal colic), Grahaṇī (malabsorption syndrome), Gulma (abdominal dump), Agnimāndya (digestive impairment), Kaphadoṣa (vitiation of Kapha doṣa)

Dose:

3 g with warm water

GHṚTA

General Description:

Ghṛtas are preparations in which the *Ghṛta* is boiled with prescribed liquid [*Svarasa/Kaṣāya* etc.] and fine paste [*Kalka*] of the drugs specified in the formulation composition. Unless specified otherwise *Ghṛta* means *Goghṛta*.

General Method of Preparation:

1. There are usually three essential components in the manufacture of *Ghṛta Kalpanā*.
 - a. *Drava* [Any liquid medium as prescribed in the composition]
 - b. *Kalka* [Fine paste of the specified drugs]
 - c. *Sneha dravya* [Fatty media - *Ghṛta*]
and, occasionally
 - d. *Gandha dravya* [Perfuming agents]
2. Unless otherwise specified in the verse, if *Kalka* is one part by weight, *Ghṛta* should be four parts and the *Drava dravya* should be sixteen parts.
3. There are a few exceptions for the above general rule:
 - a. Where *Drava dravya* is either *Kvātha* or *Svarasa*, the ratio of *Kalka* should be one-sixth and one-eighth respectively to that of *Ghṛta*.
If the *Drava dravya* is either *Kṣīra* or *Dadhi* or *Māṃsa rasa* or *Takra*, the ratio of *Kalka* should be one-eighth to that of *Ghṛta*.
When flowers are advised for use as *Kalka*, it should be one-eighth to that of *Ghṛta*.
 - b. Where the numbers of *Drava-dravya* are four or less than four, the total quantity should be four times to that of *Ghṛta*.
 - c. Where the number of *Drava-dravyas* is more than four, each *drava* should be equal to that of *Ghṛta*.
 - d. If, *Kalka dravya* is not prescribed in a formulation, the drugs specified for the *Drava-dravya* [*Kvātha* or *Svarasa*] should be used for the preparation of *Kalka*.
 - e. Where no *Drava dravya* is prescribed in a formulation, four parts of water should be added to one part of *Ghṛta*.
4. In general, the *Ghṛta* should be subjected to *Mūrcchana* process, followed by addition of increments of *Kalka* and *Drava-dravya* in specified ratio. The contents are to be stirred continuously throughout the process in order to avoid charring.
5. The process of boiling is to be continued till the whole amount of moisture gets evaporated and characteristic features of *Ghṛta* appear.

6. The whole process of *Pāka* should be carried out on a mild to moderate flame.
7. Three stages of *Pāka* are specified for therapeutic purposes.
 - a. *Mṛdu Pāka*: In this stage, the *Kalka* looks waxy and when rolled between fingers, it rolls like lac without sticking. The *Ghṛta* obtained at this stage is used for *Nasya* [Nasal instillation].
 - b. *Madhyama Pāka*: In this stage, the *Kalka* becomes harder and rolls into *Varti*. It burns without crackling sounds when exposed to fire and *phena* [froth] will disappear in *Ghṛta*. The *Ghṛta* obtained at this stage is used for *Pāna* [Internal administration] and *Basti* [Enema].
 - c. *Khara Pāka*: Further heating of the *Ghṛta*, leads to *Khara paka*. *Kalka* becomes brittle when rolled between fingers. The *Ghṛta* obtained at this stage is used only for *Abhyaṅga* [External application].
8. The period of *Pāka* depends upon the nature of liquid media used in the process.
 - a. *Takra* or *Āranala* 5 Nights
 - b. *Svarasa* 3 Nights
 - c. *Kṣīra* 2 Nights
9. *Pātra Pāka*: It is the process by which the *Ghṛta* is augmented or flavored by certain prescribed substances. The powdered drugs are suspended in a vessel containing warm, filtered *Ghṛta*.

The medicated *Ghṛta* will have the odour, colour and taste of the drugs used in the process. If a considerable amount of milk is used in the preparation, the *Ghṛta* will become thick and may solidify in cold seasons.

Ghṛtas are preserved in good quality of glass, steel or polythene containers. These medicated preparations retain the therapeutic efficacy for sixteen months.

SUKUMĀRA GHṚTA

(AFI Part-I, 6:44)

Definition:

Sukumāra ghṛta is a medicated preparation made with the ingredients in the Formulation Composition given below with mūrccita Ghṛta as the basic ingredient.

Formulation Composition:

1	Punarnava (Rakta Punarnavā API)	<i>Boerhaavia diffusa</i>	Pl.	4.800 kg
2	Daśamūla			
	a. Bilva API	<i>Aegle marmelos</i>	Rt./St. Bk.*	480 g
	b. Śyonāka API	<i>Oroxylum indicum</i>	Rt./St. Bk.*	480 g
	c. Gambhāri API	<i>Gmelina arborea</i>	Rt./St. Bk.*	480 g
	d. Pāṭalā API	<i>Stereospermum suaveolens</i>	Rt./St. Bk.*	480 g
	e. Agnimantha API	<i>Clerodendrum phlomidis</i> [<i>Premna integrifolia</i> (Official Substitute)]	Rt./St. Bk.*	480 g
	f. Śālparṇī API	<i>Desmodium gangeticum</i>	Pl.	480 g
	g. Pṛśniparṇī API	<i>Uraria picta</i>	Pl.	480 g
	h. Bṛhatī API	<i>Solanum indicum</i>	Pl.	480 g
	i. Kaṇṭhakārī API	<i>Solanum xanthocarpum</i>	Pl.	480 g
	k. Gokṣura API	<i>Tribulus terrestris</i>	Pl.	480 g
3	Payasyā (Ksīrakākolī API)	<i>Fritillaria roylei</i>	Sub. Rt.	480 g
4	Aśvagandhā API	<i>Withania somnifera</i>	Rt.	480 g
5	Eraṇḍa API	<i>Ricinus communis</i>	Rt.	480 g
6	Śatāvarī API	<i>Asparagus racemosus</i>	Rt. Tr.	480 g
7	Dvidarbhamūla			
	a. Darbha API	<i>Imperata cylindrica</i>	Rt.	480 g
	b. Kuśa API	<i>Desmostachya bipinnata</i>	Rt. Stk.	480 g
8	Śaramūla (Śara API)	<i>Saccharum munja</i> (= <i>S. bengalense</i>)	Rt. Stk.	480 g
9	Kāśamūla (Kāśa API)	<i>Saccharum spontaneum</i>	Rt. Stk.	480 g
10	Ikṣumūla (Ikṣu API)	<i>Saccharum officinarum</i>	Rt. Stk.	480 g
11	Poṭagala API	<i>Typha elephantina</i>	Rt.	480 g
12	Jala API for decoction reduced to	Potable Water	—	49.152 l 6.144 l
13	Guḍa API	Jaggery	—	1.440 kg
14	Eraṇḍa taila (API)	Castor oil	—	768 g
15	Ghṛta (Goghṛta API)	Clarified butter from cow milk	—	1.536 kg
16	Payas (Godugdha API)	Cow milk	—	1.536 kg

* Part actually used in the formulation

17	Kṛṣṇā (Pippalī API)	<i>Piper longum</i>	Fr.	96 g
18	Kṛṣṇāmūla (Pippalī API)	<i>Piper longum</i>	St.	96 g
19	Saindhava (Saindhava Lavaṇa (API))	Rock salt	–	96 g
20	Yaṣṭī API	<i>Glycyrrhiza glabra</i>	Rt.	96 g
21	Madhūka API	<i>Madhuca indica</i>	Fl.	96 g
22	Mṛdvīkā (Drākṣā API)	<i>Vitis vinifera</i>	Dr. Fr.	96 g
23	Yavānī API	<i>Trachyspermum ammi</i>	Fr.	96 g
24	Nāgara (Śuṅṭhī API)	<i>Zingiber officinale</i>	Rz.	96 g

Method of Preparation

- Take all ingredients of pharmacopoeial quality.
- Treat *Ghṛta* to prepare *mūrcchita Ghṛta* (Appendix 6.2.9.2).
- Wash, clean, dry the ingredients numbered 1 to 11 of the formulation composition, powder separately and pass through 355 I.S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for four hours, heat and reduce the volume to one-eighth. Filter through muslin cloth to obtain *kvātha*.
- Strain the *Godugdha* through *muslin cloth*.
- Wash, clean, dry the ingredients numbered 17, 18 and 20 to 24 of the Formulation Composition, powder separately and pass through 180 I.S. sieve (sieve number 85) to obtain fine powder. Roast coarsely powdered *Saindhava Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder and pass through 180 I.S. sieve (sieve number 85) (*Kalka dravya*).
- Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend.
- Take *mūrcchita Ghṛta* in a stainless steel vessel and heat it mildly.
- Add increments of *kalka*, stir thoroughly while adding *Kvātha*, *Guḍa*, *Eraṇḍa taila* and *Godugdha*.
- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating next day. Constantly check the *Kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*Madhyama pāka lakṣaṇa*) to the fingers and at the subsidence of froth (*phenaśānti*) over the *ghṛta*. Expose the *varti* and *ghṛta* to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

A greenish yellow coloured, soft, low-melting medicated fat, unctuous to touch with a pleasant odour and astringent taste

Identification:

Thin layer chromatography:

Extract 5 g of formulation in 75 ml of *n-hexane* (25 ml x3) under reflux on a water bath for 30 min. Pool the extracts, filter and concentrate to 10 ml and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *n-hexane: ethyl acetate* (8.5: 1.5) as mobile phase. After development of plate, allow it to dry in air. Spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating it at 105⁰ for about 10 min. It shows major spots at R_f 0.10, 0.19 (both green), 0.26, 0.38, 0.78 (all blue) and 0.88 (bluish black) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4618 to 1.4622,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.796 to 0.884,	Appendix 3.2
<i>Congealing point:</i>	16 ⁰ to 22 ⁰	Appendix 3.4.2
<i>Saponification value:</i>	213 to 277,	Appendix 3.10
<i>Iodine value:</i>	32 to 34,	Appendix 3.11
<i>Acid value:</i>	Not more than 3,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Vidvibandha; (constipation) Udara; (Urticaria), Gulma; (abdominal lump), Plīhā roga; (Splenic diseases), Vidradhi (abcess); Śopha (oedema); Yonishūla (pain in female genital tract); Arśa (Piles); Vṛddhi (Hydrocoele); Vātavyādhi (diseases due to Vāta Doṣa); Vātarakta (Gout)

Dose:

6 - 12 g per day in divided doses with warm water /milk

GUGGULU

General Description:

Guggulu is an oleoresin (*Niryāsa*) obtained from the plant *Commiphora wightii*. Preparations having the exudates as main effective ingredient are known as *Guggulu*. There are five different varieties of *Guggulu* described in the Ayurvedic texts. However two of the varieties, namely, *Mahiṣākṣa* and *Kanaka Guggulu* are usually preferred for medicinal preparations. *Mahiṣākṣa Guggulu* is dark greenish brown and *Kanaka Guggulu* is yellowish brown in color.

Before using, *Guggulu* is cleaned in the following manner:

1. Sand, stone, plant debris, glass etc. are first removed.
2. It is then broken into small pieces.
3. It is thereafter bundled in a piece of cloth and boiled in *Dola Yantra* containing any one of the following fluids.
 - a. *Gomūtra*,
 - b. *Triphalā kaṣāya*,
 - c. *Nirguṇḍīpatra Svarasa* with *Haridrā Cūrṇa*,
 - d. *Vāsāpatra Kaṣāya*,
 - e. *Vāsāpatra Svarasa* and
 - f. *Dugdha*.

The boiling of *Guggulu* in *Dolā Yantra* is carried on until all the *Guggulu* passes into the fluid through the cloth. By pressing with fingers, much of the fluid that can pass through is taken out. The residue in the bundle is discarded. The fluid is filtered and again boiled till it forms a mass. This mass is dried and then pounded with a pestle in a stone mortar, adding ghee in small quantities till it becomes waxy.

Guggulu cleaned as above, is soft, waxy and brown in color. Characteristics of preparations of *Guggulu* vary depending on the other ingredients added to the preparations.

Guggulu is kept in glass or porcelain jars free from moisture and stored in a cool place. The potency is maintained for two years when prepared with ingredients of plant origin and indefinitely when prepared with metals and minerals.

Note: *Guggulu* formulations can also be prepared in a tablet dosage form, without the use of excipients, but they should comply the general tests for tablets.

SAPTĀṄGA GUGGULU

(AFI Part III, 5:3)

Definition:

Saptāṅga Guggulu vaṭī is a preparation made with the ingredients in the Formulation Composition given below with Guggulu as the basic ingredient.

Formulation Composition:

1	Guggulu API - Śuddha	<i>Commiphora wightii</i>	O. R.	6 g
2	Triphalā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	1 g
	b. Bibhītaka API	<i>Terminalia belerica</i>	P.	1 g
	c. Āmalakī API	<i>Emblica officinalis</i>	P.	1 g
3	Vyoṣa			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	1 g
	b. Marica API	<i>Piper nigrum</i>	Fr.	1 g
	c. Pippalī API	<i>Piper longum</i>	Fr.	1 g
4	Ājya (Goghṛta API)	Clarified butter from cow milk	—	QS

Method of preparation:

- Take all ingredients of the pharmacopoeial quality.
- Treat *Guggulu* to prepare *Guggulu - Śuddha* (Appendix 6.2.8.4).
- Wash, clean, dry the ingredients numbered 2 and 3 of the formulation composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85) to obtain fine powder and mix them all to a homogeneous mixture.
- Crush weighed quantity of *Śuddha-Guggulu*, add fine powder of other mixed ingredients to it and pound well. Add *Ghṛta* to an extent required to facilitate the pounding and continue pounding till a semi-solid uniformly mixed mass of suitable plasticity is obtained.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove and smeared with *Ghṛta* or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Spherical pills, black in colour with agreeable odour and bitter taste

Identification:

Microscopy:

Take about 5 g of the sample, powder it and add *chloroform* (20 ml); stir for 10 min. thoroughly over a water-bath; pour out *chloroform*. Repeat the process thrice adding fresh quantities of *chloroform*; discard *chloroform*. Wash the sediment thoroughly in hot water. Take a few mg of washed material, stain with iodine solution and mount in 50 per cent *glycerin*. Clarify another few mg of the material with *chloral hydrate* and mount in 50 per cent *glycerin*. Take a few mg of washed material on glass slide, moist it with alcoholic solution of *phloroglucinol*, allow to stand until nearly dry and mount in 1-2 drops of concentrated *hydrochloric acid*. Observe the following characters in different mounts.

Fragment of thick-walled epicarp cells in surface view several showing thin septa division, a few fibres crossing each other at right angles (**Harītakī**); simple, short trichomes with a bulbous base, epicarp tissue showing cicatrices (**Bibhitaka**); fragments of polygonal parenchyma cells containing calcium oxalate crystals, abundant crushed parenchymatous large cells showing characteristic corner thickenings (**Āmalakī**); large oval as well as circular starch grains, upto 75 μm in length, with hilum at its broader end, resin containing yellow parenchymatous cells, non-lignified septate fibres several showing dentation on one side owing to pressure exerted by adjacent parenchyma cells, short, spiral xylem vessels (**Śunṭhī**); fragmented tissue from hypodermis, with groups of stone cells interspersed among parenchyma tissue, thick-walled polygonal stone cells from testa (**Marica**); spindle-shaped sclerenchymatous cells with large lumen and pitted walls (**Pippalī**); sclereids of various sizes and shapes are also present in general.

Thin layer chromatography:

Extract 5 g of formulation powder in 75 ml of *n-hexane* under reflux on a water-bath for 30 min. Filter the extract, concentrate to 25 ml and carry out the thin layer chromatography. Apply 10 μl of extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: acetone* (9: 1) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light. It shows major spots at R_f 0.17, 0.35, 0.47 and 0.56 (all fluorescent blue) under 366 nm and at R_f 0.24, 0.32, 0.38, 0.44 (all black) under 254 nm. Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.25 (yellow), 0.30 (red), and 0.38, 0.48 (both brown) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 6 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 2 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 24 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 38 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 13 per cent,	Appendix 2.2.10
<i>pH (1 % aqueous solution):</i>	3.3 to 3.5,	Appendix 3.3

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins:

Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Nāḍīvraṇa (sinus); Duṣṭavraṇa (non-healing ulcer); Bhagandara (fistula-in-ano)

Dose:

3 g daily in divided doses

Anupana:

Triphalā kvātha, Phalatrikādi kvātha, Uṣṇodaka

VARĀDI GUGGULU

(AFI Part III, 5:1)

Definition:

Varādi Guggulu vaṭī is a preparation made with the ingredients in the Formulation Composition given below with Guggulu as the basic ingredient.

Formulation Composition:

1	Varā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	4 g
	b. Bibhītaka API	<i>Terminalia belerica</i>	P.	4 g
	c. Āmalakī API	<i>Emblīca officīnalis</i>	P.	4 g
2	Nimba API	<i>Azadirachta indica</i>	St. Bk.	12 g
3	Arjuna API	<i>Terminalia arjuna</i>	St. Bk.	12 g
4	Aśvattha API	<i>Ficus religīosa</i>	St. Bk.	12 g
5	Khadira Sāra (Khadira API)	<i>Acacia catechu</i>	Wd. extract	12 g
6	Asana API	<i>Pterocarpus marsupium</i>	Ht. Wd.	12 g
7	Vāsaka (Vāsā API)	<i>Adhatoda zeylanica</i>	Rt.	12 g
8	Guggulu API - Śuddha	<i>Commiphora wightii</i>	O. R.	84 g

Method of preparation:

- Take all the ingredients of the pharmacopoeial quality.
- Treat *Guggulu* to prepare *Guggulu -Śuddha* (Appendix 6.2.8.4).
- Wash, clean, dry the ingredients numbered 1 to 7 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder and mix them all to a homogeneous mixture.
- Crush weighed quantity of *Śuddha-Guggulu*, add fine powder of other mixed ingredients to it and pound well. Add *Ghṛta* to an extent required to facilitate the pounding and continue pounding till a semi-solid uniformly mixed mass of suitable plasticity is obtained.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove and smeared with *Ghṛta* or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Spherical pills, black in colour with agreeable odour, taste bitter

Identification:

Microscopy:

Take about 5 g of the sample, powder it and add *chloroform* (20 ml); stir for 10 min thoroughly over a water-bath; pour out *chloroform*. Repeat the process thrice adding fresh quantities of

chloroform; discard *chloroform*. Wash the sediment thoroughly in hot water. Take a few mg of washed material, stain with iodine solution and mount in 50 per cent *glycerin*. Clarify another few mg with *chloral hydrate* and mount in 50 per cent *glycerin*. Take a few mg of washed material on glass slide, moisten it with alcoholic solution of *phloroglucinol*, allow to stand until nearly dry and mount in 1-2 drops of concentrated *hydrochloric acid*. Observe the following characters in different mounts.

Fragment of thick-walled epicarp cells in surface view several showing thin septa division, a few fibres crossing each other at right angles (**Harītakī**); simple, short trichomes with a bulbous base, epicarp tissue showing cicatrices (**Bibhītaka**); fragments of polygonal parenchyma cells containing calcium oxalate crystals, abundant crushed parenchymatous large cells showing characteristic corner thickenings (**Āmalakī**); groups of parenchymatous cells containing isolated prisms of calcium oxalate crystals, large groups of well developed long thick walled lignified fibres associated with phloem elements (**Nimba**); idioblasts upto 600µm or more in size, containing abundant prisms and rhombs of calcium oxalate crystals, crystal fibres associated with phloem fibres, cells containing rosette crystals of calcium oxalate, parenchyma cells from cortical tissue containing rosette crystals (**Arjuna**); groups of fibres associated with phloem tissue, but non lignified, thin walled along with thick walled phloem parenchyma containing prismatic crystals of calcium oxalate, crystal fibres from phloem, with prismatic crystals of calcium oxalate in each cell (**Aśvattha**); xylem vessels filled with tyloses, associated with tracheids and fibre tracheids with thick wall, simple pits and narrow lumen (**Asana**); tissue from cortex with group of rectangular stone cells showing distinct pits and pit canals, vessel group with simple pits and without tyloses, cortical tissue with several cells showing yellow contents (**Vasaka**); abundant dark brownish tissues from rhytidoma, associated with stone cells, abundant starch grains, isolated and compound within parenchymatous cells, abundant loose crystals of calcium oxalate in the form of prisms, rhombs and rosettes are also present in general.

Thin layer chromatography:

Extract 5 g of formulation powder in 75 ml of *n-hexane* under reflux on a water-bath for 30 min. Filter the extract, concentrate to 25 ml and carry out the thin layer chromatography. Apply 10 µl of extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: acetone* (9: 1) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light. It shows major spots at R_f 0.17, 0.35, 0.41 and 0.56 (all fluorescent blue) under 366 nm and at R_f 0.32, 0.37 (both black) under 254 nm. Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.16, 0.38, 0.48 (all brown) and 0.30 (red) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 12 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 4.0 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 18 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 29 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 14 per cent,	Appendix 2.2.10
<i>pH (1 % aqueous solution):</i>	3.7 to 3.9,	Appendix 3.3

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Upadaṃśa (syphilis/soft chancre); Rakatadoṣa (blood disorders); Duṣṭavraṇa (non-healing ulcer)

Dose:

2 g daily in divided doses

Anupana:

Triphalā kvātha, Mañjiṣṭhādi kvātha, Uṣṇodaka

VIḌAṄGĀDI GUGGULU

(AFI Part III, 5:2)

Definition:

Viḍaṅgādi Guggulu vaṭī is a preparation made with the ingredients in the Formulation Composition given below with Guggulu as the basic ingredient.

Formulation Composition:

1	Viḍaṅga API	<i>Embelia ribes</i>	Fr.	60 g
2	Triphalā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	20 g
	b. Bibhītaka API	<i>Terminalia belerica</i>	P.	20 g
	c. Āmalakī API	<i>Emblica officinalis</i>	P.	20 g
3	Vyoṣa			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	20 g
	b. Marica API	<i>Piper nigrum</i>	Fr.	20 g
	c. Pippalī API	<i>Piper longum</i>	Fr.	20 g
4	Guggulu API - Śuddha	<i>Commiphora wightii</i>	O. R.	180 g
5	Sarpi (Goghṛta API)	Clarified butter from cow milk	—	QS

Method of preparation:

- Take all the ingredients of the pharmacopoeial quality.
- Treat *Guggulu* to prepare *Guggulu - Śuddha* (Appendix 6.2.8.4).
- Wash, clean, dry the ingredients numbered 1 to 3 of the formulation composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder and mix them all to a homogeneous mixture.
- Crush weighed quantity of *Śuddha-Guggulu*, add fine powder of other mixed ingredients to it and pound well. Add *Ghṛta* to an extent required to facilitate the pounding and continue pounding till a semi-solid uniformly mixed mass of suitable plasticity is obtained.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove and smeared with *Ghṛta* or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Spherical pills, black in colour with agreeable odour, bitter taste

Identification:

Microscopy:

Take about 5 g of the sample, powder it and add *chloroform* (20 ml); stir for 10 min. thoroughly over a water-bath; pour out *chloroform*. Repeat the process thrice adding fresh quantities of *chloroform*; discard *chloroform*. Wash the sediment thoroughly in hot water. Take a few mg of washed material, stain with iodine solution and mount in 50 per cent *glycerin*. Clarify another few mg of the washed material with *chloral hydrate* and mount in 50 per cent *glycerin*. Take a few mg of washed material on glass slide, moist it with alcoholic solution of *phloroglucinol*, allow to stand until nearly dry and mount in 1-2 drops of concentrated *hydrochloric acid*. Observe the following characters in different mounts.

Fragment of thick-walled epicarp cells in surface view several showing thin septa division, a few fibres crossing each other at right angles (**Harītakī**); simple, short trichomes with a bulbous base, epicarp tissue showing cicatrices (**Bibhītaka**); fragments of polygonal parenchyma cells containing calcium oxalate crystals, abundant crushed parenchymatous large cells showing characteristic corner thickenings (**Āmalakī**); large oval as well as circular starch grains, upto 75 μm in length, with hilum at its broader end, resin containing yellow parenchymatous cells, non-lignified septate fibres several showing dentation on one side owing to pressure exerted by adjacent parenchyma cells, short, spiral xylem vessels (**Śunthī**); fragmented tissue from hypodermis, with groups of stone cells interspersed among parenchyma tissue, thick-walled polygonal stone cells from testa (**Marica**); spindle-shaped sclerenchymatous cells with large lumen and pitted walls (**Pippalī**); stone cell layers with prominent pits and narrow lumen from testa, lignified sclereids with broad lumen and pitted walls from testa layers, associated with parenchymatous cells containing prisms of calcium oxalate (**Vidanga**); sclereids of various sizes and shapes are also seen in general.

Thin layer chromatography:

Extract 5 g of formulation powder in 75 ml of *n-hexane* under reflux on a water-bath for 30 min. Filter the extract, concentrate to 25 ml and carry out the thin layer chromatography. Apply 10 μl of extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: acetone* (9: 1) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light. It shows major spots at R_f 0.11, 0.27, 0.33 and 0.48 (all fluorescent blue) under 366 nm and at R_f 0.17, 0.26, 0.32, 0.38 (all black) under 254 nm. Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105° for about 10 min. It shows major spots at R_f 0.23 (red), 0.33 (black) and 0.43 (brown) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 6 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 2 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 23 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 30 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 13 per cent,	Appendix 2.2.10
<i>pH (1 % aqueous solution):</i>	3.3 to 3.5,	Appendix 3.3

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Duṣṭa vṛāṇa (non-healing ulcer); Apacī (chronic lymphadenopathy/scrofula); Meha (excessive flow of urine); Kuṣṭha (diseases of skin); Nāḍīvṛāṇa (sinus)

Dose:

12 g daily in divided doses

Anupana:

Triphalā kvātha, Śigru kvātha, Madhu, Uṣṇodaka

TAILA

General Description:

Tailas are preparations in which *Taila* is boiled with prescribed liquid media [*Svarasa* / *Kvātha* Etc.] and a fine paste [*Kalka*] of the drugs specified in the formulation composition. Unless specified otherwise *Taila* means *Tila Taila*.

General Method of Preparation:

1. The *Taila* preferably should be fresh.
2. There are usually three essential components in the manufacture of *Taila Kalpanā*.
 - a. *Drava* [Any liquid medium as prescribed in the composition]
 - b. *Kalka* [Fine paste of the specified drug]
 - c. *Sneha dravya* [*Taila*]
and, occasionally,
 - d. *Gandha dravya* [Perfuming agents]
3. Unless otherwise specified in the verse, if *Kalka* is one part by weight, *Taila* should be four parts and the *Drava dravya* should be sixteen parts.
4. There are a few exceptions for the above general rule:
 - a. Where *Drava dravya* is either *Kvātha* or *Svarasa*, the ratio of *Kalka* should be one-sixth and one-eighth respectively to that of *Taila*.
If the *Drava dravya* is either *Kṣīra* or *Dadhi* or *Māṃsa rasa* or *Takra*, the ratio of *Kalka* should be one-eighth to that of *Taila*.
When flowers are advised for use as *Kalka*, it should be one-eighth to that of *Taila*.
 - b. Where the numbers of *Drava dravyas* are four or less than four, the total quantity should be four times to that of *Taila*.
 - c. Where the number of *Drava dravyas* is more than four, each *drava* should be equal to that of *Taila*.
 - d. If, *Kalka dravya* is not prescribed in a formulation, the drugs specified for the *Drava dravya* [*Kvātha* or *Svarasa*] should be used for the preparation of *Kalka*.
 - e. Where no *Drava dravya* is prescribed in a formulation, four parts of water should be added to one part of *Taila*.
5. In general, the *Taila* should be subjected to *Mūrccana* process, followed by addition of increments of *Kalka* and *Drava dravya* in specified ratio. The contents are to be stirred continuously throughout the process in order to avoid charring.

6. The process of boiling is to be continued till the whole amount of moisture gets evaporated and characteristic features of *Taila* appears.
7. The whole process of *Pāka* should be carried out on a mild to moderate flame.
8. Three stages of *Pāka* are specified for therapeutic purposes.
 - a. *Mṛdu Pāka*: In this stage, the *Kalka* looks waxy and when rolled between fingers, it rolls like lac without sticking. The *Taila* obtained at this stage is used for *Nasya* [Nasal instillation].
 - b. *Madhyama Pāka*: In this stage, the *Kalka* becomes harder and rolls in to *Varti*. It burns without crackling sounds when exposed to fire and *phena* [Froth] will appear over the *Taila*. *Taila* obtained at this stage is used for *Pāna* [Internal administration] and *Basti* [Enema].
 - c. *Khara Pāka*: Further heating of the *Taila*, leads to *Khara Pāka*. *Kalka* becomes brittle when rolled in between fingers. The *Taila* obtained at this stage is used only for *Abhyaṅga* [External application].
9. The period of *Pāka* depends upon the nature of liquid media used in the process.
 - a. *Takra* or *Āranala* 5 Nights
 - b. *Svarasa* 3 Nights
 - c. *Kṣīra* 2 Nights
10. *Pātra pāka*: It is the process by which the *Taila* is augmented or flavored by certain prescribed substances. The powdered drugs are suspended in a vessel containing warm, filtered *Taila*.

The medicated *Taila* will have the odour, colour and taste of the drugs used in the process. If a considerable amount of milk is used in the preparation, the *Taila* will become thick and may solidify in cold seasons.

Tailas are preserved in good quality of glass, steel or polythene containers. These medicated preparations retain the therapeutic efficacy for sixteen months.

ANU TAILA

(AFI Part I, 8:1)

Definition:

Anu Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Tila taila as the basic ingredient.

Formulation Composition:

1	Jīvantī API	<i>Leptadenia reticulata</i>	Rt.	28 g
2	Jala (Hrīvera API)	<i>Coleus vettiveroides</i>	Rt.	28 g
3	Devadāru API	<i>Cedrus deodara</i>	Ht. Wd.	28 g
4	Jalada (Mustā API)	<i>Cyperus rotundus</i>	Rz.	28 g
5	Tvak API	<i>Cinnamomum zeylanicum</i>	St. Bk.	28 g
6	Sevya (Uśīra API)	<i>Vetiveria zizanioides</i>	Rt.	28 g
7	Gopī (Śveta Sārivā API)	<i>Hemidesmus indicus</i>	Rt.	28 g
8	Hima (Śveta Candana API)	<i>Santalum album</i>	Ht. Wd.	28 g
9	Dārvī (Dāruharidrā API)	<i>Berberis aristata</i>	St.	28 g
10	Madhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	28 g
11	Plava (Kaivarta Mustā (API))	<i>Cyperus scariosus</i>	Rz.	28 g
12	Agaru API	<i>Aquilaria agallocha</i>	Ht. Wd.	28 g
13	Varī (Śatāvārī API)	<i>Asparagus racemosus</i>	Rt.	28 g
14	Puṇḍrāhva (Prapaṇḍarīka API)	<i>Nelumbo nucifera</i>	Fl.	28 g
15	Bilva API	<i>Aegle marmelos</i>	Rt.* / St. Bk.	28 g
16	Utpala API	<i>Nymphaea stellata</i>	Fl.	28 g
17	Dhāvanīdvaya			
	a. Bṛhatī API	<i>Solanum indicum</i>	Pl.	28 g
	b. Kaṇṭakārī API	<i>Solanum surratense</i> (= <i>S. xanthocarpum</i>)	Pl.	28 g
18	Surabhi (Rāsnā API)	<i>Pluchea lancolata</i> [<i>Alpinia officinarum</i> (Official Substitute)]	Rt.* / Lf.	28 g
19	Sthirādvaya			
	a. Śālaparṇī API	<i>Desmodium gangeticum</i>	Pl.	28 g
	b. Prṣniparṇī API	<i>Uraria picta</i>	Pl.	28 g
20	Kṛimihara (Viḍaṅga API)	<i>Embelia ribes</i>	Fr.	28 g
21	Patra (Tvakpatra API)	<i>Cinnamomum tamala</i>	Lf.	28 g
22	Truṭi (Sūkṣmailā API)	<i>Elettaria cardamomum</i>	Sd.	28 g
23	Reṇukā API	<i>Vitex negundo</i>	Sd.	28 g
24	Kamalakiñjalka (Kamala API)	<i>Nelumbo nucifera</i>	Adr.	28 g
25	Jala API for decoction reduced to	Potable Water	—	76.800 l 7.680 l

* Actual part used in the formulation

26	Drugs 1-24 for Kalka			192 g
27	Taila (Tila Taila API)	Sesame oil	—	768 g
28	Ājadugdha (Ājādugdha API)	Goat milk	—	768 ml

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrcchita Tila taila* (Appendix 6.2.9.4).
- Wash, clean, dry the ingredients numbered 1 to 24 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-tenth. Filter through *muslin cloth* to obtain *kvātha*.
- Divide the *kvātha* into 10 equal parts and store separately in air-tight containers protected from light and moisture.
- Wash, clean, dry the drugs under ingredient number 26 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder. Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrcchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding one part of *kvātha*.
- Heat with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop the heating when the *kalka* gets separated at the bottom of the vessel in the form of loose paste (*mṛdu pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Allow to stand overnight.
- Repeat the process for nine more times each day adding one part of *kvātha*. Along with the 10th part of *kvātha*, add *Ājādugdha* in last *pāka*.
- On the last day, constantly check the *kalka* by rolling between the fingers. Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, reddish brown in colour with characteristic odour

Identification:

Thin Layer Chromatography:

Shake 5 ml of formulation with 10 ml of *methanol*. Allow the mixture to stand till the two layers separate. Separate the methanolic layer, filter and carry out the thin layer chromatography. Apply 5 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: chloroform: methanol* (8: 0.5: 0.5: 0.2) as mobile phase. Spray the plate with 10 per cent

methanolic sulphuric acid followed by heating at 105⁰ for about 10 min. It shows spots at R_f 0.15, 0.48, 0.60, 0.73 and 0.81 (all brown).

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4646 to 1.4659,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.7706 to 0.7709,	Appendix 3.2
<i>Saponification value:</i>	188 to 200,	Appendix 3.10
<i>Iodine value:</i>	88 to 106,	Appendix 3.11
<i>Acid value:</i>	2 to 3,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Tvakrauṣya (dryness of skin); Palita (greying of hair); Ūrdhvajatrugata roga (diseases of head and neck); Skandha śuṣkatā (emaciation of shoulder); Grīvā śuṣkatā (wasting in cervical region); Vakṣa śuṣkatā (emaciation of chest muscles)

Dose:

5 to 10 drops per nostril as Nasya

APĀMĀRGA KṢĀRA TAILA

(AFI Part II, 8:1)

Definition:

Apāmārga Kṣāra Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with Tila taila as the basic ingredient.

Formulation Composition:

1	Mārga Kṣāra (Apāmārga Kṣāra (API))	<i>Achyranthes aspera</i>	Water soluble ash of Pl.	16 parts
2	Jala API	Potable Water	—	96 parts
3	Apāmārga API	<i>Achyranthes aspera</i>	Pl.	6 parts
4	Taila (Tila Taila API)	Sesame oil	—	24 parts

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccchita Tila taila* (Appendix 6.2.9.4).
- Add 6 parts of water to 1 part of *Kṣāra* (Appendix 6.2.3), and dissolve completely (*Kṣāra jala*).
- Wash, clean, dry the ingredient number 4 of the Formulation Composition, powder and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).
- Transfer the powder to wet grinder and grind with sufficient quantity of water to prepare homogenous blend (*kalka*).
- Take *mūrccchita Tila Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*. Stir thoroughly while adding the *Kṣāra jala*.
- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, pink in colour

Identification:

Thin layer chromatography:

Shake 0.5 g of formulation with 10 ml of methanol and keep the mixture over night with occasional shakings. Allow the two layers to separate and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *dichloromethane: toluene: methanol* (7: 2: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and examine under ultraviolet light (366 nm). It shows major spots at R_f 0.45 (light purple), 0.60 (light blue), 0.65 (fluorescent blue) and 0.80 (cream).

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4600 to 1.4805,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.8948 to 0.9236,	Appendix 3.2
<i>Saponification value:</i>	182 to 202,	Appendix 3.10
<i>Iodine value:</i>	103 to 106,	Appendix 3.11
<i>Acid value:</i>	Not more than 1.0,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 3,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Karṇanāda (Tinnitus); Bādhirya (Deafness)

Dose:

Ear drops - 2 to 5 drops in each ear once or twice a day

ARIMEDĀDI TAILA

(AFI Part I, 8:2)

Definition:

Arimedādi Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Tila taila as the basic ingredient.

Formulation Composition:

1	Khadira API	<i>Acacia catechu</i>	Wd.	9.600 kg
2	Arimedavalka (Arimeda API)	<i>Acacia leucophloea</i>	St. Bk.	4.800 kg
3	Jala API for decoction reduced to	Potable Water	—	49.152 l 12.288 l
4	Taila (Tila Taila API)	Sesame oil	—	3.072 kg
5	Sevya (Uśīra API)	<i>Vetiveria zizanioides</i>	Rt.	12 g
6	Ambu (Hrīvera API)	<i>Coleus vettiveroides</i>	Rt.	12 g
7	Pattaṅga API	<i>Caesalpinia sappan</i>	Ht. Wd.	12 g
8	Gairika API- Śuddha	Red ochre	—	12 g
9	Candanadvaya			
	a. Śveta candana API	<i>Santalum album</i>	Ht. Wd.	12 g
	b. Rakta candana API	<i>Pterocarpus santalinus</i>	Ht. Wd.	12 g
10	Rodhra (Lodhra API)	<i>Symplocos racemosa</i>	St. Bk.	12 g
11	Puṇḍrāhva (Prapaṇḍarīka API)	<i>Nelumbo nucifera</i>	Fl.	12 g
12	Yaṣṭyāhva (Yaśṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	12 g
13	Lākṣā (API)	<i>Lacca laccifera</i>	Secretion of Lac insect	12 g
14	Añjanadvaya			
	a. Rasāñjana (API)	<i>Berberis aristata</i>	St. Ext.	12 g
	b. Sauvīrāñjana (API) - Śuddha	<i>Lead sulphide</i>	—	12 g
15	Dhātakī API	<i>Woodfordia fruticosa</i>	Fl.	12 g
16	Kaṭphala API	<i>Myrica esculenta</i>	St. Bk.	12 g
17	Dviniśā			
	a. Haridrā API	<i>Curcuma longa</i>	Rz.	12 g
	b. Dāruharidrā API	<i>Berberis aristata</i>	St.	12 g
18	Triphlā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	12 g
	b. Bibhītaka API	<i>Terminalia belerica</i>	P.	12 g
	c. Āmalakī API	<i>Emblica officinalis</i>	P.	12 g
19	Caturjāta			
	a. Tvak API	<i>Cinnamomum zeylanicum</i>	St. Bk.	12 g
	b. Sūkṣmailā API	<i>Elettaria cardamomum</i>	Sd.	12 g
	c. Tvakpatra API	<i>Cinnamomum tamala</i>	Lf.	12 g
	d. Nāgakeśara API	<i>Mesua ferra</i>	Stmn.	12 g

20	Joṅgaka (Agaru API)	<i>Aquilaria agallocha</i>	Ht. Wd.	12 g
21	Musta (Mustā API)	<i>Cyperus rotundus</i>	Rz.	12 g
22	Mañjiṣṭhā API	<i>Rubia cordifolia</i>	St.	12 g
23	Nyagrodhapraroha (Nyagrodha API)	<i>Ficus benghalensis</i>	A. R.	12 g
24	Māṃsī (Jaṭāmāṃsī API)	<i>Nardostachys jatamansi</i>	Rz./ Rt.	12 g
25	Yavāsaka API	<i>Alhagi pseudalhagi</i>	Pl.	12 g
26	Padmaka API	<i>Prunus cerasoides</i>	Ht. Wd.	12 g
27	Aileya (Elavāluka API)	<i>Prunus avium</i>	St. Bk.	12 g
28	Samaṅgā (Lajjālu API)	<i>Mimosa pudica</i>	Pl.	12 g
29	Jātīpatrikā (Jātīphalā API)	<i>Myristica fragrans</i>	Ar.	48 g
30	Jātīphala API	<i>Myristica fragrans</i>	Sd.	48 g
31	Lavaṅga API	<i>Syzygium aromaticum</i>	Fl. Bd.	48 g
32	Kaṅkolikā (Kaṅkola API)	<i>Piper cubeba</i>	Fr.	48 g
33	Karpūra API	<i>Cinnamomum camphora</i>	Subl. Ext. of A. Pt.	192 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccchita Tila taila* (Appendix 6.2.9.4).
- Treat *Gairika* to prepare *Gairika - Śuddha* (Appendix 6.2.8.2) and *Sauvirāñjana* to prepare *Sauvirāñjana - Śuddha* (Appendix 6.2.8.18).
- Wash, clean, dry the ingredients numbered 1 and 2 of the Formulation Composition, powder separately and pass through 355 µm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, dry the ingredients numbered 5 to 7, 9 to 13 and 15 to 33 of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85) to obtain fine powder.
- Transfer the powdered ingredients to wet grinder alongwith *Gairika - Śuddha, Rasāñjana* and *Sauvirāñjana - Śuddha* and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrccchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*Madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Powder *Karpūra*, pass through 355 µm I. S. sieve, add to the oil and mix homogeneously.

- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, yellowish red in colour with a pleasant odour

Identification:

Thin Layer Chromatography:

a) Shake 1 ml of formulation with 10 ml of *methanol* for 10 min and keep the mixture for 12 h at 37°. Filter and carry out thin layer chromatography. Apply 3 µl of the extract on a TLC plate. Develop the plate up to a distance of 8 cm using *toluene: ethyl acetate* (9: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with 10 per cent *methanolic sulphuric acid reagent* followed by heating at 105° for about 10 min. The plate shows major spots at R_f 0.16, 0.22 (both faint brown), 0.30, 0.60 (both yellow) 0.35 (faint purple), 0.42 (light purple), 0.48 (light yellow), 0.74 (light brown), 0.86 (brown) in visible light.

b) *Detection of Eugenol :*

Extract 0.5 ml of formulation with 10 ml of *Chloroform* for 15 min and filter.

Dissolve 1 mg of *eugenol* in 10 ml of *chloroform*.

Apply 5 µl each of the sample extract and *eugenol* solution on TLC plate and develop the plate to 8 cm using *toluene: chloroform: acetone* (3: 5: 0.2) as mobile phase. After development, allow the plate to dry in air, spray *anisaldehyde reagent* followed by heating at 105° for about 10 min. The plate shows a spot at R_f 0.62 (brown corresponding to *eugenol*) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40°:</i>	1.4646 to 1.4665,	Appendix 3.1
<i>Specific gravity at 40°:</i>	0.8123 g to 0.8388,	Appendix 3.2
<i>Saponification value:</i>	180 to 190,	Appendix 3.10
<i>Iodine value:</i>	98 to 102,	Appendix 3.11
<i>Acid value:</i>	Not more than 1,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Mukha roga (diseases of mouth), Danta roga (disease of tooth), Nāsā roga (diseases of nose)

Dose:

Nasya: 5 to 10 drops per nostril;

External use: for Kavalagraha (gargling), local application in buccal cavity and Śirodhārā

ASANABILVĀDI TAILA

(AFI Part I, 8:3)

Definition:

Asanabilvādi Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūr̥chita Tila taila as the basic ingredient.

Formulation Composition:

1	Asana API	<i>Pterocarpus marsupium</i>	Ht. Wd.	76 g
2	Bilva API	<i>Aegle marmelos</i>	Rt. * /St. Bk.	76 g
3	Balā API	<i>Sida cordifolia</i>	Pl.	76 g
4	Amṛta (Guḍūcī API)	<i>Tinospora cordifolia</i>	St.	76 g
5	Jala API for decoction reduced to	Potable Water	—	12.288 l 3.072 l
6	Madhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	38.4 g
7	Nāgaraka (Śuṅṭhī API)	<i>Zingiber officinale</i>	Rz.	38.4 g
8	Triphlā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	38.4 g
	b. Bibhītaka API	<i>Terminalia bellirica</i>	P.	38.4 g
	c. Āmalakī API	<i>Emblica officinalis</i>	P.	38.4 g
9	Payas (Godugdha API)	Cow milk	—	768 ml
10	Taila (Tila Taila API)	Sesame oil	—	768 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūr̥chita Tila taila* (Appendix 6.2.9.4).
- Wash, clean, dry the ingredients numbered 1 to 4 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, dry the ingredients numbered 6 to 8 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder.
- Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūr̥chita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.

* Part actually used in the formulation

- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *Godugdha* and continue the process of heating. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*Madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, brown in colour with the characteristic odour of sesame oil

Identification:

Thin Layer Chromatography:

Shake 2 g of formulation with 10 ml of *toluene* for 10 min and keep the mixture for 12 h at 37⁰. Filter and dilute the extract in *toluene* in the proportion of 1: 4. Apply 5 µl of the diluted extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: methanol: glacial acetic acid* (6: 0.5: 0.2: 0.1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.41, 0.47, 0.58, 0.67 and 0.77 (all brown) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4646 to 1.4665,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.9207 to 0.9311,	Appendix 3.2
<i>Saponification value:</i>	180 to 190,	Appendix 3.10
<i>Iodine value:</i>	106 to 109,	Appendix 3.11
<i>Acid value:</i>	Not more than 1,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Nayana roga (diseases of the eye); Karṇa roga (diseases of the ear); Śīroroga (diseases of head)

Dose:

External use - for Abhyaṅga (massage on whole body)

Nasya - 2 to 4 drops per nostril once or twice a day

Eye drops - 2 to 4 drops once or twice a day

Ear drops - 5 to 10 drops once or twice a day

BALĀ TAILA

(AFI Part I, 8:33)

Definition:

Balā Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Tila taila as the basic ingredient.

Formulation Composition:

1	Balā (API)	<i>Sida cordifolia</i>	Rt.	4.800 kg
2	Chinnaruhā (Guḍūcī API)	<i>Tinospora cordifolia</i>	St.	1.200 kg
3	Rāsnā API	<i>Pluchea lancolata</i> [<i>Alpinia officinarum</i> (Official Substitute)]	Rz.	0.600 kg
4	Jala API for decoction reduced to	Potable Water	—	30.720 l 3.072 l
5	Dadhimastu (Godadhi (API))	Whey from curd of cow milk	—	3.072 l
6	Ikṣu Niryāsa (Ikṣu API)	<i>Saccharum officinarum</i>	St. juice	3.072 l
7	Śukta (Kāñjika (API))	Sour gruel	—	3.072 l
8	Taila (Tila Taila API)	Sesame oil	—	3.072 kg
9	Ājapaya (Ajādugdha (API))	Goat milk	—	1.536 l
10	Śaṭhī (Śaṭī API)	<i>Hedychium spicatum</i>	Rz.	48 g
11	Sarala API	<i>Pinus roxburghii</i>	Ht. Wd.	48 g
12	Dāru (Devadāru API)	<i>Cedrus deodara</i>	Ht. Wd.	48 g
13	Elā (Sūkṣmailā API)	<i>Elettaria cardamomum</i>	Sd.	48 g
14	Mañjiṣṭhā API	<i>Rubia cordifolia</i>	Rt.	48 g
15	Aguru (Agaru API)	<i>Aquilaria agallocha</i>	Ht. Wd.	48 g
16	Candana (Rakta candana API)	<i>Pterocarpus santalinus</i>	Ht. Wd.	48 g
17	Padmaka API	<i>Prunus cerasoides</i>	Ht. Wd.	48 g
18	Atibalā API	<i>Abutilon indicum</i>	Rt.	48 g
19	Mustā API	<i>Cyperus rotundus</i>	Rz.	48 g
20	Śūrparṇīdvaya			
	a. Mudgaparṇī API	<i>Phaseolus trilobus</i>	Pl.	48 g
	b. Māṣaparṇī API	<i>Teramnus labialis</i>	Pl.	48 g
21	Hareṇu (Reṇukā (API))	<i>Vitex agnus-castus</i> [<i>Vitex negundo</i> (Official Substitute)]	Fr.	48 g
22	Yaṣṭyāhva (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	48 g
23	Surasa (Tulasī API)	<i>Ocimum sanctum</i>	Pl.	48 g
24	Vyāghranakha API	<i>Capparis sepiaria</i>	Fr.	48 g

25	R̥ṣabhaka (API)	<i>Microstylis wallichii</i> [<i>Pueraria tuberosa</i> (Official Substitute)]	Rt. Tr.	48 g
26	Jīvaka API	<i>Microstylis wallichii</i> [<i>Pueraria tuberosa</i> (Official Substitute)]	Rt .Tr.	48 g
27	Palāśa rasa (Palāśa (API))	<i>Butea monosperma</i>	Exd.	48 g
28	Nīlikā (Nīlī API)	<i>Indigofera tinctoria</i>	Pl.	48 g
29	Jātikośa (Jātipatrī (API))	<i>Myristica fragrans</i>	Ar.	48 g
30	Spr̥kkā API	<i>Anisomeles malabarica</i>	Pl.	48 g
31	Kuñkuma API	<i>Crocus sativus</i>	Stmn./Stg.	48 g
32	Śaileya API	<i>Parmelia perlata</i>	Pl.	48 g
33	Jātikā (Jātīphala API)	<i>Myristica fragrans</i>	Sd.	48 g
34	Kaṭphala API	<i>Myrica esculenta</i> (= <i>M. nagi</i>)	St. Bk.	48 g
35	Ambu (Hrīvera API)	<i>Coleus vettiveroides</i>	Rt.	48 g
36	Tvak API	<i>Cinnamomum zeylanicum</i>	St. Bk.	48 g
37	Kunduruka (Kundurū API)	<i>Boswellia serrata</i>	Exd.	48 g
38	Karpūra API	<i>Cinnamomum camphora</i>	Subl. Ext. of A. Pt.	48 g
39	Turuṣka API	<i>Liquidambar orientalis</i> [<i>Altingia excelsa</i> (Official Substitute)]	Exd.	48 g
40	Śrīnivāsaka (Sarala API)	<i>Pinus roxburghii</i>	Exd.	48 g
41	Lavaṅga API	<i>Syzygium aromaticum</i>	Fr. Bd.	48 g
42	Nakha (Vyāghranakha API)	<i>Capparis sepiaria</i>	Fr.	48 g
43	Kaṅkola API	<i>Piper cubeba</i>	Fr.	48 g
44	Kuṣṭha API	<i>Saussurea lappa</i>	Rt.	48 g
45	Māṃsī (Jāṭāmāṃsī API)	<i>Nardostachys jatamansi</i>	Rz.	48 g
46	Priyaṅgu API	<i>Callicarpa macrophylla</i>	Fl.	48 g
47	Sthaṇḍeya API	<i>Taxus baccata</i>	Lf.	48 g
48	Tagara API	<i>Valeriana wallichii</i>	Rz.	48 g
49	Dhyāma (Rohiṣa API)	<i>Cymbopogon martini</i>	Pl.	48 g
50	Vacā API	<i>Acorus calamus</i>	Rz.	48 g
51	Madanaka (Madana API)	<i>Randia dumetorum</i>	Fr.	48 g
52	Plava (Kaivarta Mustā (API))	<i>Cyperus scariosus</i>	Rz.	48 g
53	Nāgakesara (Nāgakeśara API)	<i>Mesua ferrea</i>	Stmn.	48 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccchita Tila taila* (Appendix 6.2.9.4).
- A day prior to the *sneha pāka*, initiate to prepare *Mastu* (Appendix 6.2.5).

- Wash, clean, dry the ingredients numbered 1 to 3 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-tenth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash and clean the cut pieces of *Ikṣu*, and squeeze the juice and strain through *muslin cloth* to obtain *Ikṣu svarasa*.
- Strain the *Ajādugdha* through *muslin cloth*.
- Wash, dry the ingredients numbered 10 to 53 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder. Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrcchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha* and *svarasa*.
- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *tha Ajādugdha*, *Mastu* and *Kāñjika* and continue the process of heating intermittently over a period of five days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, light golden yellow in color with a pleasant odour

Identification:

Thin layer chromatography:

Shake 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 μl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.32, 0.43, 0.53 and 0.62 in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.451 to 1.468,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.915 to 0.940,	Appendix 3.2
<i>Saponification value:</i>	188 to 200,	Appendix 3.10
<i>Iodine value:</i>	80 to 100,	Appendix 3.11
<i>Acid value:</i>	3 to 6,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Chardi (emesis); Gulma (abdominal lump); Kāsa (cough); Śvāsa (Asthma); Jvara (fever); Mūrcchā (syncope); Kṣata (wound); Kṣaya (Pthisis); Apasmāra (Epilepsy); Vātavyādhi (diseases due to Vāta doṣa); Plīhā (splenic disease); Śoṣa (cachexia)

Dose:

5-10 ml per day in divided doses; for external use as advised by the physician

BALĀHAṬHĀDI TAILA

(AFI Part I, 8:37)

Definition:

Balāhaṭhādi Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccita Tila taila as the basic ingredient.

Formulation Composition:

1	Balā (API)	<i>Sida cordifolia</i>	Rt.	153.6 g
2	Haṭhā (Āmalakī API)	<i>Emblica officinalis</i>	Pl.	153.6 g
3	Amṛtā (Guḍūcī API)	<i>Tinospora cordifolia</i>	St.	153.6 g
4	Mudga API	<i>Phaseolus radiatus</i>	Sd.	153.6 g
5	Māṣa API	<i>Phaseolus mungo.</i>	Sd.	153.6 g
6	Jala API for decoction reduced to	Potable Water	—	12.288 l 3.072 l
7	Tilodbhava (Tila Taila API)	Sesame oil	—	768 g
8	Candana (Rakta Candana API)	<i>Pterocarpus santalinus</i>	Ht. Wd.	42.66 g
9	Āmaya (Kuṣṭha API)	<i>Saussurea lappa</i>	Rt.	42.66 g
10	Yaṣṭi (Yaṣṭī API)	<i>Glycyrriza glabra</i>	Rt.	42.66 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccita Tila taila* (Appendix 6.2.9.4).
- Wash, clean, dry the ingredients numbered 1 to 5 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash and pulverize the ingredients numbered 1 to 3 to coarse powder. Weigh *Mudga* and *Māṣa* in the required quantity and add to the prior pulverized ingredients.
- Wash, clean, dry the ingredients numbered 8 to 10 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).
- Transfer the *kalka dravya* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrccita Tila Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.

- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, brownish yellow in colour with a pleasant odour

Identification:

Thin layer chromatography:

Shake 2 g of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.16, 0.22, 0.27, 0.32, 0.43, 0.53 and 0.75 (all pink changing to purple) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.455 to 1.462,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.929 to 0.936,	Appendix 3.2
<i>Saponification value:</i>	175 to 190,	Appendix 3.10
<i>Iodine value:</i>	84 to 94,	Appendix 3.11
<i>Acid value:</i>	3 to 5,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Śīrorujā (headache)

Dose:

External application for Śīro-abhyaṅga

BHRṄGARĀJA TAILA

(AFI Part I, 8:42)

Definition:

Bhrṅgarāja Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccḥita Tila taila as the basic ingredient.

Formulation Composition:

1	Mārkava Svarasa (Bhrṅgarāja API)	<i>Eclipta alba</i>	Pl.	3.072 l
2	Taila (Tila Taila API)	Sesame oil	–	768 ml
3	Mañjiṣṭhā API	<i>Rubia cordifolia</i>	Rt.	48 g
4	Padmaka API	<i>Prunus cerasoides</i>	Ht. Wd.	48 g
5	Lodhra API	<i>Symplocos racemosa</i>	St. Bk.	48 g
6	Candana (Rakta Candana API)	<i>Pterocarpus santalinus</i>	Ht. Wd.	48 g
7	Gairika API - Śuddha	Red ochre	–	48 g
8	Balā API	<i>Sida cordifolia</i>	Rt.	48 g
9	Rajanīdvaya			
	a. Haridrā API	<i>Curcuma longa</i>	Rz.	48 g
	b. Dāruharidrā API	<i>Berberis aristata</i>	St.	48 g
10	Keśara (Nāgakeśara API)	<i>Mesua ferrea</i>	Stmn.	48 g
11	Priyaṅgu API	<i>Callicarpa macrophylla</i>	Fl.	48 g
12	Madhuyasṭikā (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	48 g
13	Prapaṇḍarīka API	<i>Nelumbo nucifera</i>	Rt.	48 g
14	Gopī (Śveta Sārivā API)	<i>Hemidesmus indicus</i>	Rt.	48 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccḥita Tila taila* (Appendix 6.2.9.4).
- Treat *Gairika* to prepare *Gairika - Śuddha* (Appendix 6.2.8.2).
- Take fresh *Bhrṅgarāja* and wash thoroughly with water. Grind and filter through *muslin cloth* to obtain *svarasa*.
- Wash, clean, dry the ingredients numbered 3 to 6 and 8 to 14 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).
- Transfer the *kalka dravya* to the wet grinder alongwith *Gairika - Śuddha* and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrccḥita Tila Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *svarasa*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.

- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

A medicated oil, light brownish yellow in color with a pleasant odour

Identification:

Thin layer chromatography:

Shake 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.18, 0.23, 0.29, 0.34, 0.44 and 0.77 (all pink changing to black) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.451 to 1.464,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.910 to 0.932,	Appendix 3.2
<i>Saponification value:</i>	188 to 194,	Appendix 3.10
<i>Iodine value:</i>	90 to 100,	Appendix 3.11
<i>Acid value:</i>	3 to 6,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Keśapāta (loss of hair); Netraroga (diseases of eyes); Karṇaroga (diseases of ear); Śīroroga (diseases of head); Manyāstambha (neck rigidity/torticollis); Galagraha (difficulty in swallowing); Khālitya (premature baldness); Indralupta (Alopecia areata)

Dose:

External application over the scalp for Śīro-Abhyaṅga

BRĤAT SAINDHAVĀDYA TAILA

(AFI Part I, 8:40)

Definition:

Bṛhat Saindhavādyā Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Eraṇḍa taila as the basic ingredient.

Formulation Composition:

1	Saindhava (Saindhava Lavaṇa (API))	Rock salt	—	24 g
2	Śreyasī (Gajapippalī API)	<i>Scindapsus officinalis</i>	Fr.	24 g
3	Rāsnā API	<i>Pluchea lanceolata</i> [<i>Alpinia galanga</i> (Official Substitute)]	Rt.	24 g
4	Śatapušpā (Śatāhvā API)	<i>Anethum sowa</i>	Fr.	24 g
5	Yamānikā (Yavānī API)	<i>Trachyspermum ammi</i>	Fr.	24 g
6	Sarjikā (Svarjīkṣāra (API))	Crude alkaline earth	—	24 g
7	Marica API	<i>Piper nigrum</i>	Fr.	24 g
8	Kuṣṭha API	<i>Saussurea lappa</i>	Rt.	24 g
9	Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	24 g
10	Sauvarcala (Sauvarcala Lavaṇa (API))	Black salt	—	24 g
11	Viḍa (Viḍa Lavaṇa)			24 g
12	Vacā API	<i>Acorus calamus</i>	Rz.	24 g
13	Ajamodā API	<i>Apium leptophyllum</i>	Fr.	24 g
14	Madhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	24 g
15	Jīraka (Śveta Jīraka API)	<i>Cuminum cyminum</i>	Fr.	24 g
16	Pauṣkara (Puṣkara API)	<i>Inula racemosa</i>	Rt.	24 g
17	Kaṇā (Pippalī API)	<i>Piper longum</i>	Fr.	24 g
18	Eraṇḍa Taila (API)	Castor oil	—	768 g
19	Śatapušpajāmbu (Śatāhvā API)	<i>Anethum sowa</i>	Fr.	768 g
	Jala API for phaṇṭa	Potable Water	—	3.0720 l
20	Kāñjika (API)	Sour gruel	—	1.536 l
21	Mastu (Godadhi (API))	Curd whey from cow milk	—	1.536 l

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat Eraṇḍa taila to prepare mūrccchita Eraṇḍa taila (Appendix 6.2.9.1).
- A day prior to the sneha pāka, initiate to prepare Mastu (Appendix 6.2.5).
- Wash, clean, dry the ingredient number 19 of the Formulation Composition, powder and pass through 355 μm I. S. sieve (sieve number 44) (kvātha dravya).

- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, clean, dry the ingredients numbered 2 to 9 and 12 to 17 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve. Roast coarsely powdered *Saindhava Lavaṇa*, *Sauvarcala Lavaṇa* and *Viḍa Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder separately and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).
- Transfer the *kalka dravya* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrcchita Eraṇḍa Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *Mastu* and *Kāñjika* and continue the process of heating intermittently over a period of five days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, golden yellow in color, odour unpleasant and characteristic of castor oil

Identification:

Thin layer chromatography:

Shake 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 μl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.25, 0.43, 0.64 and 0.71 in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.460 to 1.467,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.947 to 0.974,	Appendix 3.2
<i>Saponification value:</i>	178 to 200,	Appendix 3.10
<i>Iodine value:</i>	70 to 90,	Appendix 3.11
<i>Acid value:</i>	5 to 10,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 5,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Ānāha (distension of abdomen); Antravṛddhi (Hernia); Mūtrakṛcchra (Dysuria); Aśmarī (calculus); Hṛtśūla (Angina pectoris); Pārśvaśūla (intercostal neuralgia and polydynia); Ardita (facial palsy); Āmavāta (Rheumatism); Sandhigata Vāta (osteoarthropathy); Mandāgni (reduced digestive fire); Vāta roga (diseases due to Vāta Doṣa); Kaṭiśūla (lower backache); Jānuśūla (pain in the knee); Ūruśūla (pain in the thigh region); Pṛṣṭhaśūla (backache); Bāhyāyāma (opisthotonus)

Dose:

5 ml per day in divided doses; also used for Abhyaṅga and Basti

CITRAKĀDI TAILA

(AFI Part I, 8:16)

Definition:

Citrakādi Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Tila taila as the basic ingredient.

Formulation Composition:

1	Citraka API	<i>Plumbago zeylanica</i>	Rt.	16 g
2	Arkamūla (Arka API)	<i>Calotropis procera</i>	Rt.	16 g
3	Trivṛt API	<i>Ipomoea turpethum</i>	Rt.	16 g
4	Pāṭhā API	<i>Cissampelos pareira</i>	Rt.	16 g
5	Malapū (Phalgu API)	<i>Ficus hispida</i>	Rt. Bk.	16 g
6	Hayamāraka (Karavīra API)	<i>Nerium indicum</i>	Rt. Bk.	16 g
7	Sudhā (Snuhī API)	<i>Euphorbia neriifolia</i>	Rt.	16 g
8	Vacā API	<i>Acorus calamus</i>	Rz	16 g
9	Lāṅgalakī (Lāṅgalī API)	<i>Gloriosa superba</i>	Rz./Rt.	16 g
10	Saptaparṇa API	<i>Alstonia scholaris</i>	St. Bk.	16 g
11	Suvarcikā (Svarjīkṣāra (API))	Crude alkaline earth	—	16 g
12	Jyotiṣmatī API	<i>Celastrus paniculatus</i>	Sd.	16 g
13	Taila (Tila Taila API)	Sesame oil	—	768 g
14	Jala API	Potable Water	—	3.072 l

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccchita Tila taila* (Appendix 6.2.9.4).
- Treat *Citraka* to prepare *Citraka - Śuddha* (Appendix 6.2.8.11), *Karavīra* to prepare *Karavīra - Śuddha* (Appendix 6.2.8.10), and *Lāṅgalī* to prepare *Lāṅgalī - Śuddha* (Appendix 6.2.8.12); powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder.
- Wash, clean, dry the ingredients numbered 3 to 5, 8, 10 and 12 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder.
- Roast *Svarjīkṣāra* in a stainless steel pan on low flame till free from moisture.
- Transfer all the ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrccchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding water.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.

- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, reddish brown in colour with a faint odour

Identification:

Thin Layer Chromatography:

Shake 1 ml and carry out thin layer chromatography of formulation with 10 ml of *chloroform* and the mixture for 12 h at 37⁰. Filter the extract. Apply 3 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: glacial acetic acid* (8: 2: 0.1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with 10 per cent *methanolic sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.37, 0.42, 0.50 (all reddish brown) and 0.55 (brown) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4646 to 1.4665,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.8319 to 0.8539,	Appendix 3.2
<i>Saponification value:</i>	185 to 200,	Appendix 3.10
<i>Iodine value:</i>	100 to 102,	Appendix 3.11
<i>Acid value:</i>	4 to 5,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 1,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Bhagandara (fistula- in- ano)

Dose:

External use: for Abhyaṅga (massage on whole body) and local application

HINGVĀDI TAILA

(AFI Part II, 8:18)

Definition:

Hiṅgvādi Taila is a medicated preparation made with the ingredients in the Formulation Composition given below with mūr̄cchita Sarṣapa Taila as basic ingredient.

Formulation Composition:

1	Hiṅgu API	<i>Ferula foetida</i>	Exd.	1 part
2	Tumburu (Tejovatī API)	<i>Zanthoxylum armatum</i>	Fr.	1 part
3	Śuṅṭhī API	<i>Zingiber officinalis</i>	Rz.	1 part
4	Sārṣapa Taila (Sarṣapa Taila API)	Mustard oil	—	12 parts
5	Jala API	Potable Water	—	48 parts

Method of Preparation

- Take all ingredients of pharmacopoeial quality.
- Treat *Sarṣapa taila* to prepare *mūr̄cchita Sarṣapa taila* (Appendix 6.2.9.3).
- Treat *Hiṅgu* to prepare *Hiṅgu - Śuddha* (Appendix 6.2.8.15).
- Wash, clean, dry the ingredients numbered 2 and 3 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve. Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogenous blend (*kalka*).
- Take *mūr̄cchita Sarṣapa Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*. Stir thoroughly while adding the water in specified ratio.
- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti (khara pāka lakṣaṇa)* and at the appearance of froth (phenodgama) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, reddish in colour with characteristic odour of asafoetida

Identification:

Thin layer chromatography:

a) Shake 0.5 ml of formulation with 10 ml of *methanol* and the mixture overnight at 37⁰ with occasional shakings. Filter the extract and carry out the thin layer chromatography. Apply 10 μl

of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: acetone: methanol* (8.5: 1.5: 0.2) as mobile phase. After development, allow the plate to dry in air and spray with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and examine the plate under ultraviolet light (366nm). It shows major spots at R_f 0.35 (fluorescent blue), 0.45 (pinkish blue), 0.50 (light blue) and 0.65, 0.75 (both light pink).

b) Extract 2 ml of formulation with 10 ml of *petroleum ether* (60⁰-80⁰) by keeping the mixture for about 15 min. Filter the extract and dry the filtrate on water-bath at 35⁰ and dissolve the residue in 10 ml of *ethanol*, filter and carry out the thin layer chromatography. Apply 5 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: formic acid* (14: 8: 0.15) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and examine the plate under ultraviolet light (366nm). It shows major spots at R_f 0.25 (fluorescent light blue), 0.30 (light fluorescent blue, corresponding to ferulic acid) 0.35 (faint yellow), 0.43 (blue), 0.56 (light blue), 0.69 (fluorescent blue), 0.78 (white), 0.81, 0.84 and 0.88 (all pink).

Chemical Test:

Complies with *Test for Hingu* (Appendix 5.14.13)

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.447 to 1.478,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.880 to 0.918,	Appendix 3.2
<i>Saponification value:</i>	182 to 186,	Appendix 3.10
<i>Iodine value:</i>	100 to 105,	Appendix 3.11
<i>Acid value:</i>	Not more than 1,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Karnaśūla (otalgia)

Dose:

Externally as ear drops- 5-10 drops in the affected ear once or twice a day (Karnaṇpūraṇa)

JYOTIṢMATĪ TAILA

(AFI Part I, 8:18)

Definition:

Jyotiṣmatī Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Mayūrakakṣāra (Apāmārgakṣāra (API))	<i>Achyranthes aspera</i>	Water soluble Ash of Pl.	896 g
2	Jala API	Potable water		5.376 l
3	Jyotiṣmatī Taila (API)	<i>Celastrus paniculatus</i>	Oil	768 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Add 6 parts of water to 1 part of *Kṣāra* (Appendix 6.2.3), and dissolve completely (*Kṣāra jala*).
- Take *Jyotiṣmatī Taila* in a stainless steel vessel and heat it.
- Add equal quantity of *Kṣāra jala*. Stir thoroughly and heat with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop the heating when the *kalka* gets separated at the bottom of the vessel in the form of loose paste (*mṛdu pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Allow to stand overnight.
- Repeat the process for six more times each day adding fresh *Kṣāra jala* in equal quantity.
- On the last day, constantly check the *kalka* by rolling between the fingers. Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Very thick oil, reddish in colour

Identification:

Thin Layer Chromatography:

Shake 1 ml of formulation in 10 ml of *toluene* and the mixture for 12 h at 37⁰. Filter the extract. Apply 4 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: glacial acetic acid* (7: 2: 0.2) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and examine under ultraviolet light (366 nm). The plate shows major spots at R_f

0.15 (fluorescent green), 0.24 (faint green), 0.32 (pink), 0.35 (fluorescent green), 0.40 (pink), 0.49 (faint blue), 0.53 (light blue) and 0.67(pink).

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4232 to 1.4556,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.8506 to 0.9232,	Appendix 3.2
<i>Saponification value:</i>	106 to 108,	Appendix 3.10
<i>Iodine value:</i>	51 to 55,	Appendix 3.11
<i>Acid value:</i>	3 to 4,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 4,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	complies with Appendix 2.4
<i>Aflatoxins:</i>	complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Śvitarroga (leucoderma/ vitiligo)

Dose:

Externally for Abhyaṅga (application on affected area)

KANAKA TAILA

(AFI Part I, 8:4)

Definition:

Kanaka Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with *mūrcchita Tila taila* as the basic ingredient.

Formulation Composition:

1	Madhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	768 g
2	Jala API for decoction reduced to	Potable Water	—	3.072 ml 768 ml
3	Taila (Tila Taila API)	Sesame oil	—	192 g
4	Priyaṅgu API	<i>Calicarpa macrophylla</i>	Fl.	6.4 g
5	Mañjiṣṭhā API	<i>Rubia cordifolia</i>	St.	6.4 g
6	Candana (Rakta Candana API)	<i>Pterocarpus santalinus</i>	Ht. Wd.	6.4 g
7	Utpala API	<i>Nymphaea stellata</i>	Fl.	6.4 g
8	Keśara (Nāgakeśara API)	<i>Mesua ferrea</i>	Stmn.	6.4 g

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrcchita Tila taila* (Appendix 6.2.9.4).
- Wash, clean, dry the ingredient number 1 of the Formulation Composition, powder and pass through 355 µm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, dry the ingredients numbered 4 to 8 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85) to obtain fine powder.
- Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrcchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*Madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, reddish brown in colour with a faint odour

Identification:*Thin Layer Chromatography:*

Shake 2 ml of formulation with 10 ml of *ethyl acetate*, keep the mixture for 12 h and filter. Dilute the extract in *ethyl acetate* (1: 4). Apply 5 µl of the diluted extract on the TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: chloroform: methanol* (8.5: 1.2: 0.2: 0.2) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and examine under ultraviolet light (366 nm). The plate shows major spots at R_f 0.11, 0.31, 0.46, 0.54, 0.65 (all blue), 0.74 (black) and 0.80 (faint blue).

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4641 to 1.4660,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.9180 to 0.9230,	Appendix 3.2
<i>Saponification value:</i>	190 to 200,	Appendix 3.10
<i>Iodine value:</i>	103 to 106,	Appendix 3.11
<i>Acid value:</i>	Not more than 2,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 2,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Mukha roga (diseases of oral cavity); Vyañga (local hyper pigmentation); Nīlikā (mole)

Dose:

External use: for Abhyañga (apply on whole body)

Nasya - 2 to 4 drops per nostril once or twice a day

MAHĀNĀRĀYAṆA TAILA

(AFI Part I, 8:45)

Definition:

Mahānārāyaṇa Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccḥita Tila taila as the basic ingredient.

Formulation Composition:

1	Bilva API	<i>Aegle marmelos</i>	St. Bk.	960 g
2	Aśvagandhā API	<i>Withania somnifera</i>	Rt.	960 g
3	Bṛhatī API	<i>Solanum indicum</i>	Pl.	960 g
4	Śvadaṃṣṭrā (Gokṣura API)	<i>Tribulus terrestris</i>	Pl.	960 g
5	Śyonāka API	<i>Oroxylum indicum</i>	St. Bk.	960 g
6	Vāṭyālakā (Balā (API))	<i>Sida cordifolia</i>	Rt.	960 g
7	Pāribhadra API	<i>Erythrina indica</i>	Rt.	960 g
8	Kṣudrā (Kaṇṭakārī API)	<i>Solanum surattense</i>	Pl.	960 g
9	Kaṭhillā (Rakta Punarnavā API)	<i>Boerhavia diffusa</i>	Pl.	960 g
10	Atibalā API	<i>Abutilon indicum</i>	Rt.	960 g
11	Agnimantha API	<i>Clerodendrum phlomidis</i> [<i>Premna integrifolia</i> (Official Substitute)]	St. Bk.	960 g
12	Saraṇī (Prasārī API)	<i>Paedaria foetida</i>	Pl.	960 g
13	Pāṭalī (Pāṭalā API)	<i>Stereospermum suaveolens</i>	St. Bk.	960 g
14	Jala API for decoction reduced to	Potable Water	—	98.304 l 24.576 l
15	Taila (Tila Taila API)	Sesame oil	—	6.144 kg
16	Godugdha* API	Cow milk	—	6.144 kg
17	Śatāvarīrasa (Śatāvarī API)	<i>Asparagus racemosus</i>	Rt. Tr. juice	6.144 l
18	Rāsnā API	<i>Pluchea lanceolata</i> [<i>Alpinia galanga</i> (Official substitute)]	Rt.	96 g
19	Aśvagandhā API	<i>Withania somnifera</i>	Rt.	96 g
20	Miṣi (Miśreyā API)	<i>Foeniculum vulgare</i>	Fr.	96 g
21	Dāru (Devadāru API)	<i>Cedrus deodara</i>	Ht. Wd.	96 g
22	Kuṣṭha API	<i>Saussurea lappa</i>	Rt.	96 g
23	Parṇīcatuṣṭka			
	a. Śālaparṇī API	<i>Desmodium gangeticum</i>	Pl.	96 g
	b. Pṛṣniparṇī API	<i>Uraria picta</i>	Pl.	96 g

* Ajādugdha (Goat milk) may also be alternatively used.

c. Mudgaparṇī API	<i>Phaseolus trilobus</i>	Pl.	96 g
d. Māṣaparṇī API	<i>Teramnus labialis</i>	Pl.	96 g
24 Agarū API	<i>Aquillaria agallocha</i>	Ht. Wd.	96 g
25 Keśara (Nāgakeśara API)	<i>Messua ferrea</i>	Stmn.	96 g
26 Sindhūttha (Saindhava Lavaṇa (API))	Rock salt	—	96 g
27 Māṃsī (Jaṭāmāṃsī API)	<i>Nardostachys jatamansi</i>	Rt./Rz.	96 g
28 Rajanīdvaya			
a. Haridrā API	<i>Curcuma longa</i>	Rz.	96 g
b. Dāruharidrā API	<i>Berberis aristata</i>	St.	96 g
29 Śaileyaka (Śaileya API)	<i>Parmelia perlata</i>	Pl.	96 g
30 Candana (Rakta candana API)	<i>Pterocarpus santalinus</i>	Ht. Wd.	96 g
31 Puṣkara API	<i>Inula racemosa</i>	Rt.	96 g
32 Elā (Sūkṣmailā API)	<i>Elettaria cardamomum</i>	Sd.	96 g
33 Asra (Mañjiṣṭhā API)	<i>Rubia cordifolia</i>	St.	96 g
34 Yaṣṭī API	<i>Glycyrrhiza glabra</i>	Rt.	96 g
35 Tagara API	<i>Valeriana wallichii</i>	Rz.	96 g
36 Abda (Mustā API)	<i>Cyperus rotundus</i>	Rz.	96 g
37 Patra (Tvak patra API)	<i>Cinnamomum tamala</i>	Lf.	96 g
38 Bhr̥ṅga (Tvak API)	<i>Cinnamomum zeylanicum</i>	St. Bk.	96 g
39 Aṣṭavarga			
a. Jīvaka API	<i>Microstylis wallichii</i> [<i>Pueraria tuberosa</i> (Official Substitute)]	Rt. Tr.	96 g
b. Ṛṣabhaka API	<i>Microstylis wallichii</i> [<i>Pueraria tuberosa</i> (Official Substitute)]	Rt. Tr.	96 g
c. Medā (API)	<i>Polygonatum verticillatum</i> [<i>Asparagus racemosus</i> (Official Substitute)]	Rt. Tr.	96 g
d. Mahāmedā API	<i>Polygonatum cirrhifolium</i> [<i>Asparagus racemosus</i> (Official Substitute)]	Rt. Tr.	96 g
e. Kākolī API	<i>Lilium polyphyllum</i> [<i>Withania somnifera</i> (Official Substitute)]	Rt.	96 g
f. Kṣīrakākolī API	<i>Fritillaria roylei</i> [<i>Withania somnifera</i> (Official Substitute)]	Rt.	96 g
g. Ṛddhi API	<i>Habenaria intermedia</i> [<i>Dioscorea bulbifera</i> (Official Substitute)]	Rt. Tr.	96 g

h. Vṛddhi API	<i>Habenaria intermedia</i> [<i>Dioscorea bulbifera</i> (Official Substitute)]	Rt. Tr.	96 g
40 Ambu (Hrīvera API)	<i>Coleus vettiveroides</i>	Rt.	96 g
41 Vacā API	<i>Acorus calamus</i>	Rz.	96 g
42 Palāśa API	<i>Butea monosperma</i>	St. Bk.	96 g
43 Sthauṇeya API	<i>Taxus baccata</i>	Lf.	96 g
44 Vṛścīraka (Śveta Punarnavā API)	<i>Boerhaavia verticillata</i>	Pl.	96 g
45 Coraka API	<i>Angelica glauca</i>	Rz.	96 g
46 Karpūra API	<i>Cinnamomum camphora</i>	Subl. Ext. of A. Pt.	96 g
47 Kāśmīra (Kuṅkuma API)	<i>Crocus sativus</i>	Sty./Stg.	96 g

Method of Preparation

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrcchita Tila taila* (Appendix 6.2.9.4).
- Wash, clean, dry the ingredients numbered 1 to 13 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash and clean the cut pieces of *Śatāvarī*, grind and squeeze through *muslin cloth* to obtain *Śatāvarī svarasa*.
- Wash, clean, dry the ingredients numbered 21 to 25 and 27 to 47 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve. Roast coarsely powdered *Saindhava Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).
- Transfer the *kalka dravya* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Strain the *Godugdha* through *muslin cloth*.
- Take *mūrcchita Eraṇḍa Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*, *svarasa*.
- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *Godugdha* and continue the process of heating intermittently over a period of five days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.

- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, yellowish brown in color with an unpleasant odour

Identification:

Thin layer chromatography:

Shake 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.28, 0.41, 0.54 and 0.65 (all brown) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.458 to 1.461,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.926 to 0.955,	Appendix 3.2
<i>Saponification value:</i>	188 to 196,	Appendix 3.10
<i>Iodine value:</i>	80 to 100,	Appendix 3.11
<i>Acid value:</i>	3 to 6,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 5,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in air-tight containers, protected from light and moisture.

Therapeutic uses:

Ardita (facial palsy); Badhiratva (deafness); Paṅgutva (paraplegia); Gātra kampa (tremors); Manyāstambha (neck rigidity/torticollis) Hanustambha (lockjaw); Ekāṅgaśoṣa (wasting of one limb); Śukrakṣaya (oligospermia); Vandhyatva (infertility); Śiroroga (headache), Jihvāstambha (glossal palsy); Dantaśūla (toothache); Unmāda (Mania/psychosis); Kubja (humpback/ kyphosis); Jvara (fever); Jarā (senility); Kārśya (emaciation); Snāyubhagna (rupture of ligament); Asthibhagna (bone fracture)

Dose:

5-10 ml per day in divided doses; Nasya: 2 to 3 drops per nostril; Abhyaṅga (massage on affected parts) and Anuvāsana Basti

NĀLPĀMARĀDI TAILA

(AFI Part I, 8:24)

Definition:

Nālpāmarādi taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūr̥cchita Tila taila as the basic ingredient.

Formulation Composition:

1	Paimannal rasa (Haridrā (API)) - Ār̥dra	<i>Curcuma longa</i>	Rz. fresh	1.536 l
2	Parpaṭa API - rasa	<i>Fumaria parviflora</i>	Pl.	1.536 l
3	Eṇṇa (Tila Taila API)	Sesame oil	—	768 ml
4	Nyagrodha API	<i>Ficus bengalensis</i>	St. Bk	15 g
5	Udumbara API	<i>Ficus racemosa</i>	St. Bk	15 g
6	Aśvattha API	<i>Ficus religiosa</i>	St. Bk	15 g
7	Plakṣa API	<i>Ficus lacor</i>	St. Bk	15 g
8	Harītakī API	<i>Terminalia chebula</i>	P.	15 g
9	Bibhītaka API	<i>Terminalia bellirica</i>	P.	15 g
10	Āmalakī API	<i>Embllica officinalis</i>	P.	15 g
11	Candana (Rakta candana API)	<i>Pterocarpus santalinus</i>	Ht. Wd	15 g
12	Sevya (Uśīra API)	<i>Vetiveria zizanioides</i>	Rt.	15 g
13	Kuṣṭha API	<i>Saussurea lappa</i>	Rt.	15 g
14	Covalli (Mañjiṣṭhā API)	<i>Rubia cordifolia</i>	St.	15 g
15	Coram (API)	<i>Kaempferia galanga</i>	Rz.	15 g
16	Akil (Agaru API)	<i>Aquilaria agallocha</i>	Ht. Wd	15 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūr̥cchita Tila taila* (Appendix 6.2.9.4).
- Take fresh *Haridrā* and wash thoroughly. Grind and filter through *muslin cloth* to obtain *svarasa*.
- Wash, clean, dry the *Parpaṭa*, powder and pass through 355 μm I. S. sieve. Add eight times water to it and soak for 4 h. Boil and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *Parpaṭa rasa*. *
- Wash, dry the ingredients numbered 4 to 16 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder. Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūr̥cchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *Haridrā svarasa* and *Parpaṭa rasa*.

* Dried whole plant of *Parpaṭa* is used and *Parpaṭa rasa* is prepared as per Śār̥ṅgadhara saṃhitā-Madhyamakhaṇḍa, 1/4.

- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, reddish brown in colour with an odour of turmeric

Identification:

Thin Layer Chromatography:

Shake 5 ml of formulation with 10 ml of *acetonitrile* and keep the mixture for 12 h. Filter and carry out the thin layer chromatography. Apply 5 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toulene: ethyl acetate: chloroform: methanol* (8: 1: 0.2: 0.2) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light (254 nm). The plate shows major spots at R_f 0.21, 0.46, 0.56, 0.68, 0.78 (all black).

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4646 to 1.4665,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.7840 to 0.8235,	Appendix 3.2
<i>Saponification value:</i>	190 to 200,	Appendix 3.10
<i>Iodine value:</i>	102 to 110,	Appendix 3.11
<i>Acid value:</i>	2 to 4,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Tvagroga (skin diseases); Visarpa (Erysepales); Kuṣṭha (diseases of skin); Pāmā (eczema); Kaṇḍū (itching); Piḍikā (boil/carbuncle)

Dose:

External application for Abhyaṅga

NĪLĪBHRŅĠĠADI TAILA

(AFI Part I, 8:26)

Definition:

NĪlĪbhrŅĠĠadi Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mŭrcchita Tila taila as the basic ingredient.

Formulation Composition:

1	NĪlĪ Rasa (NĪlĪ API)	<i>Indigofera tinctoria</i>	Lf. juice	768 ml
2	BhrŅĠĠa Rasa (BhrŅĠĠarĀĵa API)	<i>Eclipta alba</i>	Pl. juice	768 ml
3	Śatakṛatu latĀ Rasa (KĀkatiktĀ API)	<i>Cardiospermum halicacabum</i>	Pl. juice	768 ml
4	DhĀtrĪ Phala Rasa (ĀmalakĪ API)	<i>Emblica officinalis</i>	P. juice	768 ml
5	Āĵaka KṣĪra (ĀĵĀdugdha (API))	Goat milk		768 ml
6	NĀlikera KṣĪra (NĀrikela (API))	<i>Cocos nucifera</i>	Milk from End.	768 ml
7	MahiṣĪ KṣĪra (MahiṣĪdugdha (API))	Buffalo milk	–	768 ml
8	Dhenŭdbhava (Godugdha API)	Cow milk	–	768 ml
9	Taila (Tila Taila API)	Sesame oil	–	768 ml
10	YaṣṭyĀhva (YaṣṭĪ API)	<i>Glycyrrhiza glabra</i>	Rt.	32 g
11	GuŅĵĀ API	<i>Abrus precatorius</i>	Rt.	32 g
12	AŅĵana (RasāŅĵana (API))	<i>Berberis sps.</i>	Ext.	32 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mŭrcchita Tila taila* (Appendix 6.2.9.4).
- Take fresh *NĪlĪ*, *BhrŅĠĠarĀĵa*, *KĀkatiktĀ* and *ĀmalakĪ* and wash thoroughly with water. Grind and filter through *muslin cloth* to obtain *svarasa*.
- Wash and clean the cut pieces of *NĀrikela*, grind and squeeze through *muslin cloth* to obtain *NĀrikela KṣĪra*.
- Strain the *ĀĵĀdugdha*, *MahiṣĪdugdha* and *Godugdha* through *muslin cloth*.
- Wash, clean, dry the ingredients numbered 10 to 12 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder. Transfer the powdered ingredients to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mŭrcchita Tila taila* in a Stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *svarasa*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *NĀrikela KṣĪra*, *ĀĵĀdugdha*, *MahiṣĪdugdha* and *Godugdha* and continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.

- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti* (*khara pāka lakṣaṇa*) and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, dark brown in colour, odour characteristic of sesame oil

Identification:

Thin layer chromatography :

Extract 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.18, 0.23, 0.35, 0.45 and 0.82 (all brown) in visible light.

Physicochemical parameters:

<i>Refractive index at 40⁰:</i>	1.451 to 1.462,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.905 to 0.928,	Appendix 3.2
<i>Saponification value:</i>	194 to 212,	Appendix 3.10
<i>Iodine value:</i>	80 to 90,	Appendix 3.11
<i>Acid value:</i>	9 to 11,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 4,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Keśapāta (hair fall) and Palita (premature greying of hair)

Dose:

External application for Śiro-abhyaṅga

PAÑCAGUṆA TAILA

(AFI Part II, 8:7)

Definition:

Pañcaguṇa Taila is a medicinal oil preparation made with the ingredients in the Formulation Composition given below with mūrccḥita Tila Taila as the basic ingredient.

Formulation Composition:

1	Harītakī API	<i>Terminalia chebula</i>	P.	60 g
2	Āmalakī API	<i>Phyllanthus emblica</i>	P.	60 g
3	Bibhītaka API	<i>Terminalia bellirica</i>	P.	60 g
4	Nimba patra (Nimba API)	<i>Azadirachta indica</i>	Lf.	180 g
5	Sambhālu patra (Nirguṇḍī API)	<i>Vitex negundo</i>	Lf.	180 g
6	Jala API for decoction reduced to	Potable Water	—	4.320 l 1.080 l
7	Tila Taila API	Sesame oil	—	960 ml
8	Moma (Madhūcchiṣṭa (API))	Hive of Apis species	Bees wax	48 g
9	Gandhavirojā (Sarala API)	<i>Pinus roxburghii</i>	Resin	48 g
10	Śīlārasa (Turuṣka (API))	<i>Liquidamber orientalis</i> [<i>Altingia excelsa</i> (Official Substitute)]	Exd.	48 g
11	Rāla (Śāla API)	<i>Shorea robusta</i>	Exd.	48 g
12	Guggulu API - Śuddha	<i>Commiphora wightii</i>	O.R.	48 g
13	Karpūra API	<i>Cinnamomum camphora</i>	Subl. Ext. of A. Pt.	60 g
14	Tārapīna Taila (API)	<i>Pinus roxburghii</i>	Volatile Oil	30 ml
15	Tailaparna Taila API	<i>Eucalyptus globulus</i>	Volatile Oil	30 ml
16	Kejopuṭī Taila (API)	<i>Melaleuca leucodendron</i>	Fixed Oil	30 ml

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccḥita Tila taila* (Appendix 6.2.9.4).
- Treat *Guggulu* to get *Guggulu - Śuddha*. (Appendix 6.2.8.4).
- Wash, clean, dry the ingredients numbered 1 to 5 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Take *mūrccḥita Tila taila* in a Stainless steel vessel and heat it.
- Add increments of ingredients numbered 8 to 12 of the Formulation Composition, stir thoroughly while adding *kvātha*.

- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days.
- Expose the oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth*, add powdered *Karpūra* and allow to cool.
- Add *Tārapīna Taila*, *Tailaparna Taila* and *Kejopuṭī Taila* and mix thoroughly.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, thick, reddish yellow in colour with characteristic odour of Eucalyptus

Identification:

Thin layer chromatography

Shake 1 ml of formulation with 10 ml of *toluene* and keep the mixture for 12 h. Filter the extract and carry out thin layer Chromatography. Apply 3 µl of the diluted extract on TLC plate. Develop the plate upto a distance of 8 cm using *toluene: ethyl acetate: methanol: glacial acetic acid* (8.5: 0.5: 0.2: 0.1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for 10 min. The plate shows major spots at R_f 0.21, 0.27, 0.32, 0.38, 0.42, 0.48, 0.55, 0.71, 0.81, 0.84 (all brown) in visible light.

Physicochemical parameters:

<i>Refractive index at 40⁰:</i>	1.4703 to 1.503,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.9106 to 0.9259,	Appendix 3.2
<i>Saponification value:</i>	160 to 184,	Appendix 3.10
<i>Iodine value:</i>	104 to 107,	Appendix 3.11
<i>Acid value:</i>	Not more than 2,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Sandhivāta (osteoarthropathy); Karṇaśūla (otalgia); Vraṇopacāra (wounds)

Dose:

Externally on wound and for Abhyaṅga on whole body

PRABHAÑJANA VIMARDANA TAILA

(AFI Part I, 8:30)

Definition:

Prabhañjana Vimardana taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Tila taila as the basic ingredient.

Formulation Composition:

1	Balā (API)	<i>Sida cordifolia</i>	Rt.	25.6 g
2	Śatāvarī API	<i>Asparagus racemosus</i>	Rt. Tr.	25.6 g
3	Śigru API	<i>Moringa oleifera</i>	St. Bk.	25.6 g
4	Varuṇa API	<i>Crataeva nurvala</i>	St. Bk.	25.6 g
5	Arka API	<i>Calotropis procera</i>	Rt.	25.6 g
6	Karañjaka (Karañja API)	<i>Pongamia pinnata</i>	St. Bk.	25.6 g
7	Eraṇḍa API	<i>Ricinus communis</i>	Rt.	25.6 g
8	Koraṇṭa (Sahacara API)	<i>Barleria prionitis</i>	Pl.	25.6 g
9	Vājigandhā (Aśvagandhā API)	<i>Withania somnifera</i>	Rt.	25.6 g
10	Prasāriṇī API	<i>Paederia foetida</i>	Pl.	25.6 g
11	Variṣṭha Pañcamūla			
	a. Bilva API	<i>Aegle marmelos</i>	St. Bk.	25.6 g
	b. Śyonāka API	<i>Oroxylum indicum</i>	Rt.	25.6 g
	c. Gambhārī API	<i>Gmelina arborea</i>	St. Bk.	25.6 g
	d. Pāṭalā API	<i>Stereospermum suaveolens</i>	St. Bk.	25.6 g
	e. Agnimantha API	<i>Clerodendrum phlomidis</i>	Rt.	25.6 g
		[<i>Premna integrifolia</i> (Official Substitute)]		
12	Jala API for decoction reduced to	Potable Water	—	24.576 l 6.144 l
13	Taila (Tila Taila API)	Sesame oil	—	1.536 kg
14	Kṣīra (Godugdha API)	Cow milk	—	3.072 l
15	Dadhi (Godadhi (API))	Curd from cow milk	—	1.536 kg
16	Kāñjika (API)	Sour gruel	—	1.536 l
17	Tagara API	<i>Valeriana wallichii</i>	Rz.	12 g
18	Amarakāṣṭha (Devadāru API)	<i>Cedrus deodar</i>	Ht. Wd.	12 g
19	Elā (Sūkṣmailā API)	<i>Elettaria cardamomum</i>	Sd.	12 g
20	Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	12 g
21	Sarṣapa API	<i>Brassica campestris</i>	Sd.	12 g
22	Coraka API	<i>Angelica glauca</i>	Rz.	12 g
23	Śatāhvā API	<i>Anethum sowa</i>	Fr.	12 g
24	Kuṣṭha API	<i>Saussurea lappa</i>	Rt.	12 g

25	Sindhūttha (Saindhava Lavaṇa (API))	Rock salt	—	12 g
26	Rāsnā API	<i>Pluchea lanceolata</i> [<i>Alpinia galanga</i> (Official Substitute)]	Rz.	12 g
27	Kālānusārikā (Methī API)	<i>Trigonella foenum-graecum</i>	Sd.	12 g
28	Vacā API	<i>Acorus calamus</i>	Rz.	12 g
29	Citraka API	<i>Plumbago zeylanica</i>	Rt.	12 g
30	Māṃsī (Jaṭāmāṃsī API)	<i>Nardostachys jatamansi</i>	Rz.	12 g
31	Sarala API	<i>Pinus roxburghii</i>	Ht. Wd.	12 g
32	Kaṭurohiṇī (Kaṭukā API)	<i>Picrorhiza kurroa</i>	Rz.	12 g

Method of Preparation

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrchita Tila taila* (Appendix 6.2.9.4).
- A day prior to the *sneha pāka*, prepare *Godadhi*.
- Wash, clean, dry the ingredients numbered 1 to 11 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, clean, dry the ingredients numbered 17 to 24 and 26 to 32 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85). Roast coarsely powdered *Saindhava Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder separately and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).
- Transfer the *kalka dravya* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Strain the *Godugdha* through *muslin cloth*.
- Take *mūrchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- next day, add *Godugdha*, *Godadhi* and *Kāñjika* and continue the process of heating intermittently over a period of five days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti (khara pāka lakṣaṇa)* and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, golden yellow in color with a pleasant odour

Identification:

Thin layer chromatography:

Extract 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air and spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105^o for about 10 min. It shows major spots at R_f 0.39 (light green), and 0.47, 0.59 (both light black) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40^o:</i>	1.451 to 1.464,	Appendix 3.1
<i>Specific gravity at 40^o:</i>	0.908 to 0.945,	Appendix 3.2
<i>Saponification value:</i>	187 to 204,	Appendix 3.10
<i>Iodine value:</i>	84 to 95,	Appendix 3.11
<i>Acid value:</i>	3 to 5,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Vātagulma (abdominal lump due to Vāta Doṣa); Vātavidradhi (abscess due to Vāta Doṣa); Antravṛddhi (Hernia); Śūla (pain); Ardita (facial palsy); Vātaroga (diseases due to Vāta Doṣa /neurological disease); Kaṭiśūla (lower backache); Pṛṣṭhaśūla (lumbago); Mūḍhagarbha (malpresentation of the foetus)

Dose:

External application for Abhyaṅga

PRASĀRIṆĪ TAILA

(AFI Part I, 8:32)

Definition:

Prasāriṇī taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below with mūrccchita Tila taila as the basic ingredient.

Formulation Composition:

1	Prasāraṇī (Prasāriṇī API)	<i>Paederia foetida</i>	Pl.	4.8 kg
2	Jala API for decoction reduced to	Potable water	—	12.288 l 3.072 l
3	Taila (Tila Taila API)	Sesame oil	—	3.072 kg
4	Dadhi (Godadhi (API))	Curd from cow milk	—	3.072 kg
5	Kāñjika (API)	Sour gruel	—	3.072 l
6	Kṣīra (Godugdha API)	Cow milk	—	12.288 l
7	Madhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	32 g
8	Pippalīmūla (Pippalī API)	<i>Piper longum</i>	St.	32 g
9	Citraka API	<i>Plumbago zeylanica</i>	Rt.	32 g
10	Saindhava (Saindhava Lavaṇa (API))	Rock salt	—	32 g
11	Vacā API	<i>Acorus calamus</i>	Rz.	32 g
12	Prasāriṇī API	<i>Paederia foetida</i>	Pl.	32 g
13	Devadāru API	<i>Cedrus deodara</i>	Ht. Wd.	32 g
14	Rāsnā API	<i>Pluchea lanceolata</i> [<i>Alpinia galanga</i> (Official Substitute)]	Rz.	32 g
15	Gajapippalī API	<i>Scindapsus officinalis</i>	Fr.	32 g
16	Bhallāta (Bhallātaka API) - Śuddha	<i>Semecarpus anacardium</i>	Fr.	32 g
17	Śatapuṣpā (Śatāhvā API)	<i>Anethum sowa</i>	Fr.	32 g
18	Māmsī (Jaṭāmāmsī API)	<i>Nardostachys jatamansi</i>	Rz.	32 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrccchita Tila taila* (Appendix 6.2.9.4).
- A day prior to the *sneha pāka*, initiate to prepare *Godadhi*.
- Wash, clean, dry the ingredients numbered 1 of the Formulation Composition, powder and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, clean, dry the ingredients numbered 7 to 9 and 11 to 18 of the formulation composition, powder separately and pass through 180 μm I. S. sieve. Roast coarsely

powdered *Saindhava Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder separately and pass through 180 μm I. S. sieve (sieve number 85) (*kalka dravya*).

- Transfer the *kalka dravya* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Strain the *Godugdha* through *muslin cloth*.
- Take *mūrcchita Tila Taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring maintaining the temperature between 50° and 90° during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *Godadhi*, *Kāñjika* and *Godugdha* and continue the process of heating intermittently over a period of five days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* breaks down into pieces on attempting to form a *varti (khara pāka lakṣaṇa)* and at the appearance of froth (*phenodgama*) over the oil. Expose the *kalka* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80°) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, brownish yellow in colour with a pleasant odour

Identification:

Thin layer chromatography:

Shake 2 ml of the formulation with 20 ml of *alcohol* for 3 h. Allow the two layers to separate. Separate the alcohol layer, filter and concentrate to 5 ml and carry out the thin layer chromatography. Apply 10 μl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: n-hexane* (6: 3: 1) as mobile phase. After development, allow the plate to dry in air and spray the plate with *ethanol-sulphuric acid reagent* followed by heating at 105° for about 10 min. It shows major spots at R_f 0.19, 0.21, 0.35, 0.46 and 0.83 (all pink changing to purple) in visible light.

Physico-chemical parameters:

<i>Refractive index at 40°:</i>	1.457 to 1.461,	Appendix 3.1
<i>Specific gravity at 40°:</i>	0.906 to 0.936,	Appendix 3.2
<i>Saponification value:</i>	190 to 200,	Appendix 3.10
<i>Iodine value:</i>	80 to 90,	Appendix 3.11
<i>Acid value:</i>	3 to 6,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Hanustambha (lockjaw); Kaṭistambha (restricted movement of the lumbo-sacral region); Gṛdhrasī (Sciatica); Khañja (limp); Kaubja (hump/kyphosis); Paṅgutva (paraplegia); Vātaśleṣmaroga (diseases due to Vāta and Kapha Doṣa); Ardita (facial palsy); Pṛṣṭhastambha (stiffness of back); Śiro-grīvā-stambha (stiffness of head and neck)

Dose:

External application for Abhyaṅga

TUVARAKA TAILA

(AFI Part I, 8:20)

Definition:

Tuvaraka Taila is a medicated oil preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Tuvaraka taila (API)	<i>Hydnocarpus laurifolia</i>	Oil	768 g
2	Khadira API	<i>Acacia catechu</i>	Ht. Wd.	576 g
3	Jala API for decoction reduced to	Potable Water	–	9.2160 l 2.304 l
4	Tuvaraka API - Kalka	<i>Hydnocarpus laurifolia</i>	Sd.	128 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Wash, clean, dry the ingredient number 2 of the formulation composition, powder separately and pass through 355 µm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Wash, clean, dry the ingredient number 4 (*kalka dravya*) of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85) to obtain fine powder.
- Transfer the powdered ingredient to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *Tuvaraka taila* in a stainless steel vessel and heat it mildly.
- Add increments of *kalka*, stir thoroughly while adding *kvātha*.
- Heat for 3 h with constant stirring, maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Next day, add *Godugdha* and continue the process of heating. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, dark reddish in colour

Identification:

Thin Layer Chromatography:

Shake 5 ml of formulation with 10 ml of *methanol* for 30 min. Allow the two layers to separate and draw the methanolic layer. Filter and carry out the thin layer chromatography. Apply 10 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: glacial acetic acid: methanol* (8: 1.5: 0.3: 0.3) as mobile phase. Spray the plate with 10 per cent *methanolic sulphuric acid* followed by heating at 105⁰ for about 10 min. Examine the plate under ultraviolet light (366 nm). It shows major spots at R_f 0.39 (blue), 0.58 (white) and 0.70 (blue).

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.4635 to 1.4748,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.831 to 0.881,	Appendix 3.2
<i>Saponification value:</i>	190 to 200,	Appendix 3.10
<i>Iodine value:</i>	75 to 85,	Appendix 3.11
<i>Acid value:</i>	3 to 6,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 6,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Kṣudrakuṣṭha (group of minor skin diseases); Mahakuṣṭha (group of major skin diseases); Meha (excessive flow of urine)

Dose:

10 to 20 drops with cow milk or butter; used externally for abhyanga also.

Pathya:

Milk, sweet, citrus fruits, apple, banana, sweet grapes, old boiled rice, barley, wheat bread, butter milk

Apathya:

Amla, Lavaṇa and Kaṭu rasa

YAṢṬĪMADHUKA TAILA

(AFI Part I, 8:47)

Definition:

Yaṣṭīmadhuka Taila is a medicinal preparation made with the ingredients in the Formulation Composition given below with Tila taila as the basic ingredient.

Formulation Composition:

1	Yaṣṭīmadhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	96 g
2	Kṣīra (Godughda API)	Cow milk	–	3.072 l
3	Dhātrīphala (Āmalakī API) - Svarasa	<i>Emblica officinalis</i>	P.	3.072 l
4	Taila (Tila Taila API)	Sesame oil	–	768 g

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Tila taila* to prepare *mūrcchita Tila taila* (Appendix 6.2.9.4).
- Wash, clean, dry the *Āmalakī*, powder separately and pass through 355 μm I. S. sieve (sieve number 44).
- To this add eight times water and reduce to one fourth of its total volume by heating on moderate flame.
- Filter while hot through *muslin cloth* to obtain *Āmalakī rasa* and keep aside.
- Strain the *Godughda* through *muslin cloth*.
- Wash, clean, dry the ingredient number 1 of the formulation composition, powder it and pass through 180 μm I. S. sieve (sieve number 85) to obtain fine powder (*kalka dravya*).
- Transfer the powdered ingredient to wet grinder and grind with sufficient quantity of water to prepare homogeneous blend (*kalka*).
- Take *mūrcchita Tila taila* in a stainless steel vessel and heat it.
- Add increments of *kalka*, stir thoroughly while adding the *Āmalakī rasa* and *Godughda*.
- Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.
- Continue the process of heating intermittently over a period of three days. Constantly check the *kalka* by rolling between the fingers.
- Stop the heating when the *kalka* easily rolls into a *varti* without sticking (*madhyama pāka lakṣaṇa*) to the fingers and at the appearance of froth (*phenodgama*) over the oil. Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture.
- Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.
- Store it in glass containers and pack them air-tight to protect from light and moisture.

Description:

Medicated oil, reddish yellow in colour with a pleasant odour

Identification:

Thin layer chromatography

a) Shake 0.5 ml of formulation in 10 ml of *methanol* and keep the mixture for 12 h. Filter and carry out thin layer chromatography. Apply 10 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: methanol: glacial acetic acid* (8: 1: 0.5: 0.2) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and observe under ultraviolet light (366 nm). It shows major spots at R_f 0.50, 0.55 (both light blue) and 0.65 (fluorescent).

b) Extract 20 ml of formulation with 50 ml of *acetone*, add 2 ml of dilute *nitric acid*, shaking occasionally for 2 h and filter. Concentrate the filtrate at 60⁰ to 20 ml on water bath. Add dilute ammonia and heat the solution till it becomes clear. Take 1 ml from this solution and extract it in the mixture of *chloroform: methanol* (1: 1). Dissolve 1 mg of *glycyrrhizic acid* in 10 ml mixture of *chloroform: methanol* (1: 1). Apply 5 µl each of the sample extract and reference solution on TLC plate. Develop the plate to a distance of 8 cm using *chloroform: methanol: glacial acetic acid: water* (6: 3: 0.5: 0.5) as mobile phase. After development, allow the plate to dry in air and observe under ultraviolet light (254 nm). The plate shows major spots of R_f 0.26, 0.31 (both black corresponding to *Glycyrrhizic acid*).

Physicochemical parameters:

<i>Refractive index at 40⁰:</i>	1.447 to 1.474,	Appendix 3.1
<i>Specific gravity at 40⁰:</i>	0.899 to 0.925,	Appendix 3.2
<i>Saponification value:</i>	192.0 to 197,	Appendix 3.10
<i>Iodine value:</i>	106 to 110,	Appendix 3.11
<i>Acid value:</i>	Not more than 1.0,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 1,	Appendix 3.13

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6
<i>Mineral oil:</i>	Absent, Appendix 3.15

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Palita (premature greying of hair); Keśapatana (hair-fall); Śmaśrupatana (hair-fall of beard and moustache)

Dose:

For external application on scalp, face and whole body

VAṬĪ AND GUṬIKĀ

General Description:

Medicines prepared in the form of pills are known as *Vaṭī* and *Guṭikās*. Such pills are small, spherical, solid dosage forms, containing one or more drugs of plant, animal or mineral origin. Pills form a convenient way of administering bitter or unpleasant substances as medicines in a pre-defined dose. In earlier days, pills were made extemporaneously by the Vaidya-Pharmacist in the following manner. The drugs of plant origin are dried and made into fine powders, separately. The minerals are made into *Bhasma* or *Sindūra*, unless otherwise mentioned. In case where *Pārada* and *Gandhaka* are mentioned, *Kajjalī* is made first and other drugs are added, one by one, according to the formula. These are put into a *Khalva* and ground to a soft paste with the prescribed fluids. When more than one liquid is mentioned for grinding, they are used in succession. *Sugandha Dravyas*, like *Kastūrī*, *Karpūra*, which are included in the formula, are added. When the mass is properly ground and kneaded to a bolus, it is in a condition to be made into pills. In cases where sugar or jaggery (*Guḍa*) is mentioned, *Pāka* of these should be made on mild fire and removed from the oven. The powders of the ingredients are added to the *Pāka* and briskly mixed. When still warm, *Vaṭī/Guṭikās* should be rolled and dried in shade. The criterion to determine the final stage of the formulation before making pills is that it should not stick to the fingers when rolled. In contemporary times, pill making machine may be recommended for turning out larger batches by the pharmacists. In this method, the prepared mass may be expelled in the form of cylindrical threads of required size, which are cut at regular intervals to give units of a pre set size. These are rolled to a spherical form by hand and dried in shade or in sun as specified in the texts.

Characteristics and Preservation

Pills made of plant drugs when kept in air tight containers can be used for two years. Pills containing minerals can be used for an indefinite period.

Pills and *Vaṭīs* should not lose their original colour, smell, taste and form. When sugar, salt or *Kṣāra* is an ingredient, the pills should be kept away from moisture.

ARKA VAṬĪ

(AFI Part II, 10:2)

Definition:

Arka Vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Sauvarcala (Sauvarcala Lavaṇa (API))	Black salt	—	1 part
2	Sādara (Narasāra API) - Śuddha	Sal ammoniac	—	1 part
3	Arkapuṣpa (Arka (API))	<i>Calotropis procera</i>	Fl. Bd.	1 part
4	Marica API	<i>Piper nigrum</i>	Fr.	1 part
5	Jala API	Potable Water	—	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Narasāra* to get *Narasāra - Śuddha*. (Appendix 6.2.8.19)
- Roast coarsely powdered *Sauvarcala Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder and pass through 180 μm I. S. sieve(sieve number 85).
- Wash, clean, dry the ingredients numbered 2 and 3 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve(sieve number 85).
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *Jala* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Spherical pills, cream coloured, with characteristic odour and taste of black salt

Identification:

Microscopy:

Take about 10 g of the formulation, powder and wash thoroughly in water, till the added salts are totally removed. Filter without loss of powder. Take a small quantity of the washed powder, add a little chloral hydrate solution and warm; wash to remove chloral hydrate and mount in 50 per cent glycerin. Take another small portion of this washed powder and mount in iodine water. Observe the following characters in different mounts:

Trichomes, epidermal cells, fibrous layer from staminal corona, rosettes of calcium oxalate, xylem vessels with spiral thickenings, cortical cells of the pedicel, fragments of pollinium, thickwalled cells, pollen grains (**Arka Puṣpa**); polygonal hypodermal cells interspersed with group of stone cells, isodiametric stone cells with thick walls, fibres, polygonal perisperm cells with starch grains and oil globules (**Marica**).

Thin layer chromatography:

Extract 1 g of sample in 10 ml of *methanol* under reflux on a water-bath for about 30 min. Filter the extract and carry out the thin layer chromatography. Apply 5 µl of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: methanol: glacial acetic acid* (8: 0.6: 1: 0.2) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.25, 0.31, 0.66 (all faint blue) 0.53 (black) and 0.60, 0.79 (both faint green) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 55 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 5.0 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 10 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 27 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 8 per cent,	Appendix 2.2.10
<i>pH (1 % aqueous solution):</i>	5 to 6.,	Appendix 3.3

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Kaphaja Agnimāndya (digestion impairment due to Kapha Doṣa)

Dose:

500 mg - 1 g per day in divided doses with warm water

CITRAKĀDI GUṬIKĀ

(AFI Part I, 12:11)

Definition:

Citrakādi Guṭikā is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Citraka API	<i>Plumbago zeylanica</i>	Rt.	1 part
2	Pippalīmūla (Pippalī API)	<i>Piper longum</i>	St.	1 part
3	Dvikṣāra			
	a. Yavakṣāra (API)	<i>Hordeum vulgare</i>	Water soluble ash of Pl.	1 part
	b. Sarjikṣāra (Svarjīkṣāra (API))	Crude alkaline earth	—	1 part
4	Pañcalavaṇa			
	a. Sauvarcala Lavaṇa (API)	Black salt	—	1 part
	b. Saindhava Lavaṇa (API)	Rock salt	—	1 part
	c. Viḍa Lavaṇa			1 part
	d. Sāmudra Lavaṇa API	Sea salt	—	1 part
	e. Audbhida Lavaṇa			1 part
5	Vyoṣa			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	1 part
	b. Marica API	<i>Piper nigrum</i>	Fr.	1 part
	c. Pippalī API	<i>Piper longum</i>	Fr.	1 part
6	Hiṅgu API - Śuddha	<i>Ferula foetida</i>	Exd.	1 part
7	Ajamodā API	<i>Apium leptophyllum</i>	Fr.	1 part
8	Cavya API	<i>Piper chaba</i>	St.	1 part
9	Mātuluṅga* API - rasa	<i>Citrus aurantifolia</i>	Fr. Juice	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Hiṅgu* to get *Hiṅgu -Śuddha*. (Appendix 6.2.8.15).
- Wash, clean, dry the ingredients number 1, 2, 5, 7 and 8 of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve(sieve number 85).
- Roast coarsely powdered ingredients numbered 4 of the Formulation Composition in a stainless steel pan on low flame till free from moisture, powder separately and pass through 180 µm I. S. sieve(sieve number 85).
- Roast ingredient number 2 of the formulation composition in a stainless steel pan on low flame till free from moisture.

* Dāḍima (seed) rasa may also be alternatively used.

- Wash and clean fresh *Mātuluṅga* fruits, cut into halves, squeeze and strain the juice through *muslin cloth* to obtain *Mātuluṅga svarasa*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *svarasa* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Light brown coloured pills with characteristic asafoetida odour and bitter taste

Identification:

Microscopy:

Take about five *Vaṭīs*, crush, wash in water repeatedly to get rid of salts; collect the residue and dry; stain a small portion in *iodine* and mount in *glycerin* (80 per cent), clear a portion in *chloral hydrate*, wash in water and mount in *glycerin* (80 per cent); clear another portion in 2 per cent *potassium hydroxide* solution, wash in water and mount in *glycerin*; observe the following characters:

Fragments of yellowish brown vittae (**Ajamodā**); a few needle and spindle shaped fibers with thick walls and narrow lumen, and isolated, circular, pitted stone cells (**Cavya**); group of isodiameric or slightly elongated stone cells with moderately thickened walls, interspersed with thin walled polygonal parenchyma cells (**Marica**), group of elongated, spindle shaped, wide lumened lignified stone cells often associated with spiral vessel units (**Pippalī**); abundant elliptic, oblong starch grains upto 60 microns in length with hilum nearer the narrower, beaked end (**Śuṅṭhī**); simple and compound starch grains having 2-7 components, round to oval with central hilum appearing like a point (**Pippalīmūla**); narrow long reticulately pitted tracheids and very long lignified pitted fibres (**Citraka**).

Thin layer chromatography:

Extract 5 g of the powdered formulation with 70 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter the extract and carry out thin layer chromatography. Apply 20 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *n-hexane: acetone* (7.5: 2.5) as mobile phase. After development, allow to dry in a current of cold air and examine under ultraviolet light (254 nm). It shows spots at R_f 0.17, 0.22 and 0.38 (all black). Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ C for about 10 min. The plate shows major spots at R_f 0.28 (black), 0.39 (red) and 0.63, 0.69 (both pink) in visible light.

Physico-chemical parameters:

Total Ash: Not more than 51 per cent, Appendix 2.2.3

<i>Acid-insoluble ash:</i>	Not more than 2 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 8 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 56 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 10 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	3.5 to 4.5,	Appendix 3.3

Assay:

Contains not less than 0.10 per cent of *piperine* when assayed by the following method.

Dissolve 1.0 mg of *piperine* in a mixture of *methanol: chloroform* (1: 1) and make up the volume to 10 ml in a volumetric flask. Apply 1, 2, 6, 10, 14, 18 µl of solution on TLC plate and develop the plate to 8 cm using *n-hexane: acetone* (7: 3) as mobile phase. After development, dry the plate in a current of hot air and scan in the TLC scanner at a wavelength of 338 nm. Note the peak area and prepare the calibration curve by plotting peak area vs concentration of piperine.

Extract accurately weighed 2g powder of vati in 75 ml of alcohol in Soxhlet apparatus for 6 h. Filter the extract while hot and dry completely and weigh. Take 100 mg in volumetric flask and dissolve in a mixture of methanol-chloroform (1:1) and make up the volume to 25 ml. Apply 25 µl of the test solutions on TLC plate. Develop, dry and scan the plate as described in the proceeding paragraph for calibration curve of piperine. Record the area under the curve for a peak corresponding to piperine in the test solution. Calculate the amount of piperine in the test solution from the calibration curve of piperine.

<i>Sodium:</i>	Not less than 12.0 per cent w/w,	Appendix 5.10.
<i>Potassium:</i>	Not less than 5.0 per cent w/w,	Appendix 5.10.
<i>Chloride:</i>	Not less than 21.00 per cent w/w,	Appendix 5.11.

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Agnimāndya (digestive impairment); Āmadoṣa (products of impaired digestion and metabolism); Grahaṇī (malabsorption syndrome)

Dose:

500 mg - 1 g per day in divided doses with warm water/ Butter milk

ELĀDI GUṬIKĀ

(AFI Part I, 12:3.)

Definition:

Elādi Guṭikā is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Elā (Sūkṣmailā API)	<i>Elettaria cardamomum</i>	Sd.	6 g
2	Patra (Tvakpatra API)	<i>Cinamomum tamala</i>	Lf.	6 g
3	Tvak API	<i>Cinamomum verum</i> (= <i>C. zeylanicum</i>)	St. Bk.	6 g
4	Pippalī API	<i>Piper longum</i>	Fr.	24 g
5	Sitā (Śarkarā API)	Sugar candy	—	48 g
6	Madhuka (Yaṣṭī API)	<i>Glycyrrhiza glabra</i>	Rt.	48 g
7	Kharjūra API	<i>Phoenix sylvestris</i>	P.	48 g
8	Mṛdvīkā (Drākṣā API)	<i>Vitis vinifera</i>	Dr. Fr.	48 g
9	Madhu API	Honey	—	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Wash, clean, dry the ingredients numbered 1 to 4 and 6 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve(sieve number 85).
- Wash, clean the ingredients numbered 7 and 8 of the Formulation Composition and grind separately to prepare soft pulp.
- Roast coarsely powdered *Sitā* in a stainless steel pan on low flame till free from moisture, powder and pass through 180 μm I. S. sieve(sieve number 85).
- Add powdered *Sitā* to the soft pulp and grind well.
- Add powdered ingredients numbered 1 to 4 and 6 of the Formulation Composition to the pulp and triturate with *Madhu* to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

A soft spherical pill, blackish brown in colour, odour cinnamon like with a sweet and pungent taste

Identification:

Microscopy:

Take about 10 *Vaṭīś*, keep in lukewarm water for 15 minutes, centrifuge at 2000 rpm for five minutes and decant the supernatant without loss of sediment. Take a small quantity of sediment and mount in 50 per cent *glycerin*, mount an other small quantity of sediment in *Iodine* and 0.001 per cent *saffranin water* respectively. Observe the following characters in different mounts.

Epidermal cells with oil cells, sclerenchymatous cells from testa (**Sūkṣmailā**), sclereids with inner walls thickened, short, very thick fibres (**Tvak**); lower epidermis with stomata, fragments of spiral vessels, short trichomes, thick-walled cells of upper epidermis, schizogenous mucilage cells (**Tvakpatra**); spindle shaped and wide lumened sclereids, perisperm cells in abundance (**Pippalī**); calcium oxalate crystals, fragments of large vessels, lignified xylem parenchyma, groups of fibres with crystal sheath, elongated fibres (**Yaṣṭī**); stone cells, vessels with spiral thickening, loose fibrous mass (**Kharjūra**); parenchyma cells with clusters of raphides and spherical pigmented cells, medullary rays, xylem vessels with spiral thickening (**Drākṣā**).

Thin Layer Chromatography:

Extract 1 g of formulation in 10 ml of *methanol* under reflux for about 30 min. Filter the extract and carry out the thin layer chromatography. Apply 4 μ l of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: methanol: glacial acetic acid* (8:1:0.7:0.2) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.25, 0.31, 0.49, 0.51, 0.66 (all faint blue) 0.53 (black), 0.56 (blue) and 0.60, 0.79 (both faint green) in visible light.

Extract 1 g of formulation in 10 ml of *methanol* under reflux for about 2 h. Filter the extract and carry out the thin layer chromatography. Apply 5 μ l of the extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate*: (7:3) as mobile phase. After development, allow the plate to dry in air. Spray the plate in *Liebermann Burchard reagent* followed by heating at 105⁰ for about 10 min and examine the plate under ultraviolet light (366nm). It shows major spots at R_f 0.11, 0.19, 0.28 (all fluorescent blue), 0.16 (faint pink), 0.24 (fluorescent green), 0.32 (black), 0.37 (light blue), 0.49 (fluorescent green, corresponding to piperine), 0.66 (faint yellow).

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 3.0 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 1.0 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 40 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 48 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 14 per cent,	Appendix 2.2.10
<i>pH (1 % aqueous solution):</i>	5 to 6,	Appendix 3.3

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Chardi (emesis); Hikkā (hiccup); Kāsa (cough); Śvāsa (asthma); Bhrama (vertigo); Mūrchā (syncope); Raktapitta (bleeding disorder); Raktaniṣṭhīvana (haemoptysis); Jvara (fever); Mada (intoxication); Tṛṣṇā (polydipsia); Aruci (anorexia); Pārśvaśūla (pleurodynia); Śoṣa (cachexia); Plīhāroga (splenic disease); Āmavāta (rheumatism); Svarabheda (hoarseness of voice); Kṣatakṣaya (emaciation due to injury); Śukrakṣaya (oligospermia)

Dose:

2- 4 g per day in divided doses with honey

LAŚUNĀDI VAṬĪ

(AFI Part I, 12:27)

Definition:

Laśunādi vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Laśuna (Rasona API)	<i>Allium sativum</i>	Bl.	1 part
2	Jīraka (Śveta Jīraka API)	<i>Cuminum cyminum</i>	Fr.	1 part
3	Saindhava (Saindhava Lavaṇa (API))	Rock salt	—	1 part
4	Gandhaka API-Śuddha	Sulphur	—	1 part
5	Trikaṭu			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	1 part
	b. Marica API	<i>Piper nigrum</i>	Fr.	1 part
	c. Pippalī API	<i>Piper longum</i>	Fr.	1 part
6	Rāmaṭha (Hiṅgu API) - Śuddha	<i>Ferula foetida</i>	Exd.	1 part
7	Nimbu (Nimbū API) - rasa	<i>Citrus limon</i>	Fr.	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Gandhaka* to get *Gandhaka - Śuddha*. (Appendix 6.2.8.3).
- Treat *Hiṅgu* to get *Hiṅgu - Śuddha*. (Appendix 6.2.8.15).
- Wash, clean, dry the ingredients numbered 2 and 5 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve(sieve number 85).
- Roast coarsely powdered *Saindhava Lavaṇa* in a stainless steel pan on low flame till free from moisture, powder and pass through 180 μm I. S. sieve(sieve number 85).
- Clean *Rasona* and grind to prepare fine paste.
- Wash and clean fresh *Nimbū* fruits, cut into halves, squeeze and strain the juice through *muslin cloth* to obtain *Nimbū svarasa*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *svrasa* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Brown coloured pills with characteristic garlic odour and saline taste

Identification:

Microscopy:

Take about 5 or 6 *Vatīś* and crush to a powder; add *chloroform* and stir thoroughly; let stand for some time to allow the heavier sulphur to settle down and the lighter plant debris to remain on top; pour the supernatant with the plant debris in another container, and repeat the process with *chloroform*; remove supernatant with plant debris to a dish, and allow chloroform to evaporate; wash the debris in alcohol once, followed by washing in water 2 or 3 times, to remove the salt; dry the debris and prepare mounts for microscopy by staining a small portion in *iodine*, and clearing another portion in *chloral hydrate*. Observe the following characteristics in the mounts.

A few fragments of thin walled elongated parenchyma cells showing stomata with spiral vessels and cells containing rhomboid crystals of calcium oxalate (**Rasona**); group of elongated stone cells from the mesocarpic layer and multicellular, multiseriate trichome (**Śveta Jīraka**); parenchyma cells, rounded to oval with small intercellular spaces, with a few cells loosely packed with cells filled with oleoresin, abundant starch granules, individual grain elliptic oblong with hilum nearer the narrower, beaked end (**Śuṅṭhī**); group of isodiameric or slightly elongated stone cells with moderately thickened walls, interspersed with thin walled polygonal parenchyma cells from hypodermis (**Marica**); some group of elongated, spindle shaped, wide lumened lignified stone cells, often associated with narrow groups of spiral vessels (**Pippalī**). In addition, abundant perisperm cells, packed with minute starch grains, characteristic of **Pippalī** and **Marica**, and loose oval starch grains upto 60 microns long from **Śuṅṭhī** are also observed.

Thin layer chromatography:

Extract 5 g of the powdered sample with 70 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter and carry out thin layer chromatography of the filtrate.

a) Apply 20.0 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *n-hexane: acetone* (7.6: 2.4) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light (254 nm). It shows spots at R_f 0.75, 0.39, 0.22 and 0.17 (all black). Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105° for about 10 min. The plate shows major spots at R_f 0.15 (violet), 0.21 (yellow), 0.29, 0.77 (both black) and 0.37, 0.44 (both purple) in visible light.

b) Apply 20.0 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *n-butanol: isopropanol: acetic acid: water* (3.5: 1.5: 1.5: 1.5) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ninhydrin reagent* followed by heating at 105° for about 10 min. The plate shows major spots at R_f 0.30 (reddish yellow) and 0.34 (red) in visible light.

Chemical tests:

Test for Sulphur:

Extract 1.0 g of the sample with 10 ml of *carbon disulphide*. Filter the carbon disulphide solution and evaporate the solvent. The residue complies with the *Test for sulphur* (Appendix 5.13.11)

Dissolve 1 g of sample in 10 ml of deionised water and filter. The filtrate complies with the *Test for Chlorides* (Appendix 5.13.6).

Complies with the *Test for Hiṅgu* (Appendix 5.14.13)

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 17 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 5 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 14 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 23 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 8 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	3 to 4,	Appendix 3.3

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Visūcikā (gastro-enteritis with piercing pain); Ajīrṇa (dyspepsia); Atisāra (diarrhoea)

Dose:

500 mg - 1 g per day in divided doses with warm water

LAVAṄGĀDI VAṬĪ

(AFI Part I, 12:26.)

Definition:

Lavaṅgādi Vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Lavaṅga API	<i>Syzigium aromaticum</i>	Fl. Bd.	1 part
2	Marica API	<i>Piper nigrum</i>	Fr.	1 part
3	Akṣaphalatvak (Bibhītaka API)	<i>Terminalia belerica</i>	P.	1 part
4	Khadirasāra (Khadira API)	<i>Acacia catechu</i>	Wd. Ext.	3 parts
5	Babbūlavṛkṣaja (Babbūla API) Kaṣāya	<i>Acacia arabica</i>	St. Bk.	QS

Method of Preparation:

- Take all ingredients of pharmacopeial quality.
- Wash, clean, dry the ingredients numbered 1 to 3 of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve (sieve number 85).
- Powder separately the ingredient number 4 of the Formulation Composition and pass through 180 µm I. S. sieve.
- Wash, clean, dry the ingredient number 5 of the Formulation Composition, powder separately and pass through 355 µm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add four times water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *kvātha* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Spherical pill, blackish brown in colour with a pleasant odour, taste bitter and astringent

Identification:

Microscopy:

Take about 10 *Vaṭīs*, powder and wash thoroughly in water. Filter without loss of powder. Take a small quantity of the washed powder, add a little *chloral hydrate* solution, warm and wash to

remove *chloral hydrate*. Take a small quantity of residue and mount in 50 per cent *glycerin*, mount another small quantity of residue in *iodine* and in 0.001 per cent *saffranin water* separately. Observe the following characters in different mounts.

Polygonal perisperm cells with starch grains and oil globules, polygonal hypodermal cells, pericarp, elongated and isodiametric stone cells, fibres (**Marica**); short trichomes with a basal cell, stone cells (**Bibhītaka**); tricolpate pollen grains, fibres, spiral tracheids, calcium oxalate crystals in rosette aggregates, schizolysigenous oil cavity embedded in parenchyma, (**Lavaṅga**).

Thin Layer Chromatography:

Extract 1 g of powdered sample with 10 ml of *n-hexane* by keeping the mixture for 2 h with occasional shakings, filter the extract and evaporate the filtrate on water bath at 60⁰. Dissolve the residue in 10 ml of methanol, filter and carry out the thin layer chromatography. Apply 10 µl on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: glacial acetic acid* (8: 2: 0.2) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *anisaldehyde sulphuric acid* followed by heating at 105⁰ for about 10 min. It shows major spots at R_f 0.20 (pink), 0.24 (faint brown), 0.29 (light blue corresponding to piperine), 0.35, 0.43 (both black), 0.54 (brown), 0.74 (dark brown corresponding to eugenol) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 5.0 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 1.0 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 26 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 22 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 14 per cent,	Appendix 2.2.10
<i>pH (1% aqueous solution):</i>	4 to 5,	Appendix 3.3

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Kāsa (cough); Śvāsa (asthma)

Dose:

500 mg - 1 g per day in divided doses (to be kept in mouth till it dissolves)

PLĪHĀRI VAṬIKĀ

(AFI Part I, 12:17)

Definition:

Plīhāri Vaṭikā is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Sahāsāra (Kumārī (API))	<i>Aloe barbadensis</i>	Fresh Lf. P.*	1 part
2	Abhra (Abhraka Bhasma (API))	Calcined Mica	–	1 part
3	Kāsīsa (Kāśīśa API) - Śuddha	Green vitreol	–	1 part
4	Laśuna (Rasona API)	<i>Allium sativum</i>	Bl.	1 part
5	Droṇapūṣṭī API - Svarasa	<i>Leucas cephalotes</i>	Juice of fresh Pl.	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Kāśīśa* to get *Kāśīśa - Śuddha* (Appendix 6.2.8.17).
- Treat *Abhraka* to get *Abhraka bhasma*.
- Clean *Rasona* and grind to prepare fine paste. .
- Take fresh *Droṇapūṣṭī* and wash thoroughly with water. Grind and filter through *muslin cloth* to obtain *svarasa*.
- Wash and clean fresh *Kumārī* and separate the pulp to obtain *Kumārī svarasa*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *svarasa* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Reddish brown coloured pills with characteristic garlic odour and bitter taste

Identification:

Microscopy:

Take about five *Vaṭīs*, crush, wash in slow running water on a 150 μm I. S. sieve to allow minerals to be washed away. Collect the material left on the sieve, without loss of material; take a

* Kanyāsāra may alternatively be used.

small amount, clear in *chloral hydrate* solution, wash in water and mount in *glycerin* (80 per cent), observe the following characters:

A few fragments of thin walled elongated parenchyma cells with stomata, spiral vessels and parenchymatous cells containing rhomboid crystals of calcium oxalate (**Rasona**); isolated acicular needles of calcium oxalate crystals (**Sahāsāra**).

Thin Layer Chromatography:

Extract 5 g of the powdered sample with 70 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter and carry out thin layer chromatography. Apply 20 µl of the filtrate on TLC plate. Develop the plate to a distance of 8 cm using *n-butanol: isopropanol: acetic acid: water* (3: 1: 1: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.41 (deep green), 0.56 (light green), 0.67 and 0.90 (pink) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 60 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 14 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 8 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 45 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 12 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	3 to 4,	Appendix 3.3

Assay:

Not less than 0.25 per cent of aloin when assayed by the following method.

Dissolve 2.5 mg of aloin in mixture of *methanol: chloroform* (1: 1) and make up the volume to 25 ml in another volumetric flask. Apply 2, 4, 6, 8, 10, 12 µl of solution on TLC plate and develop the plate to 8 cm using *ethyl acetate: methanol: water* (8: 1.35: 1) as mobile phase. After development dry the plate in a current of hot air and scan in the TLC scanner at a wavelength of 366 nm. Note the peak area and prepare the calibration curve by plotting peak area vs concentration of aloin.

Extract accurately weighed about 7 g of powdered *Vaṭīś* in 100 ml of *alcohol* in a Soxhlet apparatus for 6 h. Filter the extract while hot and dry completely and weigh. Take 25 mg of extract in volumetric flask and dissolve in a mixture of *methanol: chloroform* (1: 1) and make up the volume to 25 ml. Apply 7 µl of the test solutions on TLC plate. Develop, dry and scan the plate as described in the proceeding paragraph for calibration curve of aloin. Record the area under the curve for a peak corresponding to aloin in the test solution. Calculate the amount of aloin in the test solution from the calibration curve of aloin.

<i>Iron:</i>	Not less than 18.5 per cent w/w,	Appendix 5.6
<i>Aluminum:</i>	Not less than 1.5 per cent w/w,	Appendix 5.2
<i>Magnesium:</i>	Not less than 0.9 per cent w/w,	Appendix 5.7
<i>Potassium:</i>	Not less than 1.9 per cent w/w,	Appendix 5.10

Silica:

Not less than 9.5 per cent w/w,

Appendix 5.9

Other requirements:

Microbial limits:

Complies with Appendix 2.4

Aflatoxins:

Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Plihā-yakṛdroga (spleno-hepato disease); Gulma (abdominal lump); Agnimāndya (digestive impairment); Śoṭha (inflammation); Kāsa (cough); Śvāsa (asthma); Tṛṣṇā (thirst); Kampa (tremor); Dāha (burning sensation); Chardi (emesis); Bhrama (vertigo)

Dose:

500 mg per day in divided doses

PRABHĀKARA VAṬĪ

(AFI Part I, 12:15)

Definition:

Prabhākara Vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Mākṣika (Svarṇamākṣika Bhasma (API))	Calcined Chalcopyrite	—	1 part
2	Lauha (Lauha Bhasma (API))	Calcined Lauha	—	1 part
3	Abhra (Abhraka Bhasma (API))	Calcined Abhraka	—	1 part
4	Tugākṣīrī (Vaṃśalocana (API))	Bamboo manna	S. C.	1 part
5	Śilājatu (API) - Śuddha			1 part
6	Pārthavāri (Arjuna API) - Kvātha	<i>Terminalia arjuna</i>	St. Bk.	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Svarṇamākṣika* to get *Svarṇamākṣika bhasma*.
- Treat *Lauha* to get *Lauha bhasma*.
- Treat *Abhraka* to get *Abhraka bhasma*.
- Treat *Śilājatu* to get *Śilājatu- Śuddha* (Appendix 6.2.8.13)
- Wash, clean, dry the ingredient number 6 of the Formulation Composition, powder separately and pass through 355 μm I. S. sieve (sieve number 44) (*kvātha dravya*).
- Add four times water for decoction to the *kvātha dravya* and soak for 4 h, heat and reduce the volume to one-fourth. Filter through *muslin cloth* to obtain *kvātha*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *kvātha* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Reddish brown pills with a faint odour and acrid taste

Identification:

Microscopy:

Take about 5 *Vaṭīs*, triturate with water in a mortar, collect in a beaker stir with glass rod, swirl gently and pour off the supernatant in to another beaker; repeat this process 3 to 4 times, so that as

much of the bhasmas and minerals components can be floated and removed. Add *strong sulphuric acid*, stir, and remove sulphuric acid by pouring it off. Wash the residue, and mount a small quantity in water and another in *glycerin*; observe the characters:

Angular, flat, structure less plates, appear in good relief in lower powers in water mounts, but only faintly visible in glycerin mounts; it is invisible between crossed polars in the polarising microscope in both mounts (**Vamśalocana**).

Thin layer chromatography:

Extract 5 g of the powdered formulation with 50 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter and carry out thin layer chromatography. Apply 20.0 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *chloroform: methanol* (9: 1) as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light (366 nm). It showed spots at 0.63, 0.54, 0.45, 0.40 (all faint blue) and 0.29 (dark blue). Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.26 (red), 0.31, 0.41 (both brown), and 0.37 (black) in visible light.

Physico-chemical Parameters:

<i>Total Ash:</i>	Not more than 70 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 3 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 7 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 14 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 13 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	7.0 to 8.5,	Appendix 3.3

Assay:

<i>Iron:</i>	Not less than 21.5 per cent w/w,	Appendix 5.6
<i>Calcium:</i>	Not less than 3.0 per cent w/w,	Appendix 5.4
<i>Copper:</i>	Not less than 0.05 per cent w/w	Appendix 5.5
<i>Silicates:</i>	Not less than 36.5 per cent w/w	Appendix 5.9
<i>Sulphur:</i>	Not less than 6.0 per cent w/w	Appendix 5.12

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Hṛdroga (heart disease)

Dose:

250 mg - 500 mg per day in divided doses with water/ Arjuna kvātha/milk

RAJAḤPRAVARTINĪ VAṬĪ

(AFI Part I, 12:25)

Definition:

Rajaḥpravartinī Vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Kanyāsāra (API)	<i>Aloe barbadensis</i>	Dry Ext.	1 part
2	Kāsīsa (Kaśīśa API) - Śuddha	Green vitreol	–	1 part
3	Rāmaṭha (Hiṅgu API) - Śuddha	<i>Ferula foetida</i>	Exd.	1 part
4	Taṅkaṇa API - Śuddha	Borax	–	1 part
5	Kanyakā (Kumārī (API)) - Svarasa	<i>Aloe barbadensis</i>	Lf.	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Kāsīsa* to get *Kaśīśa - Śuddha*. (Appendix 6.2.8.17)
- Treat *Hiṅgu* to get *Hiṅgu - Śuddha*. (Appendix 6.2.8.15).
- Treat *Taṅkaṇa* to get *Taṅkaṇa - Śuddha*. (Appendix 6.2.8.5).
- Wash and clean fresh *Kumārī* and separate the pulp to obtain *Kumārī svarasa*.
- Dissolve the *Kanyāsāra* in *Kumārī svarasa*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *svarasa* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60° for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Blackish grey coloured pills with a characteristic asafetida odour and saline taste

Identification:

Thin layer chromatography:

Extract 5 g of the powdered sample with 50 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter and carry out thin layer chromatography.

a) Apply 20 µl of the extract on TLC plate. Develop the plate to a distance of 8cm using *n-hexane: acetone (7: 5: 2.5)* as mobile phase. After development, allow the plate to dry in air and examine under ultraviolet light (366 nm). It shows spots at R_f 0.25 (dark blue) and 0.30, 0.39

(both faint blue). Spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.25 (reddish brown), 0.34 (pale brown), 0.43 and 0.61 (brown) in visible light.

b) Apply 20 µl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *ethyl acetate: methanol: water* (10: 1.35: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *ethanolic potassium hydroxide reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.27 (reddish yellow) and 0.72 (red) in visible light.

Chemical Test:

Complies with *Tests for Hiñgu* (Appendix 5.13.13), *Borax* (Appendix 5.13.10) and *Anthraquinones* (Appendix 5.13.12)

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 36 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 26 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 3.0 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 30 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 12 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	6.5 to 7.5,	Appendix 3.3

Assay:

Not less than 0.56 per cent of aloin when assayed by the following method.

Dissolve accurately weighed about 2.5 mg of aloin in *methanol-chloroform* (1: 1) and make up the volume to 25 ml in another volumetric flask. Apply 2, 4, 6, 8, 10, 12 µl of solution on TLC plate and develop the plate to 7.5 cm using *ethyl acetate: methanol: water* (8: 1.35: 1) as mobile phase. After development dry the plate in a current of hot air and scan in the TLC scanner 3 at a wavelength of 366 nm. Note the peak area and prepare the calibration curve by plotting peak area vs concentration of aloin.

Extract accurately weighed about 7 g powder of *Vaiñis* in 100 ml of alcohol in Soxhlet apparatus for 6 h. Filter the extract while hot and dry completely and weigh. Take 25 mg in volumetric flask. Make the volume to 25 ml with *methanol-chloroform* (1: 1) and make up the volume to 25 ml apply 10 µl of the test solutions on TLC plate. Develop, dry and scan the plate as described in the proceeding paragraph for calibration curve of aloin. Record the area under the curve for a peak corresponding to aloin in the test solution. Calculate the amount of aloin in the test solution from the calibration curve of aloin.

<i>Sodium:</i>	Not less than 4.5 per cent w/w,	Appendix 5.10
<i>Iron:</i>	Not less than 3.5 per cent w/w,	Appendix 5.6

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Rajorodha (obstructed menstrual flow); Kaṣṭhārtava (dysmenorrhoea); Ārtavavedanā (dysmenorrhoea)

Dose:

500 mg - 1 g per day in divided doses with Warm water/ Tila kvātha/Kulattha kvātha

SAÑJĪVANĪ VAṬĪ

(AFI Part I, 12:35)

Definition:

Sañjīvanī Vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Viḍaṅga API	<i>Embelia ribes</i>	Fr	1 part
2	Nāgara (Śuṅṭhī API)	<i>Zingiber officinale</i>	Rz.	1 part
3	Kṛṣṇā (Pippalī API)	<i>Piper longum</i>	Fr.	1 part
4	Pathyā (Harītakī API)	<i>Terminalia chebula.</i>	P.	1 part
5	Āmala (Āmalakī API)	<i>Phyllanthus emblica</i>	P.	1 part
6	Bibhītaka API	<i>Terminalia bellirica</i>	P.	1 part
7	Vacā API	<i>Acorus calamus</i>	Rz.	1 part
8	Guḍūcī API	<i>Tinospora cordifolia</i>	St.	1 part
9	Bhallātaka API - Śuddha	<i>Semecarpus anacardium</i>	Fr.	1 part
10	Viṣa (Vatsanābha API) - Śuddha	<i>Aconitum chasmanthum</i>	Rt. Tr.	1 part
11	Gomūtra (API)	Cow urine	—	QS

Method of preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat *Bhallātaka* to get *Bhallātaka - Śuddha*. (Appendix 6.2.8.7).
- Treat *Vatsanābha* to get *Vatsanābha - Śuddha*. (Appendix 6.2.8.9).
- Wash, clean, dry the ingredients numbered 1 to 8 of the Formulation Composition, powder separately and pass through 180 μm I. S. sieve (sieve number 85).
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Powder *Bhallātaka - Śuddha* and *Vatsanābha - Śuddha* separately and pass through 180 μm I. S. sieve (sieve number 85) and add to the mixture to prepare a homogenous blend.
- Strain the *Gomūtra* through *muslin cloth*.
- Add *Gomūtra* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Black coloured pills with a pleasant odour and acrid taste

Identification:

Microscopy:

Take about 5 *Vaṭīś*, crush, take in a beaker containing *chloroform* sufficient to wash the powder two times; remove chloroform, allow to dry; wash the powder once in *alcohol* and again dry; take a small amount and wash in water and mount in *glycerin* (80 per cent); take another small portion, clear in *chloral hydrate* solution, wash in water and mount in *glycerin*; observe the following characters:

Groups of stone cells from testa with very thick walls and radiating pit canals (**Vidāṅga**); abundant thin walled parenchyma cells, rounded to oval with small intercellular spaces, with a few cells loosely packed with starch grains upto 60 microns, individual grain elliptic, oblong, with hilum nearer the narrower, beaked end and some cells filled with oleoresin (**Śuṅṭhī**); group of elongated, spindle shaped, wide lumened lignified stone cells often associated with spiral narrow vessels (**Pippalī**); polygonal epidermal cells from pericarp, where walls are slightly beaded, with thin septa dividing the cells (**Harītakī**); a few short stout unicellular thick walled trichomes with a basal epidermal cell (**Bibhītaka**); large mesocarpic parenchyma cells, with typical corner thickenings, cells of epidermal tissue with small crystals of calcium oxalate (**Āmalakī**); tissue debris showing moderately thick walled parenchymatous cells surrounding large, irregularly shaped air spaces, cells contain spherical starch grains (**Vacā**); rounded, hemispherical and irregularly ovoid starch grains with central hilum and isolated crystal fibres (**Guḍūcī**); a few large isolated or group of rectangular or angular stone cells with thin walls and wide pitted lumen (**Vatsanābha**).

Thin layer chromatography:

Extract 5 g of the powdered sample with 70 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter and carry out thin layer chromatography.

a) Apply 20.0 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *n-hexane: acetone* (7.5: 2.5) as mobile phase. After development, allow the plate to dry in air and spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows major spots at R_f 0.36, 0.41 (both dark pink), 0.48 (brown), 0.62, 0.69 (both purple) and 0.75 (violet) in visible light.

b) Apply 20 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: diethyl amine* (7: 2: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Dragendroff reagent* and dry the plate followed by dipping in 5 per cent *ethanolic sulphuric acid*. The plate shows major spots at R_f 0.49 and 0.59 (both orange) in visible light.

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 4 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 1 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 18 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 17 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 10 per cent,	Appendix 2.2.10

pH (10 % aqueous solution): 4.0 to 5.5,

Appendix 3.3

Assay:

Not less than 0.047 per cent of piperine when assayed by the following method.

Dissolve 1.0 mg of piperine in a mixture of *methanol: chloroform* (1: 1) and make up the volume to 10 ml in another volumetric flask. Apply 1, 2, 6, 10, 14, 18 µl of solution on TLC plate and develop the plate to 5.5 cm using *n-hexane: acetone* (7: 3) as mobile phase. After development, dry the plate in a current of hot air and scan in the TLC scanner at a wavelength of 338 nm. Note the peak area and prepare the calibration curve by plotting peak area vs concentration of piperine.

Extract accurately weighed 2 g powder of *Vaṭīś* in 75 ml of alcohol in Soxhlet apparatus for 6 h. Filter the extract while hot and dry completely and weigh. Take 100 mg in volumetric flask and dissolve in a mixture of *methanol: chloroform* (1: 1) and make up the volume to 25 ml. Apply 25 µl of the test solutions on TLC plate. Develop, dry and scan the plate as described in the proceeding paragraph for calibration curve of piperine. Record the area under the curve for a peak corresponding to piperine in the test solution. Calculate the amount of piperine in the test solution from the calibration curve of piperine.

Other requirements:

Microbial limits: Complies with Appendix 2.4

Aflatoxins: Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Ajīrṇa (indigestion); Gulma (abdominal lump); Visūcikā (gastroenteritis with piercing pain); Sarpadaṃśa (snake bite)

Dose:

500 mg - 1 g per day in divided doses with warm water/ Ārdraka svarasa

ŚAṆKHA VAṬĪ

(AFI Part I, 12:32)

Definition:

Śaṅkha Vaṭī is a pill preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Ciñcā API -kṣāra	<i>Tamarindus indica</i>	St. Bk.	48 g
2	Paṭuvajra (Pañca lavaṇa each in equal quantity)			48 g
	a. Saindhava Lavaṇa (API)	Rock salt	—	
	b. Sāmudra Lavaṇa API	Sea salt	—	
	c. Sauvarcala Lavaṇa (API)	Black salt	—	
	d. Viḍa Lavaṇa			
	e. Audbhida Lavaṇa			
3	Śaṅkha (Śaṅkha Bhasma (API))	Calcined Conch	—	48 g
4	Hiṅgu API - Śuddha	<i>Ferula foetida</i>	Exd.	48 g
5	Vyoṣa			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	48 g
	b. Marica API	<i>Piper nigrum</i>	Fr.	48 g
	c. Pippalī API	<i>Piper longum</i>	Fr.	48 g
6	Rasa (Pārada API) - Śuddha	Mercury	—	3 g
7	Amṛta (Vatsanābha API) - Śuddha	<i>Aconitum chasmanthum</i>	Rt. Tr.	3 g
8	Valī (Gandhaka API) - Śuddha	Sulphur	—	3 g
9	Nimbū API - Svarasa	<i>Citrus limon</i>	Fr. juice	QS

Method of Preparation:

- Take all ingredients of pharmacopoeial quality.
- Treat Śaṅkha to prepare Śaṅkha bhasma.*
- Treat Hiṅgu to get Hiṅgu - Śuddha (Appendix 6.2.8.15), Vatsanābha to get Vatsanābha - Śuddha (Appendix 6.2.8.9), Pārada to get Pārada - Śuddha (Appendix 6.2.8.20) and Gandhaka to get Gandhaka - Śuddha (Appendix 6.2.8.3).
- Triturate Pārada - Śuddha and Gandhaka - Śuddha together in a Khalvayantra to prepare Kajjalī. (Appendix 6.2.1).
- Wash, clean, dry the ingredients numbered 5 of the Formulation Composition, powder separately and pass through 180 µm I. S. sieve(sieve number 85).

* Alternative method: Make thin paste of Ciñcā kṣāra and Pañca Lavaṇa in Nimbū Svarasa. Heat and quench Śaṅkha - Śuddha repeatedly in this paste till it disintegrates to powder.

- Powder *Vatsanābha* - *Śuddha* separately and pass through 180 μm I. S. sieve(sieve number 85).
- Roast coarsely powdered *Saindhava*, *Sāmudra*, *Sauvarcala*, *Viḍa* and *Audbhida Lavaṅas* in a stainless steel pan on low flame till free from moisture, powder separately and pass through 180 μm I. S. sieve.
- Prepare *Ciñcā kṣāra* (Appendix 6.2.3) and roast in a stainless steel pan on low flame till free from moisture.
- Wash and clean fresh *Nimbū* fruits, cut into halves, squeeze and strain the juice through *muslin cloth* to obtain *Nimbū svarasa*.
- Mix all the ingredients thoroughly to prepare a homogenous blend.
- Add *svarasa* in required quantity and triturate well to prepare a bolus.
- Expel the mass through *vaṭī* machine to obtain cylindrical threads and cut the *vaṭīs* to a desired weight.
- Roll the *vaṭīs* on flat surface to round them by circular motion of palm covered with a glove or use suitable mechanical device.
- Dry the rounded *vaṭīs* in a tray-dryer at a temperature not exceeding 60⁰ for 10 to 12 h.
- Store *vaṭīs* in containers and pack them air-tight to protect from light and moisture.

Description:

Light grey pills with a characteristic asafoetida odour and salty taste.

Identification:

Microscopy:

Take about five *Vaṭīs*, crush, wash in slow running water on a 150 μm I. S. sieve to allow mineral and water soluble matter to be washed away. Collect the material on sieve, wash repeatedly in water and mount a small portion in *glycerin* (80 per cent); warm a small portion in *chloral hydrate* solution, wash and mount in *glycerin*; observe the following characters:

Groups of isodiametric or slightly elongated stone cells with moderately thickened walls, interspersed with thin walled polygonal parenchyma cells from hypodermis (**Marica**); group of elongated, spindle shaped, wide lumened lignified stone cells (**Pippalī**); a few large isolated or group of rectangular or angular stone cells with thin walls and wide pitted lumen (**Vatsanābha**); abundant thin walled parenchyma cells, rounded to oval with small intercellular spaces, with a few cells loosely packed with starch grains, individual grain elliptic, oblong, varying upto 60 microns with hilum nearer the narrower, beaked end and some cells filled with oleoresin (**Śunṭhī**); abundant loose starch grains and perisperm cells are general characters.

Thin layer chromatography:

Extract 5 g of the powdered formulation with 70 ml of *ethanol* in a Soxhlet apparatus on a water bath for 6 h, filter and carry out thin layer chromatography.

a) Apply 20.0 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *n-hexane: acetone* (7.5: 2.5) as mobile phase. After development, allow the plate to dry in air spray the plate with *anisaldehyde-sulphuric acid reagent* followed by heating at 105⁰ for about 10 min.

The plate shows major spots at R_f , 0.13 (brown), 0.19, 0.23 (both green), 0.35 (purple), 0.40 (blue), 0.58 (pink) and 0.60 (violet) in visible light.

b) Apply 20.0 μl of the extract on TLC plate. Develop the plate to a distance of 8 cm using *toluene: ethyl acetate: diethyl amine* (7: 2: 1) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *Dragendorff reagent* and dry the plate followed by dipping in 5 per cent *ethanolic sulphuric acid*. The plate shows two major spots at R_f . 0.49 and 0.59 (both orange) in visible light.

Chemical Tests:

Complies with *Tests for Hiingu* (Appendix 5.13.13) and *Sulphur* (Appendix 5.13.11)

Physico-chemical parameters:

<i>Total Ash:</i>	Not more than 43 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 6 per cent,	Appendix 2.2.4
<i>Alcohol-soluble extractive:</i>	Not less than 8 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	Not less than 53 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 12 per cent,	Appendix 2.2.10
<i>pH (10 % aqueous solution):</i>	8.0 to 9.0,	Appendix 3.3

Assay:

Not less than 0.15 per cent of piperine when assayed by the following method.

Dissolve 1.0 mg of piperine in a mixture of *methanol: chloroform* (1: 1) and make up the volume to 10 ml in another volumetric flask. Apply 1, 2, 6, 10, 14, 18 μl of solution on TLC plate and develop the plate to 5.5 cm using *n-hexane: acetone* (7: 3) as mobile phase. After development, dry the plate in a current of hot air and scan in the TLC scanner at a wavelength of 338 nm. Note the peak area and prepare the calibration curve by plotting peak area vs concentration of piperine.

Extract accurately weighed 2 g powder of *Vaṭīś* in 75 ml of alcohol in Soxhlet apparatus for 6 h. Filter the extract while hot and dry completely and weigh. Take 100 mg in volumetric flask and dissolve in a mixture of methanol-chloroform (1:1) and make up the volume to 25 ml. Apply 20 μl of the test solutions on TLC plate. Develop, dry and scan the plate as described in the preceding paragraph for calibration curve of piperine. Record the area under the curve for a peak corresponding to piperine in the test solution. Calculate the amount of piperine in the test solution from the calibration curve of piperine.

<i>Mercury:</i>	0.6-1.10 per cent w/w,	Appendix 5.8
-----------------	------------------------	--------------

Other requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflatoxins:</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protected from light and moisture.

Therapeutic uses:

Agnimāndya (digestive impairment); Grahaṇī (malabsorption syndrome); Arocaka (tastelessness); Pakṣīūla (duodenal ulcers); Kṣaya (pthisis)

Dose:

500 mg - 1 g per day in divided doses with Honey/Warm water/Butter milk

PUNARNAVĀDI MAṆḌŪRA (TABLET)

(AFI Part I, 19:1)

Definition:

Punarnavādi Maṇḍūra (Tablet) is a compressed tablet preparation made with the ingredients in the Formulation Composition given below.

Formulation Composition:

1	Punarnavā (Rakta Punarnavā API)	<i>Boerhaavia diffusa</i>	Rt.	48 g
2	Trivṛt API	<i>Ipomea turpethum</i>	Rt.	48 g
3	Vyoṣa			
	a. Śuṅṭhī API	<i>Zingiber officinale</i>	Rz.	48 g
	b. Marica API	<i>Piper nigrum</i>	Fr.	48 g
	c. Pippalī API	<i>Piper longum</i>	Fr.	48 g
4	Viḍaṅga API	<i>Embelia ribes</i>	Fr.	48 g
5	Dāru (Devadāru API)	<i>Cedrus deodara</i>	Ht. Wd.	48 g
6	Citraka API	<i>Plumbago zeylanica</i>	Rt.	48 g
7	Kuṣṭha API	<i>Saussurea lappa</i>	Rt.	48 g
8	Haridrādvaya			
	a. Haridrā API	<i>Curcuma longa</i>	Rz.	48 g
	b. Dāruharidrā API	<i>Berberis aristata</i>	St.	48 g
9	Triphalā			
	a. Harītakī API	<i>Terminalia chebula</i>	P.	48 g
	b. Āmalakī API	<i>Emblia officinalis</i>	P.	48 g
	c. Bibhītaka API	<i>Terminalia bellirica</i>	P.	48 g
10	Dantī API	<i>Baliospermum montanum</i>	Rt.	48 g
11	Cavya API	<i>Piper chaba</i>	St.	48 g
12	Kaliṅgaka (Indrayava API)	<i>Holarrhena antidysenterica</i>	Sd.	48 g
13	Pippalī API	<i>Piper longum</i>	Fr.	48 g
14	Pippalīmūla (Pippalī API)	<i>Piper longum</i>	St.	48 g
15	Musta (Mustā API)	<i>Cyperus rotundus</i>	Rz.	48 g
16	Maṇḍūra (Maṇḍūra Bhasma (API))	Calcined Maṇḍūra		1.920 kg
17	Gomūtra (API)	Cow urine		6.144 l

Method of preparation:

- Take all the ingredients of pharmacopoeial quality.
- Treat Maṇḍūra to prepare Maṇḍūra bhasma.
- Wash, clean, dry the ingredients numbered 1 to 15 of the formulation composition, powder separately and pass through 180 µm I. S. sieve(sieve number 85) to obtain fine powder and mix them all to a homogeneous mixture.
- Strain Gomūtra through muslin cloth.

- Add *Gomūtra* to *Maṇḍūra bhasma*, heat in a stainless steel container stirring continuously and observe the mixture for formation of a thick paste.
- Stop the heating and allow to cool to 50°.
- Add mixture of fine powders and mix thoroughly to prepare a homogeneous lumpy mass (dough). Allow to cool.
- Pass the dough through a granulator to obtain granules and add 0.5 per cent of talc powder as lubricant. Subject the granules to compression in a tablet punching machine.
- Store tablets in containers and pack them air-tight to protect from light and moisture.

Description:

Reddish brown coloured tablet with the characteristic odour of *Gomūtra*

Identifications:

Thin layer chromatography:

Extract 2 g of formulation with 20 ml of *alcohol* for 40⁰ for about 3 h. Filter and concentrate the extract to 5 ml and carry out thin layer chromatography. Apply 10 µl of extract on TLC plate and develop the plate to a distance of 8 cm using *toluene: ethyl acetate: acetic acid* (5: 4.5: 0.5) as mobile phase. After development, allow the plate to dry in air. Spray the plate with *anisaldehyde sulphuric acid reagent* followed by heating at 105⁰ for about 10 min. The plate shows spots at R_f 0.30 (light blue), 0.66 (yellow), 0.70 (green) and 0.76, 0.85 (both blue).

Physico-chemical parameters:

<i>Total ash:</i>	Not more than 60 per cent,	Appendix 2.2.3
<i>Acid-insoluble ash:</i>	Not more than 28 per cent,	Appendix 2.2.4
<i>Methanol-soluble extractive:</i>	15 to 17 per cent,	Appendix 2.2.7
<i>Water-soluble extractive:</i>	24 to 26 per cent,	Appendix 2.2.8
<i>Loss on drying:</i>	Not more than 8 per cent,	Appendix 2.2.10
<i>pH (10% aqueous solution):</i>	6 to 7	Appendix 3.3
<i>Disintegration time</i>	Not more than 20 min	Appendix 3.18
<i>Average weight</i>	250 (±5) mg	Appendix 3.19

Assay:

<i>Total Iron:</i>	Not less than 14 per cent w/w	Appendix 5.6
--------------------	-------------------------------	--------------

Other Requirements:

<i>Microbial limits:</i>	Complies with Appendix 2.4
<i>Aflotoxins :</i>	Complies with Appendix 2.6

Storage:

Store in a cool place in tightly closed containers, protect from light and moisture.

Important Therapeutic uses:

Pāṇḍu Roga (anaemia), Grahaṇī (malabsorption syndrome), Śoṭha (inflammation), Plīhā Roga (splenic disease), Viṣamajvara (intermittent fever), Arśa (haemorrhoids), Kuṣṭha (diseases of skin), Kṛmi (helminthiasis/worm infestation)

Dose:

1 to 2 g per day in divided doses

Anupāna:

Buttermilk, water

DRAFT

AYURVEDIC PHARMACOPOEIA OF INDIA

PART-II

VOL – III

(COMPOUND FORMULATIONS)

Appendix-1. Apparatus for Tests & Assays

1.1.	Nessler Cylinders.....	139
1.2.	Sieves.....	139
1.3.	Thermometers.....	140
1.4.	Ultraviolet Lamp (For general purposes and for chromatography work).....	140
1.5.	Volumetric Glassware	141
1.6.	Weights and Balances.....	141

Appendix-2. Tests and Determinations

1.7.	Muslin Cloth.....	141
2.1.	Microscopic identification:.....	142
2.2.	Determination of Quantitative Data:.....	145
2.2.1.	Net Content:.....	145
2.2.2.	Foreign Matter:.....	146
2.2.3.	Determination of Total Ash:	146
2.2.4.	Determination of Acid-Insoluble Ash:	146
2.2.5.	Determination of Water Soluble Ash:	146
2.2.6.	Determination of Sulphated Ash:	146
2.2.7.	Determination of Alcohol Soluble Extractive:.....	147
2.2.8.	Determination of Water Soluble Extractive:	147
2.2.9.	Determination of Ether Soluble Extractive (Fixed Oil Content):.....	147
2.2.10.	Determination of Moisture Content (Loss on Drying):.....	147
2.2.11.	Determination of Volatile Oil in Drugs	147
2.2.12.	Special Processes Used in Alkaloidal Assays:	149
2.2.13.	Thin-Layer Chromatography (TLC):.....	150
2.2.14.	Fatty oil estimation:	152
2.3.	Limit Tests:.....	152
2.3.1.	Limit Test for Arsenic	153
2.3.2.	Limit Test for Chlorides:.....	157
2.3.3.	Limit Test for Heavy metals:	158
2.3.4.	Limit Test for Iron.....	160
2.3.5.	Limit Test for Lead	161
2.3.6.	Limit Test for Sulphates:.....	163
2.3.7.	Heavy Metals by Atomic absorption spectrophotometry:	163
2.4.	Microbial Limit Tests:	168
2.4.1.	Total Aerobic Microbial Count:	178
2.4.2.	Tests for Specified Micro-organisms:	180
2.5.	Pesticide Residue:.....	184
2.5.1.	Qualitative and Quantitative Analysis of Pesticide Residues:.....	186
2.5.2.	Test for Pesticides:	188
2.5.3.	Quantitative Analysis:.....	189
2.6.	Test for Aflatoxins:.....	192

2.7.	Gas Chromatography:	192
2.8.	Test for the Absence of Methanol:	196

Appendix-3. Physical Tests and Determinations

3.1.	Refractive Index:	197
3.2.	Weight per Millilitre and Specific Gravity:.....	197
3.3.	Determination of pH Values:	198
3.4.	Determination of Melting Range and Congealing Range:	198
3.4.1.	Determination of Melting Range:	198
3.4.2.	Determination of Congealing Range:	201
3.5.	Determination of Boiling Range:.....	202
3.6.	Determination of Optical Rotation and Specific Optical Rotation:	203
3.7.	Determination of Viscosity:.....	205
3.8.	Determination of Total Solids:	206
3.9.	Solubility in Water:	206
3.10.	Determination of Saponification Value:	206
3.11.	Determination of Iodine Value:	207
3.12.	Determination of Acid Value:	208
3.13.	Determination of Peroxide Value:	208
3.14.	Determination of Unsaponifiable Matter:	209
3.15.	Detection of Mineral Oil (Holde's Test):	210
3.16.	Rancidity Test (Kreis Test):.....	210
3.17.	Determination of Alcohol Content:	210
3.18.	Disintegration test for tablets:.....	214
3.19.	Uniformity of Weight of Single Dose Preparations.....	216

Appendix-4. Regents and Solutions

Appendix-5. Assys and Chemical Tests

5.1.	Estimation of Sugars.....	250
5.2.	Determination of Aluminum:	252
5.3.	Determination of Borax:	252
5.4.	Determination of Calcium:	252
5.5.	Determination of Copper:	253
5.6.	Determination of Iron (Fe)	253
5.7.	Determination of Magnesium:.....	254
5.8.	Determination of Mercury:	254
5.9.	Determination of Silica (SiO ₂)	255
5.10.	Estimation of Sodium and Potassium by Flame Photometer:.....	255
5.11.	Determination of Sodium Chloride:	256

5.12.	Determination of Sulphur:	256
5.13.	Qualitative Reactions :	256
5.13.1	Sodium.....	256
5.13.2.	Potassium.....	257
5.13.3.	Magnesium	257
5.13.4.	Carbonates and Bicarbonates	257
5.13.5.	Sulphates.....	258
5.13.6.	Chlorides.....	258
5.13.7.	Calcium.....	258
5.13.8.	Sulphides	258
5.13.9.	Test for Mercury	258
5.13.10.	Test for Boron.....	258
5.13.11.	Test for Sulphur	258
5.13.12.	Test for Anthraquinones	258
5.13.13.	Test for Hingu.....	258

Appendix-6. Ayurvedic Definitions and Methods

6.1.	Kalpanā Paribhāṣā:	260
6.1.1.	Kalka:.....	260
6.1.2.	Kvātha / Kaṣāya:	260
6.1.3.	Cūrṇa:	260
6.1.4.	Putāpāka Svarasa:	260
6.1.5.	Svarasa:	260
6.1.6.	Hima Kaṣāya:	260
6.2.	Sāmānya paribhāṣā:.....	261
6.2.1.	Kajjalī:.....	261
6.2.2.	Kāñjika:	261
6.2.3.	Kṣāra Preparation:	261
6.2.4.	Cūrṇodaka:	261
6.2.5.	Mastu Preparation:	261
6.2.6.	Prakṣepa:.....	261
6.2.7.	Bhāvanā:	262
6.2.8.	Śodhana:	262
6.2.8.1.	Godantī Śodhana:	262
6.2.8.2.	Gairika Śodhana:.....	262
6.2.8.3.	Gandhaka Śodhana:.....	262
6.2.8.4.	Guggulu Śodhana:.....	263
6.2.8.5.	Ṭaṅkaṇa Śodhana:	263

6.2.8.6.	Tuttha Śodhana:	263
6.2.8.7.	Bhallātaka Śodhana:	263
6.2.8.8.	Maṇḥśilā Śodhana:.....	264
6.2.8.9.	Vatsanābha Śodhana:.....	264
6.2.8.10.	Karavīra Śodhana:	264
6.2.8.11.	Citraka Śodhana:.....	264
6.2.8.12.	Lāṅgalī Śodhana:.....	265
6.2.8.13.	Śilājatu Śodhana:	265
6.2.8.14.	Haritāla Śodhana:.....	265
6.2.8.15.	Hīṅgu Śodhana:	265
6.2.8.16.	Vijayā Śodhana:.....	266
6.2.8.17.	Kāśīśa Śodhana:	266
6.2.8.18.	Sauvīrāñjana Śodhana:	266
6.2.8.19.	Narasāra Śodhana:	266
6.2.8.20.	Pārada Sāmānya Śodhana:.....	266
6.2.8.21.	Aṣṭasaṃskāra of Pārada	267
6.2.8.21.a.	Svedana:.....	267
6.2.8.21.b.	Mardana:	267
6.2.8.21.c.	Mūrcchana:	268
6.2.8.21.d.	Utthāpana:.....	268
6.2.8.21.e.	Pātana:.....	268
6.2.8.21.f.	Rodhana / Bodhana:	269
6.2.8.21.g.	Niyāmana:.....	270
6.2.8.21. h.	Dīpana / Sandīpana:	270
6.2.9.	Mūrcchanā:	270
6.2.9.1.	Mūrcchanā of Eraṇḍa Taila:.....	271
6.2.9.2.	Mūrcchanā of Ghṛta:	272
6.2.9.3.	Mūrcchana of Saṛṣapa Taila:	273
6.2.9.4.	Mūrcchana of Tila Taila:	274
6.3.	Yantra Paribhāṣā:	275
6.3.1.	Khalva yantra:	275
6.3.2.	Tiryak pātana yantra:	275
6.3.4.	Dolāyantra:	275

Appendix-7. Weight and Measures

7.1. Metric Equivalents of Classical Weights and Measures276
7.2. Metric System.....277

Appendix-8. Classical Ayurvedic References

Appendix-9. List of Single Drugs used in Formulations

9.1. List of Single Drugs of Animal origin used in Formulations, with equivalent English name
9.2. List of Single Drugs of metal/mineral origin used in Formulations
9.3. List of Single Drugs of plant origin used in Formulation, with Latin Nomenclature.....

Appendix-10. Bibliography



APPENDICES

APPENDIX - 1

APPARATUS FOR TESTS AND ASSAYS

1.1. Nessler Cylinders

Nessler cylinders which are used for comparative tests are matched tubes of clear colourless glass with a uniform internal diameter and flat, transparent base. They comply with Indian Standard 4161-1967. They are of transparent glass with a nominal capacity of 50 ml. The overall height is about 150 mm, the external height to the 50 ml mark 110 to 124 mm, the thickness of the wall 1.0 to 1.5 mm and the thickness of the base 1.5 to 3.0 mm. The external height to the 50 ml mark of the cylinder used for a test must not vary by more than 1 mm.

1.2. Sieves

Sieves for pharmacopoeial testing are constructed from wire cloth with square meshes, woven from wire of brass, bronze, stainless steel or any other suitable material. The wires should be of uniform circular cross-section and should not be coated or plated. There must be no reaction between the material of the sieve and the substance being sifted.

Sieves conform to the following specifications –

Table 1

Approximate sieve number*	Nominal mesh aperture size	Tolerance average aperture size
	mm	± mm
4	4.0	0.13
6	2.8	0.09
8	2.0	0.07
10	1.7	0.06
12	1.4	0.05
16	1.0	0.03
--	µm	±µm
22	710	25
25	600	21
30	500	18
36	425	15

44	355	13
60	250	3 (9.9) **
85	180	11 (7.6)
100	150	9.4 (6.6)
120	125	8.1 (5.8)
150	106	7.4 (5.2)
170	90	6.6 (4.6)
200	75	6.1 (4.1)
240	63	5.3 (3.7)
300	53	4.8 (3.4)
350	45	4.8 (3.1)

* Sieve number is the number of meshes in a length of 2.54 cm. in each transverse direction parallel to the wires.

** Figures in brackets refer to close tolerances, those without brackets relate to full tolerances.

1.3. Thermometers

Unless otherwise specified, thermometers suitable for pharmacopoeial tests conform to Indian Standard 4825-1968 and are standardised in accordance with the 'Indian Standard Method of Calibrating Liquid-in-Glass Thermometers', 6274-1971.

The thermometers are of the mercury-in-glass type and are filled with a dried inert gas, preferably nitrogen. They may be standardised for total immersion or for partial immersion. Each thermometer should be employed according to the condition of immersion under which it was standardised. In the selection of the thermometer it is essential to consider the conditions under which it is to be used.

1.4. Ultraviolet Lamp (For general purposes and for chromatography work)

An instrument consisting of mercury vapour lamp and a filter which gives an emission band with maximum intensity at about 254 nm (near UV rays) and 366 nm (far UV rays) is used. To ensure that the required emission is being given by the lamp, carry out the following test periodically.

Apply to a plate coated with *silica gel* G, 5 µl of a 0.04 per cent w/v solution of *sodium salicylate* in *ethanol* (95%) for lamps of maximum output at 254 nm and 5 µl of a 0.2 per cent w/v solution in *ethanol* (95%) for lamps of maximum output at 365 nm. Examine the spot in a position normal to the

radiation. The distance between the lamp and the plate under examination used in a pharmacopoeial test should not exceed the distance used to carry out the above test.

1.5. Volumetric Glassware

Volumetric apparatus is normally calibrated at 27⁰. However, the temperature generally specified for measurements of volume in the analytical operations of the pharmacopoeia, unless otherwise stated, is 25⁰. The discrepancy is inconsequential as long as the room temperature in the laboratory is reasonably constant and is around 27⁰.

Pharmacopoeial assays involving volumetric measurements require the use of accurately calibrated glassware. Volumetric apparatus must be suitably designed to assure accuracy. The design, construction and capacity of volumetric glassware should be in accordance with those laid down by the Bureau of Indian Standards. The tolerances on capacity for volumetric flasks, pipettes and burettes, as laid down in the relevant Indian Standards, are permissible.

1.6. Weights and Balances

Pharmacopoeial tests and assays require the use of analytical balances that vary in capacity, sensitivity and reproducibility. The accuracy needed for a weighing should dictate the type of balance. Where substances are to be “accurately weighed”, the weighing is to be performed so as to limit the error to not more than 0.1 per cent. For example, a quantity of 50 mg is to be weighed to the nearest 0.05 mg; a quantity of 0.1 g is to be weighed to the nearest 0.1 mg; and quantity of 10 g is to be weighed to the nearest 10 mg. A balance should be chosen such that the value of three times the standard deviation of the reproducibility of the balance, divided by the amount to be weighed, does not exceed 0.001.

1.7. Muslin Cloth

Muslin cloth is a cotton fabric where warp is 22 per cm \pm 1 and weft is 18 \pm 1 per centimeter.

Method: Take a cardboard or an aluminium plate with a centimeter square opening. Keep the plate on the cloth to be used, so that the edges on the X or Y axis coincides with a warp or weft yarn in the fabric. Count the number of the threads of both warp and weft within the opening.

APPENDIX - 2

TESTS AND DETERMINATIONS

2.1. Microscopic identification:

Microscopic identification of the botanical ingredients is a standard for statutory purposes in several solid and semi-solid compound formulations. Microscopic identification tests are confined to those formulations where the botanical ingredients are **not more than ten**, and where they are added '*in situ*' in powder form as '*Prakṣepa Dravyas*'. Such comminuted ingredients lend themselves for microscopic identification, as they are not drastically changed in cell structure or contents while processing, and appear intact in microscopic slide preparations, after proper treatment.

Appropriate processing for separation and isolation of botanical debris from a formulation without loss of debris, by hand picking, shifting, washing, sedimentation, density separation or by floatation etc. are the preliminary steps. This is followed by clearing the debris in chemical reagents, reacting it with suitable reagents and stains and finally mounting a little part on a slide in a medium of suitable refractive index (see later part) that helps to show the unit structures in good relief. Identification of the discrete, but disoriented units from the botanical ingredients in a formulation will not be possible without proper isolation, and should not be attempted.

Monographs where the test is prescribed give both a relevant method of isolation and diagnostic features specific to the expected ingredients in that formulation. Only a brief method and a few of the characteristics for each ingredient are given, but an analyst may use other methods of isolation and choose more characteristics to draw a correct conclusion.

Although monographs prescribe standards only for the '*Prakṣepa Dravyas*', characteristics from other ingredients that are processed into extracts or decoctions prior to their addition to a formulation may also be seen in a slide preparation, giving rise to recognisable unique characteristics. In addition, cell or tissue structures common to several ingredients added to a formulation, and therefore not specific to any one of them, would also be present. Caution should therefore be exercised so that such features are not construed as parts from adulterants or substitutes or foreign parts. Proper study of the individual ingredients using authentic material and reference to their monographs in the Ayurvedic Pharmacopoeia for Single Drugs would help avoid errors of this nature. Skill in the recognition of discrete and disoriented tissue components and the knowledge required to ascribe them to their correct source should be acquired by the analyst.

A. Stains and Reagents for Microchemical Reactions:

The Ayurvedic Pharmacopoeia volumes on single drugs already include microchemical reactions for ergastic substances and may be consulted in addition to the following for use on isolated debris:

Acetic acid: Dilute 6 ml of glacial acetic acid with 100 ml of distilled water; *used for identification of cystoliths, which dissolve with effervescence.*

Aniline Chloride Solution: Dissolve 2 g in a mixture of 65 ml of 30 per cent ethyl alcohol and 15 ml distilled water and add 2 ml of conc. Hydrochloric acid. *Lignified tissues are stained bright yellow.*

Bismarck Brown: Dissolve 1 g in 100 ml of 95 per cent of ethyl alcohol; *used as a general stain for macerated material (with Schultze's).*

Chlorinated Soda Solution (Bleaching Solution): Dissolve 75 g of sodium carbonate in 125 ml of distilled water; triturate 50 g of chlorinated lime (bleaching powder) in a mortar with 75 ml of distilled water, adding it little by little. Mix the two liquids and shake occasionally for three or four hours. Filter and store, protected from light. *Used for lighting highly coloured material, by warming in it and washing the tissues thoroughly.*

Breamer's reagent: Dissolve 1 g of sodium tungstate and 2 g of sodium acetate in sufficient quantity of water to make 10 ml yellowish to brown precipitates; *indicate the presence of tannin.*

Canada Balsam (as a Mountant): Heat Canada balsam on a water bath until volatile matter is removed and the residue sets to a hard mass on cooling. Dissolve residue in xylene to form a thin syrupy liquid. *Used for making permanent mounts of reference slides of selected debris.*

Chloral Hydrate Solution: Dissolve 50 g of chloral hydrate in 20 ml of distilled water. *A valuable clarifying agent for rendering tissues transparent and clear, by freeing them from most of the ergastic substances, but leaving calcium oxalate crystals unaffected.*

Chloral Iodine: Saturate chloral hydrate solution with iodine, leaving a few crystals undissolved; *useful for detecting minute grains of starch otherwise undetectable.*

Chlorzinciodine (Iodinated Zinc Chloride solution): Dissolve 20 g of zinc chloride and 6.5 g of potassium iodide in 10 ml of distilled water. Add 0.5 g of iodine and shake for about fifteen minutes before filtering. Dilute if needed prior to use. *Renders cellulosic walls bluish violet and lignified walls yellowish brown to brown.*

Chromic acid Solution: 10 g of dissolved in 90 ml of dilute sulphuric acid: *macerating agent similar to Schultze's.*

Corallin Soda: Dissolve 5 g of corallin in 100 ml of 90 per cent ethyl alcohol. Dissolve 25 g of sodium carbonate in 100 ml distilled water; keep the solutions separate and mix when required, by adding 1 ml of the corallin solution to 20 ml of the aqueous sodium carbonate solution. Prepare fresh each time, as the mixture will not keep for long. *Used for staining sieve plates and callus bright pink and imparts a reddish tinge to starch grains and lignified tissues.*

Ammoniacal solution of Copper oxide (Cuoxam): Triturate 0.5 g of copper carbonate in a mortar with 10 ml of distilled water and gradually add 10 ml of strong solution of ammonia (sp. gr. 0.880) with continued stirring; *used for dissolving cellulosic materials.*

Eosin: 1 per cent solution in 90 per cent ethyl alcohol; *stains cellulose and aleurone grains red.*

Ferric Chloride solution: A per cent solution ferric chloride in distilled water. *Tannin containing tissues coloured bluish or greenish black.*

Glycerin: Pure or diluted as required with one or two volumes of distilled water. *Used as a general mountant.*

Haematoxylin, Delafield's: Prepare a saturated solution of ammonia alum. To 100 ml of this add a solution of 1 g of Haematoxylin in 6 ml of ethyl alcohol (97 per cent). Leave the mixed solution exposed to air and light in an unstopped bottle for three or four days. Filter and add to the filtrate 25 ml of glycerin and 25 ml of methyl alcohol. Allow the solution to stand exposed to light, till it acquires a dark colour (about two months). Refilter and store as a stock solution. Dilute it 3 or 4 times volumes with distilled water. *Stains cellulosic fibers blue; used only on water washed material.*

Iodine Water: Mix 1 volume of decinormal iodine with 4 volumes of distilled water. *Stains starch blue, and reveals crystalloids and globoids when present in aleurone grains.*

Iodine and Potassium iodide solution: Dissolve 1 g of *potassium iodide* in 200 ml of distilled water and 2 g of iodine; *stains lignified walls yellow and cellulosic walls blue.*

Lactophenol (Amman's Fluid): *Phenol 20 g, lactic acid 20 g, glycerin 40 g, distilled water 20 ml* dissolve; *reveals starch grains in polarised light with a well marked cross at hilum, and also minute crystals of calcium oxalate as brightly polarising points of light.*

Methylene blue: A solution in 25 ml of *ethyl alcohol* (95 per cent). *A general stain for nucleus and bacteria.*

Millon's Reagent: Dissolve 1 volume of mercury in 9 volumes of fuming nitric acid (sp. Gr. 1.52), keeping the mixture well cooled during reaction. Add equal volume distilled water when cool. *Stains proteins red.*

Naphthol Solution: Dissolve 10 g of Naphthol in 100 ml of *ethyl alcohol*; *a specific stain for detection of inulin; cells containing inulin turn deep reddish violet.*

Phloroglucinol: 1 g of *phloroglucinol* dissolved in 100 ml of 90 per cent *ethyl alcohol*; mount debris in a few drops, allow to react for a minute, draw off excess of reagent with a filter paper strip, and add a drop of conc. hydrochloric acid to the slide; *lignified tissues acquire a deep purplish red colour; very effective on water washed material but not in chloral hydrate washed debris.*

Picric acid Solution (Trinitrophenol Solution): A saturated aqueous solution made by dissolving 1 g of picric acid in 95 ml of distilled water; *stains animal and insect tissues, a light to deep yellow; in a solution with ethyl alcohol, aleurone grains and fungal hyphae are stained yellow.*

Potash, Caustic: A 5 per cent aqueous solution; *used to separate tenacious tissues of epidermis and also laticiferous elements and vittae, both of which are stained brown.*

Ruthenium Red: Dissolve 0.008 g of ruthenium red in 10 ml of a 10 per cent solution of lead acetate; (to be freshly prepared) *used for identification of most kinds of mucilage containing tissues,*

which turn pink. A 0.0008 g ruthenium red dissolved in 10 ml of distilled water and used immediately stains cuticular tissues in debris to a light pink.

Safranin: A 1 per cent solution in ethyl alcohol 50 per cent; used to stain lignified cell walls deep red, even after clearing with choral hydrate.

Schultze's Maceration Fluid: Add isolated debris to 50 per cent conc. *nitric acid* in a test tube and warm over water bath: add a few crystals of *potassium chlorate* while warming, till tissues soften; cool, wash with water thoroughly and tease out for mounting hard tissues; *isolated cell structures are clearly revealed, but the structures are not useful for measurement of dimensions.*

Sudan Red III: Dissolve 0.01 g of sudan red III in 5 ml of *ethyl alcohol* (90 per cent) and 5 ml of pure *glycerin*; *suberised walls of cork cells, and fatty material in cells are stained bright red.*

Sulphovanadic Acid (Mandelin's Reagent): Triturate 1 g of ammonium vandate with 100 ml conc. *sulphuric acid*. Allow the deposit to subside and use the clear liquid. *This is to be prepared fresh; useful for identification of alkaloids, particularly strychnine which turns violet in the cells containing it.*

Table 3 - Refractive Indices of Certain Mountants

Water	1.333
Lactophenol	1.444
Chloral Hydrate solution	1.44 to 1.48
Olive oil	1.46 to 1.47
Glycerol	1.473
Castor oil	1.48
Clove oil	1.53
Cresol	1.53
Cassia oil	1.6
Xylol	1.49
Alcohol	1.36
Chloroform	1.44

2.2. Determination of Quantitative Data:

2.2.1. Net Content:

The content of the final or retail pack shall not be less than 98 percent of the declared net content.

2.2.2. Foreign Matter:

The sample shall be free from visible signs of mold growth, sliminess, stones, rodent excreta, insects or any other noxious foreign matter when examined as given below.

Take a representative portion from a large container, or remove the entire contents of the packing if 100 g or less, and spread in a thin layer in a suitable dish or tray. Examine in daylight with unaided eye. Transfer suspected particles, if any, to a petri dish, and examine with 10x lens in daylight.

2.2.3. Determination of Total Ash:

Incinerate about 2 to 3 g accurately weighed, of the ground drug in a tared platinum or silica dish at a temperature not exceeding 450° until free from carbon, cool and weigh. If a carbon free ash cannot be obtained in this way, exhaust the charred mass with hot water, collect the residue on an ashless filter paper, incinerate the residue and filter paper, add the filtrate, evaporate to dryness, and ignite at a temperature not exceeding 450° . Calculate the percentage of ash with reference to the air-dried drug.

2.2.4. Determination of Acid-Insoluble Ash:

To the crucible containing total ash, add 25 ml of *dilute hydrochloric acid*. Collect the insoluble matter on an ashless filter paper (Whatman 41) and wash with hot water until the filtrate is neutral. Transfer the filter paper containing the insoluble matter to the original crucible, dry on a hot-plate and ignite to constant weight. Allow the residue to cool in a suitable desiccator for 30 minutes and weigh without delay. Calculate the content of acid-insoluble ash with reference to the air-dried drug.

2.2.5. Determination of Water Soluble Ash:

Boil the ash for 5 minutes with 25 ml of water; collect insoluble matter in a Gooch crucible or on an ashless filter paper, wash with hot water, and ignite for 15 minutes at a temperature not exceeding 450° . Subtract the weight of the insoluble matter from the weight of the ash; the difference in weight represents the water-soluble ash. Calculate the percentage of water-soluble ash with reference to the air-dried drug.

2.2.6. Determination of Sulphated Ash:

Heat a silica or platinum crucible to redness for 10 minutes, allow to cool in a desiccator and weigh. Put 1 to 2 g of the substance, accurately weighed, into the crucible, ignite gently at first, until the substance is thoroughly charred. Cool, moisten the residue with 1 ml of *sulphuric acid*, heat gently until white fumes are no longer evolved and ignite at $800^{\circ} \pm 25^{\circ}$ until all black particles have disappeared. Conduct the ignition in a place protected from air currents. Allow the crucible to cool, add a few drops of *sulphuric acid* and heat. Ignite as before, allow to cool and weigh. Repeat the operation until two successive weighing do not differ by more than 0.5 mg.

2.2.7. Determination of Alcohol Soluble Extractive:

Macerate 5 g of the air dried drug, coarsely powdered, with 100 ml of alcohol the specified strength in a closed flask for twenty-four hours, shaking frequently during six hours and allowing to stand for eighteen hours. Filter rapidly, taking precautions against loss of solvent, evaporate 25 ml of the filtrate to dryness in a tared flat bottomed shallow dish, and dry at 105⁰, to constant weight and weigh. Calculate the percentage of alcohol-soluble extractive with reference to the air-dried drug. For determination of methanol soluble extractive use methanol on place of alcohol.

2.2.8. Determination of Water Soluble Extractive:

Proceed as directed for the determination of alcohol-soluble extractive, using *chloroform-water* instead of ethanol.

2.2.9. Determination of Ether Soluble Extractive (Fixed Oil Content):

Transfer a suitably weighed quantity (depending on the fixed oil content) of the air-dried, crushed drug to an extraction thimble, extract with *solvent ether* (or *petroleum ether*, b.p. 40⁰ to 60⁰) in a continuous extraction apparatus (Soxhlet extractor) for 6 hours. Filter the extract quantitatively into a tared evaporating dish and evaporate off the solvent on a water bath. Dry the residue at 105⁰ to constant weight. Calculate the percentage of ether-soluble extractive with reference to the air-dried drug.

2.2.10. Determination of Moisture Content (Loss on Drying):

Procedure set forth here determines the amount of volatile matter (i.e., water drying off from the drug). For substances appearing to contain water as the only volatile constituent, the procedure given below, is appropriately used.

Place about 10 g of drug (without preliminary drying) after accurately weighing (accurately weighed to within 0.01 g) it in a tared evaporating dish. For example, for unground or unpowderd drug, prepare about 10 g of the sample by cutting shredding so that the parts are about 3 mm in thickness.

Seeds and fruits, smaller than 3 mm should be cracked. Avoid the use of high speed mills in preparing the samples, and exercise care that no appreciable amount of moisture is lost during preparation and that the portion taken is representative of the official sample. After placing the above said amount of the drug in the tared evaporating dish, dry at 105⁰ for 5 hours, and weigh. Continue the drying and weighing at one hour interval until difference between two successive weighing corresponds to not more than 0.25 per cent. Constant weight is reached when two consecutive weighing after drying for 30 minutes and cooling for 30 minutes in a desiccator, show not more than 0.01 g difference.

2.2.11. Determination of Volatile Oil in Drugs

The determination of volatile oil in a drug is made by distilling the drug with a mixture of *water* and *glycerin*, collecting the distillate in a graduated tube in which the aqueous portion of the distillate is

automatically separated and returned to the distilling flask, and measuring the volume of the oil. The content of the volatile oil is expressed as a percentage v/w.

The apparatus consists of the following parts (see Fig. 1). The clevenger's apparatus described below is recommended but any similar apparatus may be used provided that it permits complete distillation of the volatile oil. All glass parts of the apparatus should be made of good quality resistance glass.

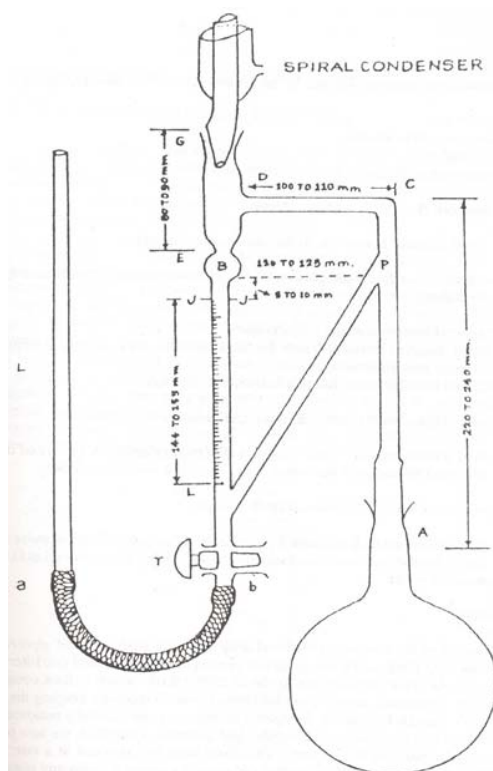


Fig. 1 Apparatus for volatile oil determination

The apparatus is cleaned before each distillation by washing successively with *acetone* and *water*, then inverting it, filling it with *chromic sulphuric acid* mixture, after closing the open end at G, and allowing to stand, and finally rinsing with water.

Method of determination:

A suitable quantity of the coarsely powdered drug together with 75 ml of *glycerin* and 175 ml of *water* in the one litre distilling flask, and a few pieces of porous earthen ware and one filter paper 15 cm cut into small strips, 7 to 12 mm wide, are also put in the distilling flask, which is then connected to the still head. Before attaching the condenser, water is run into the graduated receiver, keeping the tap T open until the water overflows, at P. Any air bubbles in the rubber tubing a—b are carefully removed by pressing the tube. The tap is then closed and the condenser attached. The contents of the flask are now heated and stirred by frequent agitation until ebullition commences. The distillation is

continued at a rate, which keeps the lower end of the condenser cool. The flask is rotated occasionally to wash down any material that adheres to its sides.

At the end of the specified time (3 to 4 hours) heating is discontinued, the apparatus is allowed to cool for 10 minutes and the tap T is opened and the tube L₁ lowered slowly; as soon as the layer of the oil completely enters into the graduated part of the receiver the tap is closed and the volume is read.

The tube L₁ is then raised till the level of water in it is above the level of B, when the tap T is slowly opened to return the oil to the bulb. The distillation is again continued for another hour and the volume of oil is again read, after cooling the apparatus as before. If necessary, the distillation is again continued until successive readings of the volatile oil do not differ.

The measured yield of volatile oil is taken to be the content of volatile oil in the drug. The dimensions of the apparatus may be suitably modified in case of necessity.

2.2.12. Special Processes Used in Alkaloidal Assays:

A-Continuous extraction of drug:

Where continuous extraction of a drug of any other substance is recommended in the monograph, the process consists of percolating it with suitable solvent at a temperature approximately that of the boiling point of the solvent. Any apparatus that permits the uniform percolation of the drug and the continuous flow of the vapour of the solvent around the percolator may be used. The type commonly known as the Soxhlet apparatus is suitable for this purpose.

B -Tests for complete extraction of alkaloids: Complete extraction is indicated by the following tests:

When extracting with an aqueous or alcoholic liquid: After extracting at least three times with the liquid, add few drops of 0.05 ml of *potassium mercuri-iodide solution* or for solanaceous alkaloids 0.05 ml of *potassium iodobismuthate solution* to a the next portion, after acidifying with 2 N *hydrochloric acid* if necessary; no precipitate or turbidity, is produced.

When extracting with an immiscible solvent: After extracting at least three times with the solvent, add to 1 to 2 ml of of 0.1 N *hydrochloric acid* to 1 to 2 ml of next portion, remove the organic solvent by evaporation, transfer the aqueous layer to a test tube, and add 0.05 ml of *potassium mercuri-iodide solution* or 0.05 ml of *potassium iodobismuthate solution* for solanaceous alkaloids or for emetine, 0.05 ml of *iodine solution*; not more than a very faint opalescence is produced.

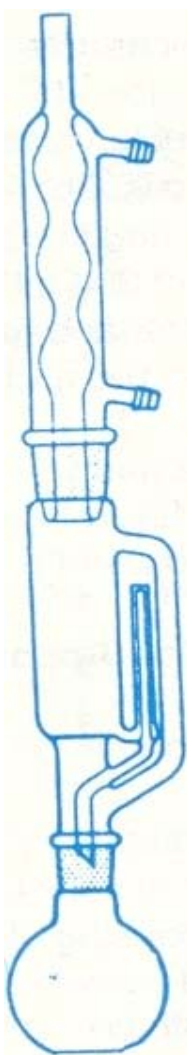


Fig. 2 - Apparatus for the continuous extraction of Drugs (Soxhlet apparatus)

2.2.13. Thin-Layer Chromatography (TLC):

Thin-layer chromatography is a technique in which a solute undergoes distribution between two phases, stationary phase acting through adsorption and a mobile phase in the form of a liquid. The adsorbent is a relatively thin, uniform layer of dry finely powdered material applied to a glass, plastic or metal sheet or plate. Precoated plates are most commonly used. Separation may also be achieved on the basis of partition or a combination of partition and adsorption, depending on the particular type of support, its preparation and its use with different solvent.

Identification can be effected by observation of spots of identical R_f value and about equal magnitude obtained, respectively, with an unknown and a reference sample chromatographed on the same plate. A visual comparison of the size and intensity of the spots usually serves for semi-quantitative estimation.

Apparatus:

- (a) Flat glass plates of appropriate dimensions which allow the application at specified points of the necessary quantities of the solution being examined and appropriate reference solutions and which allow accommodation of the specified migration path-length. The plates are prepared as described below; alternatively, commercially prepared plates may be used.
- (b) An aligning tray or a flat surface on which the plates can be aligned and rested when the coating substance is applied.
- (c) The adsorbent or coating substance consisting of finely divided adsorbent materials, normally 5 μm to 40 μm in diameter is suitable for chromatography. It can be applied directly to the plate or can be bonded to the plate by means of plaster of paris (Hydrated Calcium Sulphate) or with any other suitable binders. The adsorbent may contain fluorescing material to help in visualising spots that absorb ultra-violet light.
- (d) A spreader which, when moved over the glass plate, will apply a uniform layer of adsorbent of desired thickness over the entire surface of the plate.
- (e) A storage rack to support the plates during drying and transportation.
- (f) A developing chamber that can accommodate one or more plates and can be properly closed and sealed. The chamber is fitted with a plate support rack that supports the plates, back to back, with lid of the chamber in place.
- (g) Graduated micro-pipettes capable of delivering microlitre quantities say 10 μl and less.
- (h) A reagent sprayer that will emit a fine spray and will not itself be attacked by the reagent.
- (i) An ultra-violet light, suitable for observation at short (254 nm) and long (366 nm) ultra-violet wavelengths.

Preparation of plates:

Unless otherwise specified in the monograph, the plates are prepared in the following manner. Prepare a suspension of the coating substance in accordance with the instructions of the supplier and, using the spreading device designed for the purpose, spread a uniform layer of the suspension, 0.20 to 0.30 mm thick, on a flat glass plate 20 cm long. Allow the coated plates to dry in air, heat at 100° to 105° for at least 1 hour (except in the case of plates prepared with cellulose when heating for 10 minutes is normally sufficient) and allow to cool, protected from moisture. Store the plates protected from moisture and use within 3 days of preparation. At the time of use, dry the plates again, if necessary, as prescribed in the monographs. Now a days pre coated plates of silica gel on glass/aluminium/ plastic sheets are also available.

Method:

Unless unsaturated conditions are prescribed, prepare the tank by lining the walls with sheets of filter paper; pour into the tank, saturating the filter paper in the process, sufficient of the mobile phase to

form a layer of solvent 5 to 10 mm deep, close the tank and allow to stand for 1 hour at room temperature. Remove a narrow strip of the coating substance, about 5 mm wide, from the vertical sides of the plate. Apply the solutions being examined in the form of circular spots about 2 to 6 mm in diameter, or in the form of bands (10 to 20 mm x 2 to 6 mm unless otherwise specified) on a line parallel with, and 20 mm from, one end of the plate, and not nearer than 20 mm to the sides; the spots should be 15 mm apart. If necessary, the solutions may be applied in portions, drying between applications. Mark the sides of the plate 15 cm, or the distance specified in the monograph, from the starting line. Allow the solvent to evaporate and place the plate in the tank, ensuring that it is as nearly vertical as possible and that the spots or bands are above the level of the mobile phase. Close the tank and allow to stand at room temperature, until the mobile phase has ascended to the marked line. Remove the plate and dry and visualise as directed in the monograph; where a spraying technique is prescribed it is essential that the reagent be evenly applied as a fine spray.

For two-dimensional chromatography dry the plate after the first development and carry out the second development in a direction perpendicular to the first.

When the method prescribed in the monograph specifies ‘protected from light’ or ‘in subdued light’ it is intended that the entire procedure is carried out under these conditions.

Visualisation:

The phrases *ultra-violet light (254 nm)* and *ultra-violet light (366 nm)* indicate that the plate should be examined under an ultra-violet light having a maximum output at about 254 or at about 365 nm, as the case may be.

The term *secondary spot* means any spot other than the principal spot. Similarly, a *secondary band* is any band other than the principal band.

R_f Value :

Measure and record the distance of each spot from the point of its application and calculate the R_f value by dividing the distance travelled by the spots by the distance travelled by the front of the mobile phase.

2.2.14. Fatty oil estimation:

To estimate fatty oils, extract accurately weighed air-dried powdered plant material with *petroleum ether* (40-60^o) in a Soxhlet apparatus. Dry the extract over *anhydrous sodium sulphate* and remove the solvent under vacuum at 40^o. Weigh the residue and calculate the percentage with reference to the weight of plant material used.

2.3. Limit Tests:

Table 4- Permissible Limits of Heavy Metals and Arsenic

S.No.	Heavy Metal contents and Arsenic	Permissible limits
-------	----------------------------------	--------------------

1	Lead	10 ppm
2	Arsenic	3 ppm
3	Cadmium	0.3 ppm
4	Mercury	1 ppm

2.3.1. Limit Test for Arsenic

In the limit test for arsenic, the amount of arsenic is expressed as arsenic, As ppm

Apparatus –

A wide-mouthed bottle capable of holding about 120 ml is fitted with a rubber bung through which passes a glass tube. The latter, made from ordinary glass tubing, has a total length of 200 mm and an internal diameter of exactly 6.5 mm (external diameter about 8 mm). It is drawn out at one end to a diameter of about 1 mm and a hole not less than 2 mm in diameter is blown in the side of the tube, near the constricted part. When the bung is inserted in the bottle containing 70 ml of liquid, the constricted end of the tube is above the surface of the liquid, and the hole in the side is below the bottom of the bung. The upper end of the tube is cut off square, and is either slightly rounded or ground smooth.

Two rubber bungs (about 25 mm x 25 mm), each with a hole bored centrally and true, exactly 6.5 mm in diameter, are fitted with a rubber band or spring clip for holding them tightly together. Alternatively the two bungs may be replaced by any suitable contrivance satisfying the conditions described under *the General Test*.

Reagents:

Ammonium oxalate AsT: *Ammonium oxalate* which complies with the following additional test:

Heat 5 g with 15 ml of *water*, 5 ml of *nitric acid AsT*, and 10 ml of *sulphuric acid AsT* in narrow necked, round-bottomed flask until frothing ceases, cool, and apply the General Test; no visible stain is produced.

Arsenic solution, dilute, AsT:

<i>Strong Arsenic solution AsT</i>	1 ml
<i>Water</i> sufficient to produce	100 ml

Dilute arsenic solution, AsT must be freshly prepared.
1 ml contains 0.01 mg of arsenic (As).

Arsenic solution, strong, AsT:

<i>Arsenic trioxide</i>	0.132 g
<i>Hydrochloric acid</i>	50 ml
<i>Water</i> sufficient to produce	100 ml

Brominated hydrochloric acid AsT:

<i>Bromine solution AsT</i>	1 ml
<i>Hydrochloric acid AsT</i>	100 ml

Bromine solution AsT:

<i>Bromine</i>	30 g
<i>Potassium bromide</i>	30 g
<i>Water sufficient to produce</i>	100 ml

It complies with the following test:

Evaporate 10 ml on a water-bath nearly to dryness, add 50 ml of purified water, 10 ml of *hydrochloric acid AsT* and sufficient *stannous chloride solution AsT* to reduce the remaining bromine and apply the General Test; the stain produced is not deeper than 1 ml *standard stain*, showing that the proportion of arsenic present does not exceed 1 part per million.

Citric acid AsT: *Citric acid* which complies with the following additional tests: Dissolve 10 g in 50 ml of water add 10 ml of *stannated hydrochloric acid AsT* and apply the General Test; no visible stain is produced.

Hydrochloric acid AsT: *Hydrochloric acid* diluted with *water* to contain about 32 per cent w/w of *hydrochloride acid* and complying with the following additional tests:

- (i) Dilute 10 ml with sufficient water to produce 50 ml, add 5 ml of *ammonium thiocyanate solution* and stir immediately; no colour is produced.
- (ii) To 50 ml add 0.2 ml of *bromine solution AsT*, evaporate on a water-bath until reduced to 16 ml adding more *bromine solution AsT*, if necessary, in order that an excess, as indicated by the colour, may be present throughout the evaporation; add 50 ml of *water* and 5 drops of *stannous chloride solution AsT*, and apply the General Test; the stain produced is not deeper than a 0.2 ml *standard stain* prepared with the same acid, showing that the proportion of arsenic present does not exceed 0.05 part per million.

Hydrochloric acid (constant-boiling composition) As T : Boil *hydrochloric acid AsT* to constant boiling Composition in the presence of *hydrazine hydrate*, using 1 ml of 10 per cent w/v solution in *water* per litre of the acid.

Mercuric Chloride Paper*: Smooth white filter paper, not less than 25 mm in width, soaked in a saturated solution of *mercuric chloride*, pressed to remove superfluous solution, and dried at about 60°, in the dark. The grade of the filter paper is such that the weight is between 65 and 120 g per sq

* NOTE –*Mercuric chloride paper should be stored in a stoppered bottle in the dark. Paper which has been exposed to sunlight or to the vapour of ammonia affords a lighter stain or no stain at all when employed in the limit test for arsenic.*

mm; the thickness in mm of 400 papers is approximately equal numerically, to the weight in g per sq. mm..

Nitric acid AsT: *Nitric acid* which complies with the following additional test:

Heat 20 ml in a porcelain dish with 2 ml of *sulphuric acid AsT*, until white fumes are given off. Cool, add 2 ml of water, and again heat until white fumes are given off; cool, add 50 ml of water and 10 ml of *stannated hydrochloric acid AsT*, and apply the General Test; no visible stain is produced.

Potassium chlorate AsT: *Potassium chlorate* which complies with the following additional test:

Mix 5 g in the cold with 20 ml of *water* and 22 ml of *hydrochloric acid AsT*; when the first reaction has subsided, heat gently to expel chlorine, remove the last traces with a few drops of *stannous chloride solution AsT*, add 20 ml of water, and apply the General Test; no visible stain is produced.

Potassium iodide AsT: *Potassium iodide* which complies with the following additional test:

Dissolve 10 g in 25 ml of *hydrochloric acid AsT* and 35 ml of *water*, add 2 drops of *stannous chloride solution AsT* and apply the General Test; no visible stain is produced.

Potassium iodide AsT: *Potassium iodide* which complies with the following additional test:

Dissolve 10 g in 25 ml of *hydrochloric acid AsT* and 35 ml of *water*, add 2 drops of *stannous chloride solution AsT* and apply the General Test; no visible stain is produced.

Sodium carbonate, anhydrous AsT: *Anhydrous sodium carbonate* which complies with the following additional test:

Dissolve 5 g in 50 ml of *water*, add 20 ml of *brominated hydrochloric acid AsT*, remove the excess of bromine with a few drops of *stannous chloride solution AsT*, and apply the General Test; no visible stain is produced.

Sodium Salicylate: Of the Indian Pharmacopoeia.

Stannated hydrochloric acid AsT:

<i>Stannous chloride solution AsT</i>	1 ml
<i>Hydrochloric Acid AsT</i>	100 ml

Stannous Chloride solution AsT: Prepared from *stannous chloride solution* by adding an equal volume of *hydrochloric acid*, boiling down to the original volume, and filtering through a fine-grain filter paper.

It complies with the following test:

To 10 ml add 6 ml of water and 10 ml of *hydrochloric acid AsT*, distil and collect 16 ml. To the distillate add 50 ml of *water* and 2 drops of *stannous chloride solution AsT* and apply the General Test; the stain produced is not deeper than a 1-ml *standard stain*, showing that the proportion of arsenic present does not exceed 1 part per million.

Sulphuric acid AsT: *Sulphuric acid* which complies with the following additional test:

Dilute 10 g with 50 ml of water, add 0.2 ml of *stannous chloride solution AsT*, and apply the General Test; no visible stain is produced.

Zinc AsT: *Granulated Zinc* which complies with following additional test:

Add 10 ml of *stannated hydrochloric acid AsT* to 50 ml of *water*, and apply the General Test, using 10 of the zinc and allowing the action to continue for one hour; no visible stain is produced (limit of arsenic). Repeat the test with the addition of 0.1 ml of *dilute arsenic solution AsT*; a faint but distinct yellow stain is produced (test for sensitivity).

General Method of Testing: By a variable method of procedure suitable to the particular needs of each substance, a solution is prepared from the substance being examined which may or may not contain that substance, but contains the whole of the arsenic (if any) originally present in that substance. This solution, referred to as the 'test solution', is used in the actual test.

General Test: The glass tube is lightly packed with cotton wool, previously moistened with *lead acetate solution* and dried, so that the upper surface of the cotton wool is not less than 25 mm below the top of the tube. The upper end of the tube is then inserted into the narrow end of one of the pair of rubber bungs, either to a depth of about 10 mm when the tube has a rounded-off end, or so that the ground end of the tube is flush with the larger end of the bung. A piece of *mercuric chloride paper* is placed flat on the top of the bung and the other bung placed over it and secured by means of the rubber band or spring clip in such a manner that the borings of the two bungs (or the upper bung and the glass tube) meet to form a true tube 6.5 mm in diameter interrupted by a diaphragm of *mercuric chloride paper*.

Instead of this method of attaching the *mercuric chloride paper*, any other method may be used provided (1) that the whole of the evolved gas passes through the paper; (2) that the portion of the paper in contact with the gas is a circle 6.5 mm in diameter; and (3) that the paper is protected from sunlight during the test. The test solution prepared as specified, is placed in the wide-mouthed bottle, 1 g of *potassium iodide AsT* and 10 g of *zinc AsT* added, and the prepared glass tube is placed quickly in position. The action is allowed to proceed for 40 minutes. The yellow stain which is produced on the *mercuric chloride paper* if arsenic is present is compared by day light with the *standard stains* produced by operating in a similar manner with known quantities of *dilute arsenic solution AsT*. The comparison of the stains is made immediately at the completion of the test. The standard stains used for comparison are freshly prepared; they fade on keeping.

By matching the depth of colour with *standard stains*, the proportion of arsenic in the substance may be determined. A stain equivalent to the 1-ml standard stain, produced by operating on 10 g of substance indicates that the proportion of arsenic is 1 part per million.

NOTE: (1) The action may be accelerated by placing the apparatus on a warm surface, care being taken that the *mercuric chloride paper* remains dry throughout the test.

- (2) The most suitable temperature for carrying out the test is generally about 40^o but because the rate of the evolution of the gas varies somewhat with different batches zinc AsT, the temperature may be adjusted to obtain a regular, but not violent, evolution of gas.
- (3) The tube must be washed with *hydrochloric acid AsT*, rinsed with water and dried between successive tests.

Standard Stains: Solutions are prepared by adding to 50 ml of water, 10 ml of *stannated hydrochloric acid AsT* and quantities of *dilute arsenic solutions AsT* varying from 0.2 ml to 1 ml. The resulting solutions, when treated as described in the General Test, yield stains on the *mercuric chloride paper* referred to as the standard stains.

Preparation of the Test Solution: In the various methods of preparing the test solution given below, the quantities are so arranged unless otherwise stated, that when the stain produced from the solution to be examined is not deeper than the 1-ml standard stain, the proportion of arsenic present does not exceed the permitted limit.

Ammonium Chloride: Dissolve 2.5 g in 50 ml of *water*, and 10 ml of *stannated hydrochloric acid AsT*.

Boric acid: Dissolve 10 g with 2 g of *citric acid AsT* in 50 ml *water*, and add 12 ml of *stannated hydrochloric acid AsT*.

Ferrous Sulphate: Dissolve 5 g in 10 ml of *water* and 15 ml of *stannated hydrochloric acid AsT* and distil 20 ml; to the distillate add a few drops of *bromine solution AsT*. Add 2 ml of *stannated hydrochloric acid AsT*, heat under a reflux condenser for one hour, cool, and add 10 ml of *water* and 10 ml of *hydrochloric acid AsT*.

Glycerin: Dissolve 5 g in 50 ml of *water*, and add 10 ml of *stannated hydrochloric acid AsT*.

Hydrochloric acid: Mix 10 g with 40 ml of *water* and 1 ml of *stannous chloride solution AsT*.

Magnesium Sulphate: Dissolve 5 g in 50 ml of *water* and add 10 ml of *stannated hydrochloric acid AsT*.

Phosphoric acid: Dissolve 5 g in 50 ml of *water* and add 10 ml of *stannated hydrochloric acid AsT*

Potassium iodide: Dissolve 5 g in 50 ml of *water* and add 2 ml of *stannated hydrochloric acid AsT*.

Sodium bicarbonate: Dissolve 5 g in 50 ml of *water* and add 15 ml of *brominated hydrochloric acid AsT*, and remove the excess of bromine with a few drops of *stannous chloride solution AsT*.

Sodium hydroxide: Dissolve 2.5 g in 50 ml of *water*, add 16 ml of *brominated hydrochloric acid AsT*, and remove the excess of *bromine* with a few drops of *stannous chloride solution AsT*.

2.3.2. Limit Test for Chlorides:

Dissolve the specified quantity of the substance in *water* or prepare a solution as directed in the text and transfer to a *Nessler cylinder*. Add 10 ml of *dilute nitric acid*, except when nitric acid is used in

the preparation of the solution, dilute to 50 ml with water, and add 1 ml of *silver nitrate solution*. Stir immediately with a glass rod and allow to stand for 5 minutes. The opalescence produced is not greater than the *standard opalescence*, when viewed transversely.

Standard Opalescence:

Place 1.0 ml of a 0.05845 per cent w/v solution of *sodium chloride* and 10 ml of *dilute nitric acid* in a *Nessler cylinder*. Dilute to 50 ml with water and add 1 ml of *silver nitrate solution*. Stir immediately with a glass rod and allow to stand for five minutes.

2.3.3. Limit Test for Heavy metals:

The test for heavy metals is designed to determine the content of metallic impurities that are coloured by sulphide ion, under specified conditions. The limit for heavy metals is indicated in the individual monographs in terms of the parts of lead per million parts of the substance (by weight), as determined by visual comparison of the colour produced by the substance with that of a control prepared from a standard lead solution.

Determine the amount of heavy metals by one of the following methods and as directed in the individual monographs. Method A is used for substances that yield clear colourless solutions under the specified test conditions. Method B is used for substances that do not yield clear, colourless solutions under the test conditions specified for method A, or for substances which, by virtue of their complex nature, interfere with the precipitation of metals by sulphide ion. Method C is used for substances that yield clear, colourless solutions with *sodium hydroxide solution*.

Special Reagents:

Acetic acid Sp.: *Acetic acid* which complies with the following additional test : Make 25 ml alkaline with *dilute ammonia solution Sp.*, add 1 ml of *potassium cyanide solution Sp.*, dilute to 50 ml with *water* and add two drops of *sodium sulphide solution*; no darkening is produced.

Dilute acetic acid Sp.: *Dilute acetic acid*, which complies with the following additional test – Evaporate 20 ml in a porcelain dish, nearly to dryness on a water-bath. Add to the residue 2 ml of the acid and dilute with water to 25 ml, add 10 ml of *hydrogen sulphide solution*. Any dark colour produced is not more than that of a control solution consisting of 2 ml of the acid and 4.0 ml of *standard lead solution* diluted to 25 ml with *water*.

Ammonia solution Sp.: *Strong ammonia solution* which complies with the following additional test: Evaporate 10 ml to dryness on a water-bath; to the residue add 1 ml of *dilute hydrochloric acid Sp.* and evaporate to dryness. Dissolve the residue in 2 ml of dilute acetic acid Sp. Add sufficient water to produce 25 ml.

Add 10 ml of *hydrogen sulphide solution*. Any darkening produced is not greater than in a blank solution containing 2 ml of dilute acetic acid Sp. 1.0 ml of *standard lead solution* and sufficient *water* to produce 25 ml.

Dilute ammonia solution Sp.: *Dilute ammonia solution* which complies with the following additional test: To 20 ml, add 1 ml of *potassium cyanide solution Sp.*, dilute to 50 ml with *water*, and add two drops of *sodium sulphide solution*; no darkening is produced.

Hydrochloric acid: *Hydrochloric acid* which complies with the following additional test: Evaporate off the acid in a beaker to dryness on a water-bath. Dissolve the residue in 2 ml of *dilute acid Sp.*, dilute to 17 ml with *water* and add 10 ml of *hydrogen sulphide solution*; any darkening produced is not greater than in a blank solution containing 2.0 ml of *standard lead solution*, 2 ml of *dilute acetic acid Sp.* and dilute to 40 ml with *water*.

Dilute hydrochloric acid Sp.: *Dilute hydrochloric acid*, which complies with the following additional test: Treat 10 ml of the acid in the manner described under *Hydrochloric acid Sp.*

Lead nitrate stock solution: Dissolve 0.1598 g of *lead nitrate* in 100 ml of *water* to which has been added 1 ml of *nitric acid*, then dilute with *water* to 1000 ml. This solution must be prepared and stored in polyethylene or glass containers free from soluble lead salts.

Standard lead solution: On the day of use, dilute 10.0 ml of *lead nitrate* stock solution with *water* to 100.0 ml. Each ml of *standard lead solution* contains the equivalent of 10 µg of lead. A control comparison solution prepared with 2.0 ml of *standard lead solution* contains, when compared to a solution representing 1.0 g of the substance being tested, the equivalent of 20 parts per million of lead.

Nitric acid Sp.: *Nitric acid* which complies with the following additional test: Dilute 10 ml with 10 ml of *water*, make alkaline with *ammonia solution Sp.*, add 1 ml of *potassium cyanide solution Sp.*, dilute to 50 ml with *water*, and add two drops of *sodium sulphide solution*; no darkening is produced.

Potassium cyanide solution Sp.: See Appendix 2.3.5.

Sulphuric acid Sp.: Sulphuric acid which complies with following additional test: Add 5 g to 20 ml of *water* make alkaline with *ammonia solution Sp.*, add 1 ml of *potassium cyanide solution Sp.*, dilute to 50 ml with *water* and add two drops of *sodium sulphide solution*; no darkening is produced.

Method A

Standard solution: Into a 50 ml *Nessler cylinder*, pipette 2 ml of *standard lead solution* and dilute with *water* to 25 ml. Adjust with *dilute acetic acid Sp.* or *dilute ammonia solution Sp.* to a pH between 3.0 and 4.0, dilute with *water* to about 35 ml, and mix.

Test solution: In to a 50 ml *Nessler cylinder*, place 25 ml of the solution prepared for the test as directed in the individual monograph, or using the stated volume of acid when specified in the individual monograph, dissolve and dilute with *water* to 25 ml the specified quantity of the substance

being tested. Adjust with *dilute acetic acid Sp.* or *dilute ammonia solution Sp.* to a pH between 3.0 and 4.0, dilute with *water* to about 35 ml and mix.

Procedure: To each of the cylinders containing the *standard solution* and test solution, respectively, add 10 ml of freshly prepared *hydrogen sulphide solution*, mix, dilute with *water* to 50 ml, allow to stand for five minutes, and view downwards over a white surface; the colour produced in the *test solution* is not darker than that produced in the *standard solution*.

Method B

Standard solution: Proceed as directed under Method A.

Test solution: Weigh in a suitable crucible the quantity of the substance specified in individual monograph, add sufficient *sulphuric acid Sp.* to wet the sample, and ignite carefully at a low temperature until thoroughly charred. Add to the charred mass 2 ml of *nitric acid Sp.* and five drops of *sulphuric acid Sp.* and heat cautiously until white fumes are no longer evolved. Ignite, preferably in a muffle furnace, at 500⁰ to 600⁰ until the carbon is completely burnt off. Cool, add 4 ml of *hydrochloric acid Sp.*, cover, digest on a water bath for 15 minutes, uncover and slowly evaporate to dryness on a water-bath. Moisten the residue with one drop of *hydrochloric acid Sp.*, add 10 ml of hot water and digest for two minutes. Add *ammonia solution sp.*, dropwise, until the solution is just alkaline to *litmus paper*, dilute with *water* to 25 ml and adjust with dilute acetic acid *Sp.* to a pH between 3.0 and 4.0. Filter if necessary, rinse the crucible and the filter with 10 ml of water, combine the filtrate and washings in a 50 ml *Nessler cylinder*, dilute with *water*, to about 35 ml, and mix. Procedure: Proceed as directed under Method A.

Method C

Standard solution: Into a 50 ml *Nessler cylinder*, pipette 2 ml of *standard lead solution*, add 5 ml of *dilute sodium hydroxide solution.*, dilute with *water* to 50 ml and mix.

Test solution: Into a 50 ml *Nessler cylinder*, place 25 ml of the solution prepared for the test as directed in the individual monograph; or, if not specified otherwise in the individual monograph, dissolve the specified quantity in a mixture of 20 ml of *water* and 5 ml of *dilute sodium hydroxide solution*. Dilute 50 ml with water and mix.

Procedure: To each of the cylinders containing the *standard solution* and the *test solution*, respectively add 5 drops of *sodium sulphide solution*, mix, allow to stand for five minutes and view downwards over a white surface; the colour produced in the *test solution* is not darker than that produced in the *standard solution*.

2.3.4. Limit Test for Iron

Standard Iron solution: Weigh accurately 0.1726 g of *ferric ammonium sulphate* and dissolve in 10 ml of 0.1 N *sulphuric acid* and sufficient *water* to produce 1000.0 ml. Each ml of this solution contains 0.02 mg of Fe.

Method:

Dissolve the specified quantity of the substance being examined in 40 ml of *water*, or use 10 ml of the solution prescribed in the monograph, and transfer to a *Nessler cylinder*. Add 2 ml of a 20 per cent w/v solution of *iron-free citric acid* and 0.1 ml of *thioglycollic acid*, mix, make alkaline with *iron-free ammonia solution*, dilute to 50 ml with *water* and allow to stand for five minutes. Any colour produced is not more intense than the standard colour.

Standard colour: Dilute 2.0 ml of *standard iron solution* with 40 ml of *water* in a *Nessler cylinder*. Add 2 ml of a 20 per cent w/v solution of *iron-free citric acid* and 0.1 ml of *thioglycollic acid*, mix, make alkaline with *iron-free ammonia solution*, dilute to 50 ml with *water* and allow to stand for five minutes.

2.3.5. Limit Test for Lead

The following method is based on the extraction of lead by solutions of *dithizone*. All reagents used for the test should have as low a content of lead as practicable. All reagent solutions should be stored in containers of borosilicate glass. Glassware should be rinsed thoroughly with warm *dilute nitric acid*, followed by *water*.

Special Reagents:

- (1) **Ammonia-cyanide solution Sp.:** Dissolve 2 g of *potassium cyanide* in 15 ml of *strong ammonia solution* and dilute with *water* to 100 ml.
- (2) **Ammonium citrate solution Sp.:** Dissolve 40 g of *citric acid* in 90 ml *water*. Add two drops of *phenol red solution* then add slowly *strong ammonia solution* until the solution acquires a reddish colour. Remove any lead present by extracting the solution with 20 ml quantities of *dithizone* extraction solution until the *dithizone* solution retains its orange-green colour.
- (3) **Dilute standard lead solution:** Dilute 10.0 ml of *standard lead solution* with sufficient 1 per cent v/v solution of nitric acid to produce 100 ml. Each ml of this solution contains 1 µg of lead per ml.
- (4) **Dithizone extraction solution:** Dissolve 30 mg of *diphenylthiocarbazon*e in 1000 ml of *chloroform* and add 5 ml of *alcohol*. Store the solution in a refrigerator. Before use, shake a suitable volume of the solution with about half its volume of 1 per cent v/v solution of *nitric acid* and discard the acid.
- (5) **Hydroxylamine hydrochloride solution Sp.:** Dissolve 20 g of *hydroxylamine hydrochloride* in sufficient *water* to produce about 65 ml. Transfer to separator, add five drops of *thymol blue solution*, add *strong ammonia solution* until the solution becomes yellow. Add 10 ml of a 4 per cent w/v solution of *sodium diethyldithiocarbamate* and allow to stand for five minutes. Extract with successive quantities, each of 10 ml, of *chloroform* until a 5 ml portion of the extract does not assume a yellow colour when shaken with dilute copper sulphate solution. Add *dilute*

hydrochloric acid until the solution is pink and then dilute with sufficient water to produce 100 ml.

- (6) **Potassium cyanide solution Sp.:** Dissolve 50 g of *potassium cyanide* in sufficient *water* to produce 100 ml. Remove the lead from this solution by extraction with successive quantities, each of 20 ml of *dithizone extraction solution* until the dithizone solution retains its orange-green colour. Extract any dithizone remaining in the cyanide solution by shaking with *chloroform*. Dilute this cyanide solution with sufficient *water* to produce a solution containing 10 g of *potassium cyanide* in each 100 ml.
- (7) **Standard dithizone solution:** Dissolve 10 ml of *diphenylthiocarbazone* in 1000 ml of *chloroform*. Store the solution in a glass-stoppered, lead-free bottle, protected from light and in a refrigerator.
- (8) **Citrate-cyanide wash solution:** To 50 ml of *water* add 50 ml of *ammonium citrate solution Sp.* and 4 ml of *potassium cyanide solution Sp.*, mix, and adjust the pH, if necessary, with strong *ammonia solution* to 9.0.
- (9) **Buffer solution pH 2.5:** To 25.0 ml of 0.2 M *potassium hydrogen phthalate* add 37.0 ml of 0.1 N *hydrochloric acid*, and dilute with sufficient *water* to produce 100.0 ml.
- (10) **Dithizone-carbon tetrachloride solution:**— Dissolve 10 mg of *diphenylthiocarbazone* in 1000 ml of carbon tetrachloride. Prepare this solution fresh for each determination.
- (11) **pH 2.5 wash solution:** To 500 ml of a 1 per cent v/v *nitric acid* add strong *ammonia solution* until the pH of the mixture is 2.5, then add 10 ml of *buffer solution* pH 2.5 and mix.
- (12) **Ammonia-cyanide wash solution:** To 35 ml of pH 2.5 *wash solution* add 4 ml of *ammonia-cyanide solution Sp.*, and mix.

Method

Transfer the volume of the prepared sample directed in the monograph to a separator and unless otherwise directed in monograph, add 6 ml of *ammonium citrate solution Sp.*, and 2 ml *hydroxylamine hydrochloride solution Sp.*, (For the determination of lead in iron salts use 10 ml of *ammonium citrate solution Sp.*). Add two drops of *phenol red solution* and make the solution just alkaline (red in colour) by the addition of *strong ammonia solution*. Cool the solution if necessary, and add 2 ml of *potassium cyanide solution Sp.* Immediately extract the solution with several quantities each of 5 ml, of *dithizone extraction solution*, draining off each extract into another separating funnel, until the dithizone extraction solution retains its green colour. Shake the combine dithizone solutions for 30 seconds with 30 ml of a 1 per cent w/v solution of *nitric acid* and discard the chloroform layer. Add to the solution exactly 5 ml of *standard dithizone solution* and 4 ml of *ammonia-cyanide solution Sp.* and shake for 30 seconds; the colour of the chloroform layer is of no deeper shade of violet than that of a control made with a volume of *dilute standard lead solution* equivalent to the amount of lead permitted in the sample under examination.

2.3.6. Limit Test for Sulphates:

Reagents

Barium Sulphate reagent: Mix 15 ml of 0.5 M *barium chloride*, 55 ml of *water*, and 20 ml of *sulphate free alcohol*, add 5 ml of a 0.0181 per cent w/v solution of potassium sulphate, dilute to 100 ml with *water*, and mix. Barium sulphate reagent must be freshly prepared.

0.5 M Barium Chloride: *Barium chloride* dissolved in *water* to contain in 1000 ml 122.1 g of $\text{BaCl}_2 \cdot 2\text{H}_2\text{O}$.

Method

Dissolve the specified quantity of the substance in *water*, or prepare a solution as directed in the text, transfer to a *Nessler cylinder*, and add 2 ml of *dilute hydrochloric acid*, except where *hydrochloric acid* is used in the preparation of the solution. Dilute to 45 ml with *water*, add 5 ml of barium sulphate reagent. Stir immediately with a glass rod, and allow to stand for five minutes. The turbidity produced is not greater than the *standard turbidity*, when viewed transversely. Standard turbidity : Place 1.0 ml of 0.1089 per cent w/v solution of potassium sulphate and 2 ml of *dilute hydrochloric acid* in a *Nessler cylinder*, dilute to 45 ml with *water*, add 5 ml of barium sulphate reagent, stir immediately with a glass rod and allow to stand for five minutes.

2.3.7. Heavy Metals by Atomic absorption spectrophotometry:

Atomic absorption spectrophotometry is used in the determination of heavy metal elements and some nonmetal elements in the atomic state.

The light of characteristic wave length emitted from a cathodic discharge lamp is absorbed when it passes through the atomic vapor generated from sample containing the element being examined atomized to the ground state. The assay of the element being examined is tested by determining the decreased degree of light intensity of radiation. Atomic absorption obeys the general rule for absorption spectrophotometry. The assay is carried out by comparing the absorbance of the test preparation with that of the reference preparation.

Apparatus

An atomic absorption spectrophotometer consists of a light source, an atomic generator, a monochromator and a detector system. Some are equipped with a background compensation system and automatic sampling system, etc.

1.Light Source: A hollow-cathode discharge lamp is usually used. The cathode is made of the element being examined.

2.Atomic Generator: There are four main types : flame atomizer, graphite furnace atomizer, hydride-generated atomizer, cold vapor atomizer.

(1) **Flame atomizer:** It mainly consists of a nebulizer and a burner. Its function is to nebulize the test solution into aerosol, which is mixed with combustion gas. And the mixture is introduced into the

flame generated by the burner. So that the substance being examined is to be dried, evaporated to form the ground state atoms of the element being examined. The burning flame is generated by different mixtures of gases, acetylene-air is mostly used. By modifying the proportion of combustion gas, the temperature of the flame can be controlled and a better stability and a better sensitivity can be obtained.

(2) **Furnace atomizer:** It consists of electric furnace and a power supply. Its function is to dry and incinerate the substance being examined. During the stage of high temperature atomization, the ground state atoms of the element being examined are to be formed. Graphite is commonly used as the heater. Protection gas is introduced into the furnace to avoid oxidation and used to transfer the sample vapor.

(3) **Hydride-generated atomizer:** It consists of hydride generator and atomic absorption cell. It is used for the determination of the elements such as arsenic, selenium and antimony etc. Its function is to reduce the element to be examined in acidic medium to the low-boiling and easily pyrolyzed hydride. The hydride is then swept by a stream of carrier gas into the atomic absorption cell which consists of quartz tube and heater etc., in which the hydride is pyrolyzed by heating to form the ground-state atom.

(4) **Cold vapor atomizer:** It consists of a mercury vapor atomizer and an absorption cell. It is suitable for the determination of mercury. Its function is to reduce the mercuric ion into mercury vapor which is swept into the quartz absorption cell by carrier gas.

3. Monochromator: Its function is to separate the specified wavelength radiation from the electromagnetic radiations erradiated from the light source. The optical path of the apparatus should assure the good spectra resolution and has the ability to work well at the condition of narrow spectral band (0.2 nm). The commonly used wavelength region is 190.0 - 900.0 nm.

4. Detector system: It consists of a detector, a signal processor and a recording system. It should have relatively higher sensitivity and better stability and can follow the rapid change of the signal absorption.

5. Background compensation system: System employed for the correction of atmospheric effects on the measuring system. Four principles can be utilized for background compensation: continuous spectrum sources (a deuterium lamp is often used in the UV region), the Zeeman effect, the self inversion phenomenon and the non resonance spectrum. In the analysis using atomic absorption spectrophotometry, the interference to the determination caused by background and other reasons should be noticed. Changes of some experimental conditions, such as the wavelength, the slit width, the atomizing condition, etc., may affect the sensitivity, the stability and the interference. If it is flame, the suitable wavelength, slit width and flame temperature, the addition of complexing agents and releasing agents and the use of Standard addition method may eliminate interference. If it is furnace, system, the selection of suitable background compensation system and the addition of suitable matrix modifying agents, etc may remove the interference. Background compensation method shall be selected as specified in the individual monograph.

Procedure

Method (direct calibration method)

Prepare not less than 3 reference solutions of the element being examined of different concentrations, covering the range recommended by the instrument manufacturer and add separately the corresponding reagents as that for the test solution and prepare the blank reference solution with the corresponding reagents. Measure the absorbances of the blank reference solution and each reference solution of different concentrations separately, record the readings and prepare a calibration curve with the average value of 3 readings of each concentration on the ordinate and the corresponding concentration on the abscissa.

Prepare a test solution of the substance being examined as specified in the monograph, adjust the concentration to fall within the concentration range of the reference solution. Measure the absorbance 3 times, record the readings and calculate the average value. Interpolate the mean value of the readings on the calibration curve to determine the concentration of the element.

When used in the test for impurities, prepare two test preparations of the same concentration as specified in the monograph. To one of the test preparation add an amount of the reference substance equivalent to the limit of the element specified in the monograph. Proceed as directed above and measure this solution to give an appropriate reading a; then measure the test preparation without the addition of the reference substance under the same condition and record the reading b; b is not greater than (a-b).

Determination of Lead, Cadmium, Arsenic, Mercury and Copper:

(1) Determination of lead (graphite oven method):

Determination conditions Reference condition: dry temperature: 100-120⁰, maintain 20 seconds; ash temperature: 400-750⁰, maintain 20-25 seconds; atomic temperature: 1700-2100⁰, maintain 4-5 seconds; measurement wavelength: 283.3 nm; background calibration: deuterium lamp (D lamp) or Zeeman effect.

Preparation of lead standard stock solution: Measure accurately a quantity of lead single-element standard solution to prepare standard stock solution with 2 per cent nitric acid solution, which containing 1 µg per ml, stored at 0-5⁰.

Preparation of calibration curve: Measure accurately a quantity of lead standard stock solutions respectively, diluted with 2 per cent nitric acid solution to the concentration of 0, 5, 20, 40, 60, 80 ng per ml, respectively. Measure respectively accurately 1 ml the above solution, add respectively 1 ml of 1 per cent ammonium dihydrogen phosphate and 0.2 per cent *magnesium nitrate* mix well, pipette accurately 20 µl to inject into the atomic generator of graphite oven and determine their absorbance, then draw the calibration curve with absorbance as vertical axis and concentration as horizontal ordinate.

Preparation of test solution:

Method: Weigh accurately 0.5 g of the coarse powder of the substance being examined, transfer into a casparian flask, add 5-10 ml of the mixture of *nitric acid* and *perchloric acid* (4 : 1), add a small hopper on the flask-top, macerate overnight, heat to slake on the electric hot plate, keep somewhat-boiling, if brownish-black, add again a quantity of the above mixture, continuously heat till the solution becomes clean and transparent, then raise temperature, heat continuously to thick smoke, till white smoke disperse, the slaked solution becomes colourless and transparent or a little yellow, cool, transfer it into a 50 ml volumetric flask, wash the container with 2 per cent *nitric acid solution* add the washing solution into the same volumetric flask and dilute with the same solvent to the volume, shake well. Prepare synchronously the reagent blank solution according to the above procedure.

Determination: Measure accurately 1 ml of the test solution and its corresponding reagent blank solution respectively, add 1 ml of solution containing 1per cent *ammonium dihydrogen phosphate* and 0.2 per cent *magnesium nitrate*, shake well, pipette accurately 10-20 μl to determine their absorbance according to the above method of “Preparation of calibration curve”. Calculate the content of lead (Pb) in the test solution from the calibration curve.

(2) Determination of cadmium (Cd) (graphite oven method):

Determination conditions Reference condition: dry temperature: 100-120⁰, maintain 20 seconds; ash temperature: 300-500⁰, maintain 20-25 seconds; atomic temperature: 1500-1900⁰, maintain 4-5 seconds; measurement wavelength: 228.8 nm; background calibration: deuterium lamp (D lamp) or Zeeman effect.

Preparation of Cd standard stock solution: Measure accurately a quantity of Cd single-element standard solution to prepare standard stock solution Cd with 2 per cent nitric acid, which containing 0.4 μg per ml Cd, stored at 0-5⁰.

Preparation of calibration curve: Measure accurately a quantity of cadmium standard stock solutions, diluted to the concentration of 1.6, 3.2, 4.8, 6.4 and 8.0 ng per ml with 2 per cent nitric acid, respectively. Pipette accurately 10 μl the above solutions respectively, inject them into the graphite oven, determine their absorbance, and then draw the calibration curve with absorbance as vertical axis and concentration as horizontal ordinate.

Preparation of test solution: Reference to “Preparation of test solution” of Pb in the above.

Determination: Pipette accurately 10-20 μl of the test solution and its corresponding reagent blank solution respectively, determine their absorbance according to the above method of “Preparation of calibration curve. If interference occurs, weigh accurately respectively 1 ml of the standard solution, blank solution and test solution, add 1 ml of a solution containing 1per cent *ammonium dihydrogen phosphate* and 0.2 per cent *magnesium nitrate*, shake well, determine their absorbance according to the method above, calculate the content of Cd in the test solution from the calibration curve.

(3) Determination of Arsenic (As) (hydride method):

Determination conditions Apparatus: suitable hydride generator device, reducing agent: a solution containing 1 per cent *sodium borohydride* and 0.3 per cent *sodium hydroxide*; carrier liquid: 1 per cent *hydrochloric acid*; carrier gas: nitrogen; measurement wavelength: 193.7 nm; background calibration: deuterium lamp (D lamp) or Zeeman effect.

Preparation of As standard stock solution: Measure accurately a quantity of As single-element standard solution to prepare standard stock solution with 2 per cent *nitric acid* solution, which contains 1.0 μg per ml As, stored at 0-5⁰.

Preparation of calibration curve: Measure accurately proper quantity of arsenic standard stock solutions, diluted with 2 per cent *nitric acid* to the concentration of 2, 4, 8, 12 and 16 ng per ml respectively. Accurately transfer 10 ml of each into 25 ml volumetric flask respectively, add 1 ml of 25 per cent *potassium iodide solution* (prepared prior to use), shake well, add 1 ml of *ascorbic acid solution* (prepared prior to use), shake well, dilute with hydrochloric acid solution (20-100) to the volume, shake well, close the stopper and immerse the flask in a water bath at 80⁰ for 3 minutes. Cool, transfer proper quantities of each solution respectively into the hydride generator device, determine the absorbance, then plot the calibration curve with peak area (absorbance) as vertical axis and concentration as horizontal ordinate.

Preparation of test solution: Reference to A or B method of "Preparation of test solution" of Pb in the above.

Determination: Pipette accurately 10 ml of the test solution and its corresponding reagent blank solution respectively, proceed as described under "Preparation of calibration curve" beginning at the words "add 1 ml of 25 per cent *potassium iodide solution*". Calculate the content of As in the test solution from the calibration curve.

(4) Determination of Mercury (Hg) (cold absorption method):

Determination conditions: Apparatus: suitable hydride generator device; reducing agent: a solution containing 0.5 per cent *sodium borohydride* and 0.1 per cent *sodium hydroxide*; carrier liquid: 1 per cent *hydrochloric acid*; carrier gas: nitrogen; measurement wavelength: 253.6 nm; background calibration: deuterium lamp (D lamp) or Zeeman effect.

Preparation of mercury standard stock solution: Measure accurately a proper quantity of mercury single-element standard solution to prepare standard stock solution with 2 per cent nitric acid solution, which containing 1.0 μg per ml Hg, stored at 0-5⁰.

Preparation of calibration curve: Measure accurately 0, 0.1, 0.3, 0.5, 0.7 and 0.9 ml of mercury standard stock solution, transfer into a 50 ml volumetric flask respectively, add 40 ml 4 per cent *sulphuric acid solution* and 0.5 ml of 5 per cent *potassium permanganate solution*, shake well, drop 5 per cent *hydroxylamine hydrochloride solution* until the violet red just disappears, dilute with 4 per cent *sulfuric acid solution* to the volume, shake well. A quantity of each solution is injected to the

hydride generator device, determine the absorbance, then plot the calibration curve with peak area (absorbance) as vertical axis and concentration as horizontal ordinate.

Preparation of test solution:

Method : Transfer 1 g of the coarse powder of the substance being examined, accurately weighed, into a casparian flask, add 5-10 ml of the mixture solution of *nitric acid* and *perchloric acid* (4 : 1), mix well, fix a small hopper on the flask-top, immerse overnight, heat to slake on the electric hot plate at 120-140⁰ for 4-8 hours until *slaking* completely, cool, add a quantity of 4 per cent *sulfuric acid solution* and 0.5 ml of 5 per cent *potassium permanganate solution*, shake well, drop 5 per cent *hydroxylamine hydrochloride solution* until the violet red colour just disappears, dilute with 4 per cent *sulphuric acid solutions* to 25 ml, shake well, centrifugate if necessary, the supernatant is used as the test solution. Prepare synchronally the reagent blank solute based on the same procedure.

Determination: Pipette accurately a quantity of the test solution and its corresponding reagent blank solution, respectively, proceed as described under “Preparation of calibration curve” beginning at the words “add 1 ml of 25 per cent *potassium iodide solution*”. Calculate the content of mercury (Hg) in the test solution from the calibration curve.

(5) Determination of Copper (flame method):

Determination conditions: Measurement wavelength: 324.7 nm; flame: air -acetylene flame; background calibration: deuterium lamp or Zeeman effect.

Preparation of copper standard stock solution: Measure accurately a proper quantity of copper single-element standard solution, to prepare the standard stock solution with 2 per cent *nitric acid solution*, which containing 10 µg per ml Cu, stored at 0-5⁰.

Preparation of calibration curve: Measure accurately a quantity of copper standard stock solutions, dilute with 2 per cent *nitric acid* to the concentrations of 0.05, 0.2, 0.4, 0.6 and 0.8 µg per ml, respectively. Inject each standard solution into the flame and determine the absorbance, respective, then plot the calibration curve with absorbance as vertical axis and concentration as horizontal ordinate.

Preparation of test solution: Reference to “Preparation of test solution” of Pb in the above.

Determination: Pipette accurately quantities of the test solution and its corresponding reagent blank solution respectively, proceed as described under “Preparation of calibration curve”. Calculate the content of Cu in the test solution from the calibration curve.

2.4. Microbial Limit Tests:

The following tests are designed for the estimation of the number of viable aerobic micro-organisms present and for detecting the presence of designated microbial species in pharmaceutical substances. The term ‘growth’ is used to designate the presence and presumed proliferation of viable micro-organisms.

Preliminary Testing

The methods given herein are invalid unless it is demonstrated that the test specimens to which they are applied do not, of themselves, inhibit the multiplication under the test conditions of micro-organisms that can be present. Therefore, prior to doing the tests, inoculate diluted specimens of the substance being examined with separate viable cultures of *Escherichia coli*, *Salmonella* species, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. This is done by adding 1 ml of not less than 10^{-3} dilutions of a 24 h broth culture of the micro-organisms to the first dilution (in buffer solution pH 7.2, fluid soyabean-casein digest medium or fluid lactose medium) of the test material and following the test procedure. If the organisms fail to grow in the relevant medium the procedure should be modified by (a) increasing the volume of diluent with the quantity of test material remaining the same, or (b) incorporating a sufficient quantity of a suitable inactivating agent in the diluents, or (c) combining the aforementioned modifications so as to permit growth of the organisms in the media. If inhibitory substances are present in the sample, 0.5 per cent of soya lecithin and 4 per cent of polysorbate 20 may be added to the culture medium. Alternatively, repeat the test as described in the previous paragraph, using fluid casein digest-soya lecithin-polysorbate 20 medium to demonstrate neutralization of preservatives or other antimicrobial agents in the test material. Where inhibitory substances are contained in the product and the latter is soluble, the Membrane filtration method described under Total Aerobic Microbial Count may be used.

If in spite of incorporation of suitable inactivating agents and a substantial increase in the volume of diluent it is still not possible to recover the viable cultures described above and where the article is not suitable for applying the membrane filtration method it can be assumed that the failure to isolate the inoculated organism may be due to the bactericidal activity of the product. This may indicate that the article is not likely to be contaminated with the given species of micro-organisms. However, monitoring should be continued to establish the spectrum of inhibition and bactericidal activity of the article.

Media

Culture media may be prepared as given below or dehydrated culture media may be used provided that, when reconstituted as directed by the manufacturer, they have similar ingredients and / or yield media comparable to those obtained from the formulae given below.

Where agar is specified in a formula, use agar that has a moisture content of not more than 15 per cent. Where water is called for in a formula, use purified water. Unless otherwise indicated, the media should be sterilized by heating in an autoclave at 115° for 30 minutes.

In preparing media by the formulas given below, dissolve the soluble solids in the water, using heat if necessary, to effect complete solution and add solutions of hydrochloric acid or sodium hydroxide in quantities sufficient to yield the required pH in the medium when it is ready for use. Determine the pH at $25^{\circ} \pm 2^{\circ}$.

Baird-Parker Agar Medium

Pancreatic digest of casein	10.0 g
Beef extract	5.0 g
Yeast extract	1.0 g
Lithium chloride	5.0 g
Agar	20.0 g
Glycine	12.0 g
Sodium pyruvate	10.0 g
Water to	1000 ml

Heat with frequent agitation and boil for 1 minute. Sterilise, cool to between 45° and 50°, and add 10 ml of a one per cent w/v solution of sterile *potassium tellurite* and 50 ml of egg-yolk emulsion. Mix intimately but gently and pour into plates. (Prepare the egg-yolk emulsion by disinfecting the surface of whole shell eggs, aseptically cracking the eggs, and separating out intact yolks into a sterile graduated cylinder. Add sterile saline solution, get a 3 to 7 ratio of egg-yolk to saline. Add to a sterile blender cup, and mix at high speed for 5 seconds). Adjust the pH after sterilization to 6.8 ± 0.2.

Bismuth Sulphite Agar Medium

Solution (1)

Beef extract	6 g
Peptone	10 g
Agar	24 g
Ferric citrate	0.4 g
Brilliant green	10 mg
Water to	1000 ml

Dissolve with the aid of heat and sterilise by maintaining at 115° for 30 minutes.

Solution (2)

Ammonium bismuth citrate	3 g
Sodium sulphite	10 g
Anhydrous disodium hydrogen Phosphate	5 g
Dextrose monohydrate	5 g
Water to	100 ml

Mix, heat to boiling, cool to room temperature, add 1 volume of solution (2) to 10 volumes of solution (1) previously melted and cooled to a temperature of 55° and pour. Bismuth Sulphite Agar Medium should be stored at 2° to 8° for 5 days before use.

Brilliant Green Agar Medium

Peptone	10.0 g
Yeast extract	3.0 g
Lactose	10.0 g
Sucrose	10.0 g
Sodium chloride	5.0 g
Phenol red	80.0 g
Brilliant green	12.5 mg
Agar	12.0 g
Water to	1000 ml

Mix, allow to stand for 15 minutes, sterilise by maintaining at 115⁰ for 30 minutes and mix before pouring.

Buffered Sodium Chloride-Peptone Solution pH 7.0

Potassium dihydrogen phosphate	3.56 g
Disodium hydrogen phosphate	7.23 g
Sodium chloride	4.30 g
Peptone (meat or casein)	1.0 g
Water to	1000 ml

0.1 to 1.0 per cent w/v polysorbate 20 or polysorbate 80 may be added. Sterilise by heating in an autoclave at 121⁰ for 15 minutes.

Casein Soyabean Digest Agar Medium

Pancreatic digest of casein	15.0 g
Papaic digest of soyabean meal	5.0 g
Sodium chloride	5.0 g
Agar	15.0 g
Water to	1000 ml

Adjust the pH after sterilization to 7.3 ± 0.2.

Cetrimide Agar Medium

Pancreatic digest of gelatin	20.0 g
Magnesium chloride	1.4 g
Potassium sulphate	10.0 g
Cetrimide	0.3 g
Agar	13.6 g
Glycerin	10.0 g

Water to 1000 ml

Heat to boiling for 1 minute with shaking. Adjust the pH so that after sterilization it is 7.0 to 7.4. Sterilise at 121⁰ for 15 minutes.

Desoxycholate-Citrate Agar Medium

Beef extract	5.0 g
Peptone	5.0 g
Lactose	10.0 g
Trisodium citrate	8.5 g
Sodium thiosulphate	5.4 g
Ferric citrate	1.0 g
Sodium desoxycholate	5.0 g
Neutral red	0.02 g
Agar	12.0 g
Water to	1000 ml

Mix and allow to stand for 15 minutes. With continuous stirring, bring gently to the boil and maintain at boiling point until solution is complete. Cool to 80⁰, mix, pour and cool rapidly.

Care should be taken not to overheat Desoxycholate Citrate Agar during preparation. It should not be remelted and the surface of the plates should be dried before use.

Fluid Casein Digest-Soya Lecithin-Polysorbate 20 Medium

Pancreatic digest of casein	20 g
Soya lecithin	5 g
Polysorbate 20	40 ml
Water to	1000 ml

Dissolve the pancreatic digest of casein and soya lecithin in water, heating in a water-bath at 48⁰ to 50⁰ for about 30 minutes to effect solution. Add polysorbate 20, mix and dispense as desired.

Fluid Lactose Medium

Beef extract	3.0 g
Pancreatic digest of gelatin	5.0 g
Lactose	5.0 g
Water to	1000 ml

Cool as quickly as possible after sterilization. Adjust the pH after sterilization to 6.9 ± 0.2.

Lactose Broth Medium

Beef extract	3.0 g
Pancreatic digest of gelatin	5.0 g
Lactose	5.0 g
Water to	1000 ml

Adjust the pH after sterilisation to 6.9 ± 0.2 .

Levine Eosin-Methylene Blue Agar Medium

Pancreatic digest of gelatin	10.0 g
Dibasic potassium phosphate	2.0 g
Agar	15.0 g
Lactose	10.0 g
Eosin Y	400 mg
Methylene blue	65 mg
Water to	1000 ml

Dissolve the pancreatic digest of gelatin, dibasic potassium phosphate and agar in water with warming and allow to cool. Just prior to use, liquefy the gelled agar solution and the remaining ingredients, as solutions, in the following amounts and mix. For each 100 ml of the liquefied agar solution use 5 ml of a 20 per cent w/v solution of lactose, and 2 ml of a 2 per cent w/v solution of eosin Y, and 2 ml of a 0.33 per cent w/v solution of methylene blue. The finished medium may not be clear. Adjust the pH after sterilisation to 7.1 ± 0.2 .

MacConkey Agar Medium

Pancreatic digest of gelatin	17.0g
Peptone (meat and casein, equal parts)	3.0 g
Lactose	10.0 g
Sodium chloride	5.0 g
Bile salts	1.5 g
Agar	13.5 g
Neutral red	30 mg
Crystal violet	1 mg
Water to	1000 ml

Boil the mixture of solids and water for 1 minute to effect solution. Adjust the pH after sterilisation to 7.1 ± 0.2 .

MacConkey Broth Medium

Pancreatic digest of gelatin	20.0g
------------------------------	-------

Lactose	10.0 g
Dehydrated ox bile	5.0 g
Bromocresol purple	10 mg
Water to	1000 ml

Adjust the pH after sterilisation to 7.3 ± 0.2 .

Mannitol-Salt Agar Medium

Pancreatic digest of gelatin	5.0 g
Peptic digest of animal tissue	5.0 g
Beef extract	1.0 g
D-Mannitol	10.0 g
Sodium chloride	75.0 g
Agar	15.0 g
Phenol red	25 mg
Water to	1000 ml

Mix, heat with frequent agitation and boil for 1 minute to effect solution. Adjust the pH after sterilisation to 7.4 ± 0.2 .

Nutrient Agar Medium : Nutrient broth gelled by the addition of 1 to 2 per cent w/v of agar.

Nutrient Broth Medium

Beef extract	10.0 g
Peptone	10.0 g
Sodium chloride	5 mg
Water to	1000 ml

Dissolve with the aid of heat. Adjust the pH to 8.0 to 8.4 with *5M sodium hydroxide* and boil for 10 minutes. Filter, and sterilise by maintaining at 115° for 30 minutes and adjust the pH to 7.3 ± 0.1 .

Pseudomonas Agar Medium for Detection of Flourescein

Pancreatic digest of casein	10.0 g
Peptic digest of animal tissue	10.0 g
Anhydrous dibasic potassium phosphate	1.5 g
Magnesium sulphate hepta hydrate	1.5 g
glycerin	10.0 ml
Agar	15.0 g
Water to	1000 ml

Dissolve the solid components in water before adding glycerin. Heat with frequent agitation and boil for 1 minute to effect solution. Adjust the *pH* after sterilisation to 7.2 ± 0.2 .

Pseudomonas Agar Medium for Detection of Pyocyanin

Pancreatic digest of gelatin	20.0 g
Anhydrous magnesium chloride	1.4 g
Anhydrous potassium sulphate	10.0 g
Agar	15.0 g
Glycerin	10.0 ml
Water to	1000 ml

Dissolve the solid components in water before adding glycerin. Heat with frequent agitation and boil for 1 minute to effect solution. Adjust the *pH* after sterilisation to 7.2 ± 0.2 .

Sabouraud Dextrose Agar Medium

Dextrose	40 g
Mixture of equal parts of peptic digest of animal tissue and Pancreatic digest of casein	10 g
Agar	15 g
Water to	1000 ml

Mix, and boil to effect solution. Adjust the *pH* after sterilisation to 5.6 ± 0.2 .

Sabouraud Dextrose Agar Medium with Antibiotics

To 1 liter of Sabouraud Dextrose Agar Medium add 0.1 g of benzylpenicillin sodium and 0.1 g of tetracycline or alternatively add 50 mg of chloramphenicol immediately before use.

Selenite F Broth

Peptone	5 g
Lactose	4 g
Disodium hydrogen phosphate	10 g
Sodium hydrogen selenite	4 g
Water to	1000 ml

Dissolve, distribute in sterile containers and sterilise by maintaining at 100° for 30 minutes.

Fluid Selenite-Cystine Medium

Pancreatic digest of casein	5.0 g
Lactose	4.0 g
Sodium phosphate	10.0 g

Sodium hydrogen selenite	4.0 g
L-Cystine	10.0 mg
Water to	1000 ml

Mix and heat to effect solution. Heat in flowing steam for 15 minutes. Adjust the final *pH* to 7.0 ± 0.2 . Do not sterilise.

Tetrathionate Broth Medium

Beef extract	0.9 g
Peptone	4.5 g
Yeast extract	1.8 g
Sodium chloride	4.5 g
Calcium carbonate	25.0 g
Sodium thiosulphate	40.7 g
Water to	1000 ml

Dissolve the solids in water and heat the solution to boil. On the day of use, add a solution prepared by dissolving 5 g of potassium iodide and 6 g of iodine in 20 ml of water.

Tetrathionate-Bile-Brilliant Green Broth Medium

Peptone	8.6 g
Dehydrated ox bile	8.0 g
Sodium chloride	6.4 g
Calcium carbonate	20.0 g
Potassium tetrathionate	20.0 g
Brilliant green	70 mg
Water to	1000 ml

Heat just to boiling; do not reheat. Adjust the *pH* so that after heating it is 7.0 ± 0.2 .

Triple Sugar-Iron Agar Medium

Mix, allow standing for 15 minutes, bringing to boil and maintain at boiling point until solution is complete, mix, distributing in tubes and sterilising by maintaining at 115° for 30 minutes. Allow to stand in a sloped form with a butt about 2.5 cm long.

Urea Broth Medium

Potassium dihydrogen orthophosphate	9.1 g
Anhydrous disodium hydrogen phosphate	9.5 g
Urea	20.0 g
Yeast extract	0.1 g

Phenol red	10 mg
Water to	1000 ml

Mix, sterilise by filtration and distribute aseptically in sterile containers.

Vogel-Johnson Agar Medium

Pancreatic digest of casein	10.0 g
Yeast extract	5.0 g
Mannitol	10.0 g
Dibasic potassium phosphate	5.0 g
Lithium chloride	5.0 g
glycerin	10.0 g
Agar	16.0 g
Phenol red	25.0 mg
Water to	1000 ml

Boil the solution of solids for 1 minute. Sterilise, cool to between 45⁰ to 50⁰ and add 20 ml of a 1 per cent w/v sterile solution of potassium tellurite. Adjust the pH after sterilisation to 7.0±0.2.

Xylose-Lysine-Desoxycholate Agar Medium

Xylose	3.5 g
L-Lysine	5.0 g
Lactose	7.5 g
Sucrose	7.5 g
Sodium chloride	5.0 g
Yeast extract	3.0 g
Phenol red	80 mg
Agar	13.5 g
Sodium desoxycholate	2.5 g
Sodium thiosulphate	6.8 g
Ferric ammonium citrate	800 mg
Water to	1000 ml

Heat the mixture of solids and water, with swirling, just to the boiling point. Do not overheat or sterilise. Transfer at once to a water-bath maintained at about 50⁰ and pour into plates as soon as the medium has cooled. Adjust the final pH to 7.4 ± 0.2.

Sampling : Use 10 ml or 10 g specimens for each of the tests specified in the individual monograph.

Precautions : The microbial limit tests should be carried out under conditions designed to avoid accidental contamination during the test. The precautions taken to avoid contamination must be such that they do not adversely affect any micro-organisms that should be revealed in the test.

2.4.1. Total Aerobic Microbial Count:

Pretreat the sample of the product being examined as described below.

Water-soluble products : Dissolve 10 g or dilute 10 ml of the preparation being examined, unless otherwise specified, in buffered sodium chloride-peptone solution *pH* 7.0 or any other suitable medium shown to have no antimicrobial activity under the conditions of test and adjust the volume to 100 ml with the same medium. If necessary, adjust the *pH* to about 7.

Products insoluble in Water (non-fatty): Suspend 10 g or 10 ml of the preparation being examined, unless otherwise specified, in buffered sodium chloride-peptone solution *pH* 7.0 or any other suitable medium shown not to have antimicrobial activity under the conditions of the test and dilute to 100 ml with the same medium. If necessary, divide the preparation being examined and homogenize the suspension mechanically.

A suitable surface-active agent such as 0.1 per cent w/v of polysorbate 80 may be added to assist the suspension of poorly wettable substances. If necessary, adjust the *pH* of the suspension to about 7.

Fatty products: Homogenise 10 g or 10 ml of the preparation being examined, unless otherwise specified, with 5 g of polysorbate 20 or polysorbate 80. If necessary, heat to not more than 40°. Mix carefully while maintaining the temperature in the water-bath or in an oven. Add 85 ml of buffered sodium chloride-peptone solution *pH* 7.0 or any other suitable medium shown to have no antimicrobial activity under the conditions of the test, heated to not more than 40° if necessary. Maintain this temperature for the shortest time necessary for formation of an emulsion and in any case for not more than 30 minutes. If necessary, adjust the *pH* to about 7.

Examination of the sample: Determine the total aerobic microbial count in the substance being examined by any of the following methods.

Membrane filtration : Use membrane filters 50 mm in diameter and having a nominal pore size not greater than 0.45 µm the effectiveness of which in retaining bacteria has been established for the type of preparation being examined.

Transfer 10 ml or a quantity of each dilution containing 1 g of the preparation being examined to each of two membrane filters and filter immediately. If necessary, dilute the pretreated preparation so that a colony count of 10 to 100 may be expected. Wash each membrane by filtering through it three or more successive quantities, each of about 100 ml, of a suitable liquid such as *buffered sodium chloride-peptone solution pH 7.0*. For fatty substances add to the liquid *polysorbate 20* or *polysorbate 80*. Transfer one of the membrane filters, intended for the enumeration of bacteria, to the surface of a

plate of *casein soyabean digest agar* and the other, intended for the enumeration of fungi, to the surface of a plate of *Sabouraud dextrose agar* with antibiotics.

Incubate the plates for 5 days, unless a more reliable count is obtained in shorter time, at 30⁰ to 35⁰ in the test for bacteria and 20⁰ to 25⁰ in the test for fungi. Count the number of colonies that are formed. Calculate the number of micro-organisms per g or per ml of the preparation being examined, if necessary counting bacteria and fungi separately.

Plate count for bacteria: Using Petri dishes 9 to 10 cm in diameter, add to each dish a mixture of 1 ml of the pretreated preparation and about 15 ml of liquefied *casein soyabean digest agar* at not more than 45⁰. Alternatively, spread the pretreated preparation on the surface of the solidified medium in a Petri dish of the same diameter. If necessary, dilute the pretreated preparation as described above so that a colony count of not more than 300 may be expected. Prepare at least two such Petri dishes using the same dilution and incubate at 30⁰ to 35⁰ for 5 days, unless a more reliable count is obtained in a shorter time. Count the number of colonies that are formed. Calculate the results using plates with the greatest number of colonies but taking 300 colonies per plate as the maximum consistent with good evaluation.

Plate count for fungi: Proceed as described in the test for bacteria but use *Sabouraud dextrose agar with antibiotics* in place of *casein soyabean digest agar* and incubate the plates at 20⁰ to 25⁰ for 5 days, unless a more reliable count is obtained in a shorter time. Calculate the results using plates with not more than 100 colonies.

Multiple-tube or serial dilution method : In each of fourteen test-tubes of similar size place 9.0 ml of sterile *fluid soyabean casein digest medium*. Arrange twelve of the tubes in four sets of three tubes each. Put aside one set of three tubes to serve as controls. Into each of three tubes of one set (“100”) and into fourth tube (A) pipette 1 ml of the solution of suspension of the test specimen and mix. From tube A pipette 1 ml of its contents into the one remaining tube (B) not included in the set and mix. These two tubes contain 100 mg (or 100 µl) and 10 mg (or 10 µl) of the specimen respectively. Into each of the second set (“10”) of three tubes pipette 1 ml from tube

Table 5 – Most Probable Total Count by Multiple-Tube Or Serial Dilution Method

Observed combination of numbers of tubes showing growth in each set			Most probable number of micro-organisms per g or per ml
No. of mg (or ml) of specimen per tube			
100 (100 µl)	10 (10 µl)	1 (1 µl)	

3	3	3	>1100
3	3	2	1100
3	3	1	500
3	3	0	200
3	2	3	290
3	2	2	210
3	2	1	150
3	2	0	90
3	1	3	160
3	1	2	120
3	1	1	70
3	1	0	40
3	0	3	95
3	0	2	60
3	0	1	40
3	0	0	23

A, and into each tube of the third set (“1”) pipette 1 ml from tube B. Discard the unused contents of tube A and B. Close well and incubate all of the tubes. Following the incubation period, examine the tubes for growth. The three control tubes remain clear. Observations in the tubes containing the test specimen, when interpreted by reference to Table 1, indicate the most probable number of micro-organisms per g or per ml of the test specimen.

2.4.2. Tests for Specified Micro-organisms:

Pretreatment of the sample being examined: Proceed as described under the test for total aerobic microbial count but using lactose broth or any other suitable medium shown to have no antimicrobial activity under the conditions of test in place of buffered sodium chloride-peptone solution pH 7.0.

Escherichia coli : Place the prescribed quantity in a sterile screw-capped container, add 50 ml of nutrient broth, shake, allow to stand for 1 hour (4 hours for gelatin) and shake again. Loosen the cap and incubate at 37⁰ for 18 to 24 hours.

Primary test: Add 1.0 ml of the enrichment culture to a tube containing 5 ml of MacConkey broth. Incubate in a water-bath at 36⁰ to 38⁰ for 48 hours. If the contents of the tube show acid and gas carry out the secondary test.

Secondary test: Add 0.1 ml of the contents of the tubes containing (a) 5 ml of MacConkey broth, and (b) 5 ml of peptone water. Incubate in a water-bath at 43.5⁰ to 44.5⁰ for 24 hours and examine tube (a) for acid and gas and tube (b) for indole. To test for indole, add 0.5 ml of Kovac’s reagent,

shake well and allow to stand for 1 minute; if a red colour is produced in the reagent layer indole is present. The presence of acid and gas and of indole in the secondary test indicates the presence of *Escherichia coli*.

Carry out a control test by repeating the primary and secondary tests adding 1.0 ml of the enrichment culture and a volume of broth containing 10 to 50 *Escherichia coli* (NCTC 9002) organisms, prepared from a 24-hour culture in nutrient broth, to 5 ml of MacConkey broth. The test is not valid unless the results indicate that the control contains *Escherichia coli*.

Alternative test: By means of an inoculating loop, streak a portion from the enrichment culture (obtained in the previous test) on the surface of MacConkey agar medium. Cover and invert the dishes and incubate. Upon examination, if none of the colonies are brick-red in colour and have a surrounding zone of precipitated bile the sample meets the requirements of the test for the absence of *Escherichia coli*.

If the colonies described above are found, transfer the suspect colonies individually to the surface of Levine eosin-methylene blue agar medium, plated on Petri dishes. Cover and invert the plates and incubate. Upon examination, if none of the colonies exhibits both a characteristic metallic sheen under reflected light and a blue-black appearance under transmitted light, the sample meets the requirements of the test for the absence of *Escherichia coli*. The presence of *Escherichia coli* may be confirmed by further suitable cultural and biochemical tests.

Salmonella : Transfer a quantity of the pretreated preparation being examined containing 1 g or 1 ml of the product to 100 ml of nutrient broth in a sterile screw-capped jar, shake, allow to stand for 4 hours and shake again. Loosen the cap and incubate at 35⁰ to 37⁰ for 24 hours.

Primary test: Add 1.0 ml of the enrichment culture to each of the two tubes containing (a) 10 ml of selenite F broth and (b) tetrathionate-bile-brilliant green broth and incubate at 36⁰ to 38⁰ for 48 hours. From each of these two cultures subculture on at least two of the following four agar media: bismuth sulphate agar, brilliant green agar, deoxycholatecitrate agar and xylose-lysine-deoxycholate agar. Incubate the plates at 36⁰ to 38⁰ for 18 to 24 hours. Upon examination, if none of the colonies conforms to the description given in Table 2, the sample meets the requirements of the test for the absence of the genus *Salmonella*.

If any colonies conforming to the description in Table 2 are produced, carry out the secondary test.

Secondary test: Subculture any colonies showing the characteristics given in Table 2 in triple sugar-iron agar by first inoculating the surface of the slope and then making a stab culture with the same inoculating needle, and at the same time inoculate a tube of urea broth. Incubate at 36⁰ to 38⁰ for 18 to 24 hours. The formation of acid and gas in the stab culture (with or without concomitant blackening) and the absence of acidity from the surface growth in the triple sugar iron agar, together with the absence of a red colour in the urea broth, indicate the presence of *Salmonella*. If acid but no gas is produced in the stab culture, the identity of the organisms should be confirmed by agglutination tests.

Carry out the control test by repeating the primary and secondary tests using 1.0 ml of the enrichment culture and a volume of broth containing 10 to 50 *Salmonella abony* (NCTC 6017) organisms, prepared from a 24-hour culture in nutrient broth, for the inoculation of the tubes (a) and (b). The test is not valid unless the results indicate that the control contains *Salmonella*.

Table 6 – Test for *Salmonella*

Medium	Description of colony
Bismuth sulphite agar	Black or green
Brilliant green agar	Small, transparent and colourless, or opaque, pinkish or white (frequently surrounded by a pink or red zone)
Deoxycholate-citrate agar	Colourless and opaque, with or without black centers
Xylose-lysine-desoxy-cholate agar	Red with or without black centres

Pseudomonas aeruginosa : Pretreat the preparation being examined as described above and inoculate 100 ml of fluid soyabean-casein digest medium with a quantity of the solution, suspension or emulsion thus obtained containing 1 g or 1 ml of the preparation being examined. Mix and incubate at 35⁰ to 37⁰ for 24 to 48 hours. Examine the medium for growth and if growth is present, streak a portion of the medium on the surface of cetrimide agar medium, each plated on Petri dishes. Cover and incubate at 35⁰ to 37⁰ for 18 to 24 hours.

If, upon examination, none of the plates contains colonies having the characteristics listed in Table 3 for the media used, the sample meets the requirement for freedom from *Pseudomonas aeruginosa*. If any colonies conforming to the description in Table 3 are produced, carry out the oxidase and pigment tests.

Streak representative suspect colonies from the agar surface of cetrimide agar on the surfaces of *Pseudomonas* agar medium for detection of fluorescein and *Pseudomonas* agar medium for detection of pyocyanin contained in Petri dishes. Cover and invert the inoculated media and incubate at 33⁰ to 37⁰ for not less than 3 days. Examine the streaked surfaces under ultra-violet light. Examine the plates to determine whether colonies conforming to the description in Table 3 are present.

If growth of suspect colonies occurs, place 2 or 3 drops of a freshly prepared 1per cent w/v solution of *N,N,N',N'*-tetramethyl-4-phenylenediamine dihydrochloride on filter paper and smear with the colony; if there is no development of a pink colour, changing to purple, the sample meets the requirements of the test for the absence of *Pseudomonas aeruginosa*.

Staphylococcus aureus : Proceed as described under *Pseudomonas aeruginosa* if, upon examination of the incubated plates, none of them contains colonies having the characteristics listed in Table 8 for the media used, the sample meets the requirements for the absence of *Staphylococcus aureus*.

If growth occurs, carry out the coagulase test. Transfer representative suspect colonies from the agar surface of any of the media listed in Table 4 to individual tubes, each containing 0.5 ml of mammalian, preferably rabbit or horse, plasma with or without additives. Incubate in water-bath at 37° examining the tubes at 3 hours and subsequently at suitable intervals up to 24 hours. If no coagulation in any degree is observed, the sample meets the requirements of the test for the absence of *Staphylococcus aureus*.

Validity of the tests for total aerobic microbial count:

Grow the following test strains separately in tubes containing fluid soyabean-casein digest medium at 30° to 35° for 18 to 24 hours or, for *Candida albicans*, at 20° for 48 hours.

Table 7 – Tests for *Pseudomonas aeruginosa*

Medium	Characteristic colonial morphology	Fluorescence in UV light	Oxidase test	Gram stain
Cetrimide agar	Generally greenish	Greenish	Positive	Negative rods
Pseudomonas agar medium for detection of fluorescein	Generally colourless to yellowish	Yellowish	Positive	Negative rods
Pseudomonas agar medium for detection of pyocyanin	Generally greenish	Blue	Positive	Negative rods

Table 8 – Tests for *Staphylococcus aureus*

Selective medium	Characteristic colonial morphology	Gram stain
Vogel-Johnson agar	Black surrounded by yellow zones	Positive cocci (in clusters)
Mannitol-salt agar	Yellow colonies with yellow zones	Positive cocci (in clusters)
Baird-Parker agar	Black, shiny, surrounded by clear zones of 2 to 5 mm	Positive cocci (in clusters)

<i>Staphylococcus aureus</i>	(ATCC 6538; NCTC 10788)
<i>Bacillus subtilis</i>	(ATCC 6633; NCIB 8054)
<i>Escherichia coli</i>	(ATCC 8739; NCIB 8545)
<i>Candida albicans</i>	(ATCC 2091; ATCC 10231)

Dilute portions of each of the cultures using buffered sodium chloride-peptone solution pH 7.0 to make test suspensions containing about 100 viable micro-organisms per ml. Use the suspension of each of the micro-organisms separately as a control of the counting methods, in the presence and absence of the preparation being examined, if necessary.

A count for any of the test organisms differing by not more than a factor of 10 from the calculated value for the inoculum should be obtained. To test the sterility of the medium and of the diluent and the aseptic performance of the test, carry out the total aerobic microbial count method using sterile buffered sodium chloride-peptone solution pH 7.0 as the test preparation. There should be no growth of micro-organisms.

Validity of the tests for specified micro-organisms: Grow separately the test strains of *Staphylococcus aureus* and *Pseudomonas aeruginosa* in fluid soyabean-casein digest medium and *Escherichia coli* and *Salmonella typhimurium* at 30⁰ to 35⁰ for 18 to 24 hours. Dilute portions of each of the cultures using buffered sodium chloride-peptone solution pH 7.0 to make test suspensions containing about 10³ viable micro-organisms per ml. Mix equal volume of each suspension and use 0.4 ml (approximately 10² micro-organisms of each strain) as an inoculum in the test for *E. coli*, *Salmonella*, *P. aeruginosa* and *S. aureus*, in the presence and absence of the preparation being examined, if necessary. A positive result for the respective strain of micro-organism should be obtained.

Table 9- Microbial Contamination Limits

S.No.	Parameters	Permissible limits
1.	<i>Staphylococcus aureus</i> /g	Absent
2.	<i>Salmonella</i> sp./g	Absent
3.	<i>Pseudomonas aeruginosa</i> /g	Absent
4.	<i>Escherichia coli</i>	Absent
5.	Total microbial plate count (TPC)	10 ⁵ /g*
6.	Total Yeast & Mould	10 ³ /g

*For topical use, the limit shall be 10⁷/g.

2.5. Pesticide Residue:

Definition: For the purposes of the Pharmacopoeia, a pesticide is any substance or mixture of substances intended for preventing, destroying or controlling any pest, unwanted species of plants or animals causing harm during or otherwise interfering with the production, processing, storage,

transport or marketing of vegetable drugs. The item includes substances intended for use as growth-regulators, defoliant or desiccants and any substance applied to crops either before or after harvest to protect the commodity from deterioration during storage and transport.

Limits: Unless otherwise indicated in the monograph, the drug to be examined at least complies with the limits indicated in Table -1, The limits applying to pesticides that are not listed in the table and whose presence is suspected for any reason comply with the limits set by European Community directives 76/895 and 90/642, including their annexes and successive updates. Limits for pesticides that are not listed in Table.-1 nor in EC directives are calculated using the following expression:

$$\frac{ADI \times M}{MDD \times 100}$$

ADI = Acceptable Daily Intake, as published by FAO-WHO, in milligrams per kilogram of body mass,

M = body mass in kilograms (60 kg),

MDD = daily dose of the drug, in kilograms.

If the drug is intended for the preparation of extracts, tinctures or other pharmaceutical forms whose preparation method modifies the content of pesticides in the finished product, the limits are calculated using the following expression:

$$\frac{ADI \times M \times E}{MDD \times 100}$$

E = Extraction factor of the method of preparation, determined experimentally.

Higher limits can also be authorised, in exceptional cases, especially when a plant requires a particular cultivation method or has a metabolism or a structure that gives rise to a higher than normal content of pesticides.

The competent authority may grant total or partial exemption from the test when the complete history (nature and quantity of the pesticides used, date of each treatment during cultivation and after the harvest) of the treatment of the batch is known and can be checked precisely.

Sampling

Method: For containers up to 1 kg, take one sample from the total content, thoroughly mixed, sufficient for the tests. For containers between 1 kg and 5 kg, take three samples, equal in volume, from the upper, middle and lower parts of the container, each being sufficient to carry out the tests. Thoroughly mix the samples and take from the mixture an amount sufficient to carry out the tests. For containers of more than 5 kg, take three samples, each of at least 250 g from the upper, middle and

lower parts of the container. Thoroughly mix the samples and take from the mixture an amount sufficient to carry out the tests.

Size of sampling: If the number (n) of containers is three or fewer, take samples from each container as indicated above under Method. If the number of containers is more than three, take n+1 samples for containers as indicated under Method, rounding up to the nearest unit if necessary.

The samples are to be analysed immediately to avoid possible degradation of the residues. If this is not possible, the samples are stored in air-tight containers suitable for food contact, at a temperature below 0⁰, protected from light.

Reagents: All reagents and solvents are free from any contaminants, especially pesticides, that might interfere with the analysis. It is often necessary to use special quality solvents or, if this is not possible, solvents that have recently been re-distilled in an apparatus made entirely of glass. In any case, suitable blank tests must be carried out.

Apparatus: Clean the apparatus and especially glassware to ensure that they are free from pesticides, for example, soak for at least 16 h in a solution of phosphate-free detergent, rinse with large quantities of *distilled water* and wash with *acetone* and *hexane* or *heptane*.

2.5.1. Qualitative and Quantitative Analysis of Pesticide Residues:

The analytical procedures used are validated according to the regulations in force. In particular, they satisfy the following criteria :

- the chosen method, especially the purification steps, are suitable for the combination pesticide residue/substance to be analysed and not susceptible to interference from co-extractives; the limits of detection and quantification are measured for each pesticide-matrix combination to be analysed.
- between 70 per cent to 110 per cent of each pesticide is recovered.
- the repeatability of the method is not less than the values indicated in Table 10
- the reproducibility of the method is not less than the values indicated in Table 11
- the concentration of test and reference solutions and the setting of the apparatus are such that a linear response is obtained from the analytical detector.

Table -10

Substance	Limit (mg/kg)
Alachlor	0.02
Aldrin and Dieldrin (sum of)	0.05
Azinphos-methyl	1.0
Bromopropylate	3.0

Chlordane (sum of cis-, trans – and Oxythlordane)	0.05
Chlorfenvinphos	0.5
Chlorpyrifos	0.2
Chlorpyrifos-methyl	0.1
Cypermethrin (and isomers)	1.0
DDT (sum of p,p-‘DDT, o,p-‘DDT, p,p-‘DDE and p,p-‘TDE	1.0
Deltamethrin	0.5
Diazinon	0.5
Dichlorvos	1.0
Dithiocarbamates (as CS ₂)	2.0
Endosulfan (sum of isomers and Endosulfan sulphate)	3.0
Endrin	0.05
Ethion	2.0
Fenitrothion	0.5
Fenvalerate	1.5
Fonofos	0.05
Heptachlor (sum of Heptachlor and Heptachlorepoxyde)	0.05
Hexachlorobenzene	0.1
Hexachlorocyclohexane isomers (other than γ)	0.3
Lindane (γ -Hexachlorocyclohexane)	0.6
Malathion	1.0
Methidathion	0.2
Parathion	0.5
Parathion-methyl	0.2
Permethrin	1.0
Phosalone	0.1
Piperonyl butoxide	3.0
Pirimiphos-methyl	4.0
Pyrethrins (sum of)	3.0

Quintozene (sum of quintozene, pentachloroaniline and methyl pentachlorophenyl sulphide)	1.0
--	-----

Table -11

Concentration of the pesticide (mg/kg)	Repeatability (difference, ± mg/kg)	Reproducibility (difference, ± mg/kg)
0.010	0.005	0.01
0.100	0.025	0.05
1.000	0.125	0.25

2.5.2. Test for Pesticides:

Organochlorine, Organophosphorus and Pyrethroid Insecticides:

The following methods may be used, in connection with the general method above, depending on the substance being examined, it may be necessary to modify, sometimes extensively, the procedure described hereafter. In any case, it may be necessary to use, in addition, another column with a different polarity or another detection method (mass spectrometry) or a different method (immunochemical methods) to confirm the results obtained.

This procedure is valid only for the analysis of samples of vegetable drugs containing less than 15 per cent of water. Samples with a higher content of water may be dried, provided it has been shown that the drying procedure does not affect significantly the pesticide content.

Extraction

To 10 g of the substance being examined, coarsely powdered, add 100 ml of *acetone* and allow to stand for 20 min. Add 1 ml of a solution containing 1.8 µg/ml of *carbophenothion* in *toluene*. Homogenise using a high-speed blender for 3 min. Filter and wash the filter cake with two quantities, each of 25 ml, of *acetone*. Combine the filtrate and the washings and heat using a rotary evaporator at a temperature not exceeding 40⁰ C until the solvent has almost completely evaporated. To the residue add a few milliliters of *toluene* and heat again until the acetone is completely removed. Dissolve the residue in 8 ml of *toluene*. Filter through a membrane filter (45 µm), rinse the flask and the filter with *toluene* and dilute to 10.0 ml with the same solvent (solution A).

Purification

Organochlorine, organophosphorus and pyrethroid insecticides:

Examine by size-exclusion chromatography.

The chromatographic procedure may be carried out using:

- a stainless steel column 0.30 m long and 7.8 mm in internal diameter packed with styrene-divinylbenzene copolymer (5 μm).
- as mobile phase *toluene* at a flow rate of 1 ml/min.

Performance of the column: Inject 100 μl of a solution containing 0.5 g/l of *methyl red* and 0.5 g/l of *oracet blue* in *toluene* and proceed with the chromatography. The column is not suitable unless the colour of the eluate changes from orange to blue at an elution volume of about 10.3 ml. If necessary calibrate the column, using a solution containing, in *toluene*, at a suitable concentration, the insecticide to be analysed with the lowest molecular mass (for example, dichlorvos) and that with the highest molecular mass (for example, deltamethrin). Determine which fraction of the eluate contains both insecticides.

Purification of the test solution: Inject a suitable volume of solution A (100 μl to 500 μl) and proceed with the chromatography. Collect the fraction as determined above (solution B). Organophosphorus insecticides are usually eluted between 8.8 ml and 10.9 ml. Organochlorine and pyrethroid insecticides are usually eluted between 8.5 ml and 10.3 ml.

Organochlorine and pyrethroid insecticides: In a chromatography column, 0.10 m long and 5 mm in internal diameter, introduce a piece of defatted cotton and 0.5 g of silica gel treated as follows: heat silica gel for chromatography in an oven at 150⁰ for at least 4 h. Allow to cool and add dropwise a quantity of *water* corresponding to 1.5 per cent of the mass of silica gel used; shake vigorously until agglomerates have disappeared and continue shaking for 2 h using a mechanical shaker. Condition the column using 1.5 ml of *hexane*. Prepacked columns containing about 0.50 g of a suitable silica gel may also be used provided they are previously validated.

Concentrate solution B in a current of helium for chromatography or oxygen-free nitrogen almost to dryness and dilute to a suitable volume with *toluene* (200 μl to 1 ml according to the volume injected in the preparation of solution B). Transfer quantitatively onto the column and proceed with the chromatography using 1.8 ml of *toluene* as the mobile phase. Collect the eluate (solution C).

2.5.3. Quantitative Analysis:

A. Organophosphorus insecticides: Examine by gas chromatography, using *carbophenothion* as internal standard. It may be necessary to use a second internal standard to identify possible interference with the peak corresponding to carbophenothion.

Test solution: Concentrate solution B in a current of helium for chromatography almost to dryness and dilute to 100 μl with *toluene*.

Reference solution: Prepare at least three solutions in *toluene* containing the insecticides to be determined and *carbophenothion* at concentrations suitable for plotting a calibration curve.

The chromatographic procedure may be carried out using:

- a fused-silica column 30 m long and 0.32 mm in internal diameter the internal wall of which is covered with a layer 0.25 μm thick of poly (dimethyl) siloxane.
- hydrogen for chromatography as the carrier gas. Other gases such as helium for chromatography or nitrogen for chromatography may also be used provided the chromatography is suitably validated.
- a phosphorus-nitrogen flame-ionisation detector or a atomic emission spectrometry detector.

Maintaining the temperature of the column at 80° for 1 min, then raising it at a rate of $30^{\circ}/\text{min}$ to 150° , maintaining at 150° for 3 min, then raising the temperature at a rate of $4^{\circ}/\text{min}$ to 280° and maintaining at this temperature for 1 min and maintaining the temperature of the injector port at 250° and that of the detector at 275° . Inject the chosen volume of each solution. When the chromatograms are recorded in the prescribed conditions, the relative retention times are approximately those listed in Table 12 Calculate the content of each insecticide from the peak areas and the concentrations of the solutions.

B. Organochlorine and Pyrethroid Insecticides:

Examine by gas chromatography, using *carbophenothion* as the internal standard. It may be necessary to use a second internal standard to identify possible interference with the peak corresponding to *carbophenothion*.

Test solution: Concentrate solution C in a current of helium for chromatography or oxygen-free nitrogen almost to dryness and dilute to 500 μl with *toluene*.

Reference solution: Prepare at least three solutions in *toluene* containing the insecticides to be determined and *carbophenothion* at concentrations suitable for plotting a calibration curve.

Table 12- Relative Retention Times of Pesticides

Substance	Relative retention times
Dichlorvos	0.20
Fonofos	0.50
Diazinon	0.52
Parathion-methyl	0.59
Chlorpyrifos-methyl	0.60
Pirimiphos-methyl	0.66
Malathion	0.67
Parathion	0.69
Chlorpyrifos	0.70
Methidathion	0.78

Ethion	0.96
Carbophenothion	1.00
Azinphos-methyl	1.17
Phosalon	1.18

The chromatographic procedure may be carried out using:

- a fused silica column 30 m long and 0.32 mm in internal diameter the internal wall of which is covered with a layer 0.25 μm thick of *poly (dimethyl diphenyl) siloxane*.
- hydrogen for chromatography as the carrier gas. Other gases such as helium for chromatography or nitrogen for chromatography may also be used, provided the chromatography is suitably validated.
- an electron-capture detector.
- a device allowing direct cold on-column injection.

maintaining the temperature of the column at 80⁰ for 1 min, then raising it at a rate of 30⁰/min to 150⁰, maintaining at 150⁰ for 3 min, then raising the temperature at a rate of 4⁰/min to 280⁰ and maintaining at this temperature for 1 min and maintaining the temperature of the injector port at 250⁰ and that of the detector at 275⁰. Inject the chosen volume of each solution. When the chromatograms are recorded in the prescribed conditions, the relative retention times are approximately those listed in Table 13. Calculate the content of each insecticide from the peak areas and the concentrations of the solutions.

Table 13- Relative Retention Times of Insecticides

Substance	Relative retention times
α -Hexachlorocyclohexane	0.44
Hexachlorobenzene	0.45
β -Hexachlorocyclohexane	0.49
Lindane	0.49
δ -Hexachlorocyclohexane	0.54
ϵ -Hexachlorocyclohexane	0.56
Heptachlor	0.61
Aldrin	0.68
<i>cis</i> -Heptachlor-epoxide	0.76
<i>o,p'</i> - DDE	0.81

α -Endosulfan	0.82
Dieldrin	0.87
<i>p,p'</i> -DDE	0.87
<i>o,p'</i> -DDD	0.89
Endrin	0.91
β -Endosulfan	0.92
<i>o,p'</i> -DDT	0.95
Carbophenothion	1.00
<i>p,p'</i> -DDT	1.02
<i>cis</i> -Permethrin	1.29
<i>trans</i> -Permethrin	1.31
Cypermethrin*	1.40
Fenvalerate*	1.47 and 1.49
Deltamethrin	1.54

*The substance shows several peaks.

2.6. Test for Aflatoxins:

Caution: Aflatoxins are highly dangerous and extreme care should be exercised in handling aflatoxin materials.

This test is provided to detect the possible presence of aflatoxins B₁, B₂, G₁ and G₂ in any material of plant origin. Unless otherwise specified in the individual monograph, use the following method.

Zinc Acetate – Aluminum Chloride Reagent: Dissolve 20 g of *zinc acetate* and 5 g of *aluminum chloride* in sufficient water to make 100 ml.

Sodium Chloride Solution: Dissolve 5 g of *sodium chloride* in 50 ml of purified water.

Test Solution 1: Grind about 200 g of plant material to a fine powder. Transfer about 50 g of the powdered material, accurately weighed, to a glass-stoppered flask. Add 200 ml of a mixture of *methanol* and *water* (17: 3). Shake vigorously by mechanical means for not less than 30 minutes and filter. [Note – If the solution has interfering plant pigments, proceed as directed for Test Solution 2.] Discard the first 50 ml of the filtrate and collect the next 40 ml portion. Transfer the filtrate to a separatory funnel. Add 40 ml of sodium chloride solution and 25 ml of *hexane* and shake for 1 minute. Allow the layers to separate and transfer the lower aqueous layer to a second separatory funnel. Extract the aqueous layer in the separatory funnel twice, each time with 25 ml of *methylene*

chloride, by shaking for 1 minute. Allow the layers to separate each time, separate the lower organic layer and collect the combined organic layers in a 125 ml conical flask. Evaporate the organic solvent to dryness on a water bath. Cool the residue. If interferences exist in the residue, proceed as directed for *Cleanup Procedure*; otherwise, dissolve the residue obtained above in 0.2 ml of a mixture of *chloroform* and *acetonitrile* (9.8: 0.2) and shake by mechanical means if necessary.

Test Solution 2: Collect 100 ml of the filtrate from the start of the flow and transfer to a 250 ml beaker. Add 20 ml of *Zinc Acetate-Aluminum Chloride Reagent* and 80 ml of water. Stir and allow to stand for 5 minutes. Add 5 g of a suitable filtering aid, such as diatomaceous earth, mix and filter. Discard the first 50 ml of the filtrate, and collect the next 80 ml portion. Proceed as directed for *Test Solution 1*, beginning with “Transfer the filtrate to a separatory funnel.”

Cleanup Procedure: Place a medium-porosity sintered-glass disk or a glass wool plug at the bottom of a 10 mm x 300 mm chromatographic tube. Prepare slurry of 2 g of silica gel with a mixture of *ethyl ether* and *hexane* (3: 1), pour the slurry into the column and wash with 5 ml of the same solvent mixture. Allow the absorbent to settle and add to the top of the column a layer of 1.5 g of *anhydrous sodium sulfate*. Dissolve the residue obtained above in 3 ml of *methylene chloride* and transfer it to the column. Rinse the flask twice with 1 ml portions of *methylene chloride*, transfer the rinses to the column and elute at a rate not greater than 1 ml per minute. Add successively to the column 3 ml of *hexane*, 3 ml of *diethyl ether* and 3 ml of *methylene chloride*; elute at a rate not greater than 3 ml per minute; and discard the eluates. Add to the column 6 mL of a mixture of *methylene chloride* and *acetone* (9: 1) and elute at a rate not greater than 1 ml per minute, preferably without the aid of vacuum. Collect this eluate in a small vial, add a boiling chip if necessary and evaporate to dryness on a water bath. Dissolve the residue in 0.2 ml of a mixture of *chloroform* and *acetonitrile* (9.8 : 0.2) and shake by mechanical means if necessary.

Aflatoxin Solution: Dissolve accurately weighed quantities of aflatoxin B₁, aflatoxin B₂, aflatoxin G₁ and aflatoxin G₂ in a mixture of *chloroform* and *acetonitrile* (9.8: 0.2) to obtain a solution having concentrations of 0.5 µg /per ml each for aflatoxin B₁ and G₁ and 0.1µg per ml each for aflatoxins for B₂ and G₂.

Procedure: Separately apply 2.5 µl, 5 µl, 7.5 µl and 10 µl of the Aflatoxin Solution and three 10 µl applications of either *Test Solution 1* or *Test Solution 2* to a suitable thin-layer chromatographic plate coated with a 0.25-mm layer of chromatographic silica gel mixture. Superimpose 5 µl of the *Aflatoxin Solution* on one of the three 10 µl applications of the *Test Solution*. Allow the spots to dry and develop the chromatogram in an unsaturated chamber containing a solvent system consisting of a mixture of *chloroform*, *acetone* and *isopropyl alcohol* (85:10:5) until the solvent front has moved not less than 15 cm from the origin. Remove the plate from the developing chamber, mark the solvent front and allow the plate to air-dry. Locate the spots on the plate by examination under UV light at 365 nm: the four applications of the *Aflatoxin Solution* appear as four clearly separated blue fluorescent spots; the spot obtained from the *Test Solution* that was superimposed on the *Aflatoxin Solution* is no more intense than that of the corresponding *Aflatoxin Solution*; and no spot from any

of the other *Test Solutions* corresponds to any of the spots obtained from the applications of the *Aflatoxin Solution*. If any spot of aflatoxins is obtained in the *Test Solution*, match the position of each fluorescent spot of the *Test Solution* with those of the *Aflatoxin Solution* to identify the type of aflatoxin present. The intensity of the aflatoxin spot, if present in the *Test Solution*, when compared with that of the corresponding aflatoxin in the *Aflatoxin Solution* will give an approximate concentration of aflatoxin in the *Test Solution*.

Table14 - Permissible Limit of Aflatoxins*

S.No	Aflatoxins	Permissible Limit
1.	B ₁	0.5 ppm
2.	G ₁	0.5 ppm
3.	B ₂	0.1 ppm
4.	G ₂	0.1 ppm

*For Domestic use only

2.7. Gas Chromatography:

Gas chromatography (GC) is a chromatographic separation technique based on the difference in the distribution of species between two non-miscible phases in which the mobile phase is a carrier gas moving through or passing the stationary phase contained in a column. It is applicable to substances or their derivatives, which are volatilized under the temperatures employed.

GC is based on mechanisms of adsorption, mass distribution or size exclusion.

Apparatus

The apparatus consists of an injector, a chromatographic column contained in an oven, a detector and a data acquisition system (or an integrator or a chart recorder). The carrier gas flows through the column at a controlled rate or pressure and then through the detector.

The chromatography is carried out either at a constant temperature or according to a given temperature programme.

Injectors

Direct injections of solutions are the usual mode of injection, unless otherwise prescribed in the monograph. Injection may be carried out either directly at the head of the column using a syringe or an injection valve, or into a vaporization chamber which may be equipped with a stream splitter.

Injections of vapour phase may be effected by static or dynamic head-space injection systems.

Dynamic head-space (purge and trap) injection systems include a sparging device by which volatile substances in solution are swept into an absorbent column maintained at a low temperature. Retained substances are then desorbed into the mobile phase by rapid heating of the absorbent column.

Static head-space injection systems include a thermostatically controlled sample heating chamber in which closed vials containing solid or liquid samples are placed for a fixed period of time to allow the volatile components of the sample to reach equilibrium between the non-gaseous phase and the vapour phase. After equilibrium has been established, a predetermined amount of the head-space of the vial is flushed into the gas chromatograph.

Stationary Phases

Stationary phases are contained in columns, which may be:

- a capillary column of fused-silica close wall is coated with the stationary phase.
- a column packed with inert particles impregnated with the stationary phase.
- a column packed with solid stationary phase.

Capillary columns are 0.1 mm to 0.53 mm in internal diameter (Φ) and 5 to 6 m in length. The liquid or stationary phase, which may be chemically bonded to the inner surface, is a film 0.1 μm to 5.0 μm thick.

Packed columns, made of glass or metal, are usually 1 m to 3 m in length with an internal diameter (Φ) of 2 mm to 4 mm. Stationary phases usually consist of porous polymers or solid supports impregnated with liquid phase.

Supports for analysis of polar compounds on columns packed with low-capacity, low-polarity stationary phase must be inert to avoid peak tailing. The reactivity of support materials can be reduced by silanising prior to coating with liquid phase. Acid-washed, flux-calcinated diatomaceous earth is often used. Materials are available in various particle sizes, the most commonly used particles are in the ranges of 150 μm to 180 μm and 125 μm to 150 μm .

Mobile Phases

Retention time and peak efficiency depend on the carrier gas flow rate; retention time is directly proportional to column length and resolution is proportional to the square root of the column length. For packed columns, the carrier gas flow rate is usually expressed in milliliters per minute at atmospheric pressure and room temperature, flow rate is measured at the detector outlet, either with a calibrated mechanical device or with a bubble tube, while the column is at operating temperature. The linear velocity of the carrier gas through a packed column is inversely proportional to the square root of the internal diameter of the column for a given flow volume. Flow rates of 60 ml/min in a 4 mm internal diameter column and 15 ml/min in a 2 mm internal diameter column, give identical linear velocities and thus similar retention times.

Helium or nitrogen is usually employed as the carrier gas for packed columns, whereas commonly used carrier gases for capillary columns are nitrogen, helium and hydrogen.

Detectors

Flame-ionisation detectors are usually employed but additional detectors which may be used include: electron-capture, nitrogen-phosphorus, mass spectrometric, thermal conductivity, Fourier transform infrared spectrophotometric and others, depending on the purpose of the analysis.

Method

Equilibrate the column, the injector and the detector at the temperatures and the gas flow rates specified in the monograph until a stable baseline is achieved. Prepare the test solution (s) and the reference solutions (s) as prescribed. The solutions must be free from solid particles.

Criteria for assessing the suitability of the system are described in the chapter on *Chromatographic separation techniques*. The extent to which adjustments of parameters of the chromatographic system can be made to satisfy the criteria of system suitability are also given in this chapter.

2.8. Test for the Absence of Methanol:

Take 1 drop of the sample in a 15 ml test tube. Add 1 drop of water with 1-drop dilute phosphoric acid (10 per cent of water) followed by 1 drop of potassium permanganate solution (1 per cent w/v of water). Add sodium bisulphate solution dropwise until the permanganate color is discharged. If brown color remains add 1 drop of dilute phosphoric acid followed by 5 ml of chromotropic acid solution (5 mg chromotropic acid sodium salt in 10 ml mixture of 9 ml *sulphuric acid* & 4 ml water) and heat to 60⁰ C for 10 minutes. If no violet color is produced indicates the absence of *methanol*.

APPENDIX - 3

PHYSICAL TESTS AND DETERMINATIONS

3.1. Refractive Index:

The refractive index (η) of a substance with reference to air, is the ratio of the sine of the angle of incidence to the sine of the angle of refraction of a beam of light passing from air into the substance. It varies with the wavelength of the light used in its measurement.

Unless otherwise prescribed, the refractive index is measured at 25° (± 0.5) with reference to the wavelength of the D line of sodium (λ 589.3 nm). The temperature should be carefully adjusted and maintained since the refractive index varies significantly with temperature.

The Abbe's refractometer is convenient for most measurements of refractive index but other refractometer of equal or greater accuracy may be used. Commercial refractometers are normally constructed for use with white light but are calibrated to give the refractive index in terms of the D line of sodium light.

To achieve accuracy, the apparatus should be calibrated against *distilled water* which has a refractive index of 1.3325 at 25° or against the reference liquids given in the following table.

Table 15

Reference Liquid	$\eta_D 20^{\circ}$	Temperature Co-efficient $\Delta n / \Delta t$
Carbon tetrachloride	1.4603	-0.00057
Toluene	1.4969	-0.00056
α -Methylnaphthalene	1.6176	-0.00048

* Reference index value for the D line of sodium, measured at 20° .

The cleanliness of the instrument should be checked frequently by determining the refractive index of distilled water, which at 25° is 1.3325.

3.2. Weight per Millilitre and Specific Gravity:

A. Weight per millilitre: The weight per millilitre of a liquid is the weight in g of 1 ml of a liquid when weighed in air at 25° , unless otherwise specified.

Method

Select a thoroughly clean and dry pycnometer. Calibrate the pycnometer by filling it with recently boiled and cooled *water* at 25° and weighing the contents. Assuming that the weight of 1 ml of *water* at 25° when weighed in air of density 0.0012 g per ml, is 0.99602 g. Calculate the capacity of the pycnometer. (Ordinary deviations in the density of air from the value given do not affect the result of

a determination significantly). Adjust the temperature of the substance to be examined, to about 20° and fill the pycnometer with it. Adjust the temperature of the filled pycnometer to 25°, remove any excess of the substance and weigh. Subtract the tare weight of the pycnometer from the filled weight of the pycnometer. Determine the weight per milliliter dividing the weight in air, expressed in g, of the quantity of liquid which fills the pycnometer at the specified temperature, by the capacity expressed in ml, of the pycnometer at the same temperature.

B. Specific gravity: The specific gravity of a liquid is the weight of a given volume of the liquid at 25° (unless otherwise specified) compared with the weight of an equal volume of water at the same temperature, all weighing being taken in air.

Method

Proceed as described under wt. per ml. Obtain the specific gravity of the liquid by dividing the weight of the liquid contained in the pycnometer by the weight of water contained, both determined at 25° unless otherwise directed in the individual monograph.

3.3. Determination of pH Values:

The *pH* value of an aqueous liquid may be defined as the common logarithm of the reciprocal of the hydrogen ion concentration expressed in g per litre. Although this definition provides a useful practical means for the quantitative indication of the acidity or alkalinity of a solution, it is less satisfactory from a strictly theoretical point of view. No definition of *pH* as a measurable quantity can have a simple meaning, which is also fundamental and exact.

The *pH* value of a liquid can be determined potentiometrically by means of the glass electrode, a reference electrode and a pH meter either of the digital or analogue type.

3.4. Determination of Melting Range and Congealing Range:

3.4.1. Determination of Melting Range:

The melting-range of a substance is the range between the corrected temperature at which the substance begins to form droplets and the corrected temperature at which it completely melts, as shown by formation of a meniscus.

Apparatus:

- (a) A capillary tube of soft glass, closed at one end, and having the following dimensions:
 - (i) thickness of the wall, about 0.10 to 0.15 mm.
 - (ii) length about 10 cm or any length suitable for apparatus used.
 - (iii) internal diameter 0.9 to 1.1 mm for substances melting below 100° or 0.8 to 1.2 mm for substances melting above 100°.

Thermometers:

Accurately standardized thermometers covering the range 10° to 300° , the length of two degrees on the scale being not less than 0.8 mm. These thermometers are of the mercury-in-glass, solid-stem type; the bulb is cylindrical in shape, and made of approved thermometric glass suitable for the range of temperature covered; each thermometer is fitted with a safety chamber. The smallest division on the thermometer scale should vary between 0.1° to 1.5° , according to the melting point of the substance under test.

The following form of heating apparatus is recommended.

A glass heating vessel of suitable, construction and capacity fitted with suitable stirring device, capable of rapidly mixing the liquids.

Suitable liquids for use in the heating vessel:

<i>Glycerin</i>	Upto 150°
<i>Sulphuric acid</i> to which a small crystal of <i>potassium nitrate</i> or 4 Drops of <i>nitric acid</i> per 100 ml has been added	Upto 200°
A liquid paraffin of sufficiently high boiling range	Upto 250°
Seasame oil	Upto 300°
30 parts of <i>potassium sulphate</i> , dissolved by heating in 70 parts of <i>sulphuric acid</i>	Upto 300°

Any other apparatus or method, preferably, the electric method may be used subject to a check by means of pure substances having melting temperature covering the ranges from 0° to 300° and with suitable intervals.

The following substances are suitable for this purpose.

Substance	Melting range
Vanillin	81° to 83°
Acetanilide	114° to 116°
Phenacetin	134° to 136°
Sulphanilamide	164° to 166.5°
Sulphapyridine	191° to 193°
Caffeine (Dried at 100°)	234° to 237°

Procedure

Method I: Transfer a suitable quantity of the powdered and thoroughly dried substance to a dry capillary tube and pack the powder by tapping the tube on a hard surface so as to form a tightly packed column of 2 to 4 mm in height. Attach the capillary tube and its contents to a standardized thermometer so that the closed end is at the level of the middle of the bulb; heat in a suitable apparatus (preferably a round-bottom flask) fitted with an auxiliary thermometer regulating the rise of temperature in the beginning to 3^o per minute. When the temperature reached is below the lowest figure of the range for the substance under examination, the heating of the apparatus is adjusted as desired; if no other directions are given, the rate of rise of temperature should be kept at 1^o to 2^o per minute. The statement ‘determined by rapid heating’ means that the rate of rise of temperature is 5^o per minute during the entire period of heating.

Unless otherwise directed, the temperature at which the substance forms droplets against the side of the tube and the one at which it is completely melted as indicated by the formation of a definite meniscus, are read.

The following emergent stem corrections should be applied to the temperature readings.

Before starting the determination of the melting temperature the auxiliary thermometer is attached so that the bulb touches the standard thermometer at a point midway between the graduation for the expected melting temperature and the surface of the heating material. When the substance has melted, the temperature is read on the auxiliary thermometer. The correction figure to be added to the temperature reading of the standardized thermometer is calculated from the following formula

$$0.00015 N (T-t)$$

Where ‘T’ is the temperature reading of the standardized thermometer

‘t’ is the temperature reading of the auxiliary thermometer

‘N’ is the number of degrees of the scale of the standardized thermometer between the surface of the heating material and level of mercury

The statement “melting range, a^o to b^o” means that the corrected temperature at which the material forms droplets must be at least a^o, and that the material must be completely melted at the corrected temperature, b^o.

Method II: The apparatus employed for this test is the same as described for method I except for such details as are mentioned in the procedure given below

Procedure: A capillary tube open at both ends is used for this test. Melt the material under test at as low a temperature as possible. Draw into the capillary a column of the material about 10 mm high. Cool the charged tube in contact with ice for at least 2 hours. Attach the tube to the thermometer by means of rubber band and adjust it in the heating vessel containing water so that the upper edge of the material is 10 mm below the water level. Heat in the manner as prescribed in Method I, until the temperature is about 5^o below the expected melting point and then regulate the rate of rise of

temperature between 0.5° to 1° per minute. The temperature at which the material is observed to rise in the capillary tube is the melting temperature of the substance.

3.4.2. Determination of Congealing Range:

The congealing temperature is that point at which there exists a mixture of the liquid (fused) phase of a substance and a small but increasing proportion of the solid phase. It is distinct from the freezing point which is the temperature at which the liquid and solid phases of a substance are in equilibrium. In certain cases, this may happen over a range of temperatures.

The temperature at which a substance solidifies upon cooling is a useful index of its purity if heat is liberated when solidification takes place.

The following method is applicable to substances that melt between -20° and 150° .

Apparatus

A test-tube (About 150 mm \times 25 mm) placed inside another test-tube (about 160 mm \times 40 mm) the inner tube is closed by a stopper that carries a stirrer and a thermometer (About 175 mm long and with 0.2° graduations) fixed so that the bulb is about 15 mm above the bottom of the tube. The stirrer is made from a glass rod or other suitable material formed at one end into a loop of about 18 mm overall diameter at right angles to the rod. The inner tube with its jacket is supported centrally in a 1-litre baker containing a suitable cooling liquid to within 20 mm of the top. The thermometer is supported in the cooling bath.

Method

Melt the substance, if a solid, at a temperature not more than 20° above its expected congealing point, and pour it into the inner test-tube to a height of 50 to 57 mm. Assemble the apparatus with the bulb of the thermometer immersed half-way between the top and bottom of the sample in the test-tube. Fill the bath to almost 20 mm from the top of the tube with a suitable fluid at a temperature 4° to 5° below the expected congealing point. If the substance is a liquid at room temperature, carry out the determination using a bath temperature about 15° below the expected congealing point. When the sample has cooled to about 5° above its expected congealing point stir it continuously by moving the loop up and down between the top and bottom of the sample at a regular rate of 20 complete cycles per minute. If necessary, congelation may be induced by scratching the inner walls of the test-tube with the thermometer or by introducing a small amount of the previously congealed substance under examination. Pronounced supercooling may result in deviation from the normal pattern of temperature changes. If it happens, repeat the test introducing small fragments of the solid substance under examination at 1° intervals when the temperature approaches the expected congealing point.

Record the reading of the thermometer every 30 seconds and continue stirring only so long as the temperature is falling. Stop the stirring when the temperature is constant to starts to rise slightly. Continue recording the temperature for at least 3 minutes after the temperature again begins to fall after remaining constant.

The congealing point will be mean of not less than four consecutive readings that lie within a range of 0.2° .

3.5. Determination of Boiling Range:

The boiling-range of a substance is the range of temperature within which the whole or a specified portion of the substance distils.

Apparatus

The boiling-range is determined in a suitable apparatus, the salient features of which are described below:

(a) **Distillation flask:** The flask shall be made of colourless transparent heat-resistant glass and well annealed. It should have a spherical bulb having a capacity of about 130 ml. The side tube slopes downwards in the same plane as the axis of the neck at angle of between 72° to 78° . Other important dimensional details are as under:

Internal diameter of neck	15 to 17 mm
Distance from top of neck to center of side tube	72 to 78 mm
Distance from the center of the side tube to surface of the liquid when the flask contains 100 ml liquid	87 to 93 mm
Internal diameter of side tube	3.5 to 4.5 mm
Length of side tube	97 to 103 mm

(b) **Thermometer:** Standardised thermometers calibrated for 100 mm immersion and suitable for the purpose and covering the boiling range of the substance under examination shall be employed; the smallest division on the thermometer scale may vary between 0.2° to 1° according to requirement.

(c) **Draught Screen:** suitable draught screen, rectangular in cross section with a hard asbestos board about 6 mm thick closely fitting horizontally to the sides of the screen, should be used. The asbestos board shall have a centrally cut circular hole, 110 mm in diameter. The asbestos board is meant for ensuring that hot gases from the heat source do not come in contact with the sides or neck of the flask.

(d) **Asbestos Board:** A 150 mm square asbestos board 6 mm thick provided with a circular hole located centrally to hold the bottom of the flask, shall be used. For distillation of liquids boiling below 60° the hole shall be 30 mm in diameter; for other liquid it should be 50 mm in diameter. This board is to be placed on the hard asbestos board of the draught screen covering its 110 mm hole.

(e) **Condenser:** A straight water-cooled glass condenser about 50 cm long shall be used.

Procedure: 100 ml of the liquid to be examined is placed in the distillation flask, and a few glass beads or other suitable substance is added. The bulb of the flask is placed centrally over a circular hole varying from 3 to 5 cm in diameter (according to the boiling range of the substance under

examination), in a suitable asbestos board. The thermometer is held concentrically in the neck of the flask by means of a well fitting cork in such a manner that the bulb of the thermometer remains just below the level of the opening of the side-tube. Heat the flask slowly in the beginning and when distillation starts, adjust heating in such a manner that the liquid distils at a constant rate of 4 to 5 ml per minute. The temperature is read when the first drop runs from the condenser, and again when the last quantity of liquid in the flask is evaporated.

The boiling ranges indicated, apply at a barometric pressure of 760 mm of mercury. If the determination is made at some other barometric pressure, the following correction is added to the temperatures read:

$$K - (760 - p)$$

Where p is the barometric pressure (in mm) read on a mercury barometer, without taking into account the temperature of the air;

K is the boiling temperature constant for different liquids having different boiling ranges as indicated below:-

Observed Boiling range	'K'
Below 100 ⁰	0.04
100 ⁰ to 140 ⁰	0.045
141 ⁰ to 190 ⁰	0.05
191 ⁰ to 240 ⁰	0.055
above 240 ⁰	0.06

If the barometric pressure is below 760 mm of mercury the correction is added to the observed boiling-range; if above, the correction is subtracted.

The statement 'distils between a⁰ and b⁰', means that temperature at which the first drop runs from the condenser is not less than a⁰ and that the temperature at which the liquid is completely evaporated is not greater than b⁰.

Micro-methods of equal accuracy may be used.

3.6. Determination of Optical Rotation and Specific Optical Rotation:

A. Optical Rotation :Certain substances, in a pure state, in solution and in tinctures possess the property of rotating the plane of polarized light, i.e., the incident light emerges in a plane forming an angle with the plane of the incident light. These substances are said to be optically active and the property of rotating the plane of polarized light is known as optical rotation. The optical rotation is defined as the angle through which the plane of polarized light is rotated when polarized light obtained from sodium or mercury vapour lamp passes through one decimeter thick layer of a liquid or a solution of a substance at a temperature of 25⁰ unless as otherwise stated in the monograph.

Substances are described as dextrorotatory or laevoretatory according to the clockwise or anticlockwise rotation respectively of the plane of polarized light. Dextrorotation is designated by a plus (+) sign and laevorotation by a minus (-) sign before the number indicating the degrees of rotation.

Apparatus: A polarimeter on which angular rotation accurate 0.05° can be read may be used.

Calibration: The apparatus may be checked by using a solution of previously dried *sucrose* and measuring the optical rotation in a 2-dm tube at 25° and using the concentrations indicated in Table.

Concentration (g/100 ml)	Angle of Rotation (+) at 25°
10.0	13.33
20.0	26.61
30.0	39.86
40.0	53.06
50.0	66.23

Procedure: For liquid substances, take a minimum of five readings of the rotation of the liquid and also for an empty tube at the specified temperature. For a solid dissolve in a suitable solvent and take five readings of the rotation of the solution and the solvent used. Calculate the average of each set of five readings and find out the corrected optical rotation from the observed rotation and the reading with the blank (average).

B. Specific Rotation : The apparatus and the procedure for this determination are the same as those specified for optical rotation.

Specific rotation is denoted by the expression

$$[\alpha]_{\lambda}^t$$

t denotes the temperature of rotation; λ denotes the wave length of light used or the characteristic spectral line. Specific rotations are expressed in terms of sodium light of wave length 589.3 nm (D line) and at a temperature of 25°, unless otherwise specified.

Specific rotation of a substance may be calculated from the following formulae:

For liquid substances

$$[\alpha]_{\lambda}^t = \frac{\alpha}{ld}$$

For solutions of substances

$$[\alpha]^t \leftrightarrow = \frac{a \times 100}{lc}$$

Where a is the corrected observed rotation in degrees

l is the length of the polarimeter tube in decimeters.

D is the specific gravity of the liquid C is the concentration of solution expressed as the number of g of the substance in 100 ml of solution.

3.7. Determination of Viscosity:

Viscosity is a property of a liquid, which is closely related to the resistance to flow.

In C.G.S. system, the dynamic viscosity (η) of a liquid is the tangential force in dynes per square centimeter exerted in either of the two parallel planes placed, 1 cm apart when the space between them is filled with the fluid and one of the plane is moving in its own plane with a velocity of 1 cm per second relatively to the other. The unit of dynamic viscosity is the poise (abbreviated p). The centi poise (abbreviated cp) is 1/100th of one poise.

While on the absolute scale, viscosity is measured in poise or centi poise, it is not convenient to use the kinematic scale in which the units are stokes (abbreviated S) and centi-stokes (abbreviated CS). The centistokes is 1/100th of one stoke. The kinematic viscosity of a liquid is equal to the quotient of the dynamic viscosity and the density of the liquid at the same temperature, thus :

$$\text{Kinematic Viscosity} = \frac{\text{Dynamic Viscosity}}{\text{Density}}$$

Viscosity of liquid may be determined by any method that will measure the resistance to shear offered by the liquid.

Absolute viscosity can be measured directly if accurate dimensions of the measuring instruments are known but it is more common practice to calibrate the instrument with a liquid of known viscosity and to determine the viscosity of the unknown fluid by comparison with that of the known.

Procedure: The liquid under test is filled in a U tube viscometer in accordance with the expected viscosity of the liquid so that the fluid level stands within 0.2 mm of the filling mark of the viscometer when the capillary is vertical and the specified temperature is attained by the test liquid. The liquid is sucked or blown to the specified weight of the viscometer and the time taken for the meniscus to pass the two specified marks is measured. The kinematic viscosity in centistokes is calculated from the following equation:

$$\text{Kinematic viscosity} = kt$$

Where k = the constant of the viscometer tube determined by observation on liquids of known kinematic viscosity; t = time in seconds for meniscus to pass through the two specified marks.

3.8. Determination of Total Solids:

Determination of total solids in Asava/ Aristha is generally required. Asava/ Aristha containing sugar or honey should be examined by method 1, sugar or honey free Asava/ Aristha and other material should be examined by method 2.

Method 1: Transfer accurately 50 ml of the clear Asava/ Aristha in an evaporable dish and evaporate to a thick extract on a water bath. Unless specified otherwise, extract the residue with 4 quantities, each of 10 ml, of dehydrated ethanol with stirring and filter. Combine the filtrates to another evaporating dish which has been dried to a constant weight and evaporate nearly to dryness on a water bath, add accurately 1 g of diatomite (dry at 105⁰ for 3 hours and cooled in a desiccator for 30 min), stir thoroughly, dry at 105⁰ for 3 hours, cool the dish in a desiccator for 30 min, and weigh immediately. Deduct the weight of diatomite added, the weight of residue should comply with the requirements stated under the individual monograph.

Method 2: Transfer accurately 50 ml of the clear Asava/ Aristha to an evaporable dish, which has been dried to a constant weight and evaporate to dryness on a water bath, then dry at 105⁰ for 3 hours. After cooling the dish containing the residue in a desiccator for 30 min, weigh it immediately. The weight of residue should comply with the requirements stated under the individual monograph.

3.9. Solubility in Water:

Take 100 ml of distilled water in a *Nessler cylinder* and add air-dried and coarsely powdered drug up to saturation. Then stir the sample continuously by twirling the spatula (rounded end of a microspatula) rapidly. After 1 minute, filter the solution using Hirsch funnel, evaporate the filtrate to dryness in a tared flat bottomed shallow dish and dry at 105⁰ to constant weight and calculate the solubility of the drug in water (wt. in mg/100ml).

3.10. Determination of Saponification Value:

The saponification value is the number of mg of potassium hydroxide required to neutralize the fatty acids, resulting from the complete hydrolysis of 1 g of the oil or fat, when determined by the following method :

Dissolve 35 to 40 g of potassium hydroxide in 20 ml water, and add sufficient alcohol to make 1,000 ml. Allow it to stand overnight, and pour off the clear liquor.

Weigh accurately about 2 g of the substance in a tared 250 ml flask, add 25 ml of the alcoholic solution of potassium hydroxide, attach a reflux condenser and boil on a water-bath for one hour, frequently rotating the contents of the flask cool and add 1 ml of solution of phenolphthalein and titrate the excess of alkali with 0.5 N hydrochloric acid. Note the number of ml required (a). Repeat the experiment with the same quantities of the same reagents in the manner omitting the substance. Note the number of ml required (b) Calculate the saponification value from the following formula:-

$$\text{Saponification Value} = \frac{(b-a) \times 0.02805 \times 1000}{W}$$

Where 'W' is the weight in g of the substance taken.

3.11. Determination of Iodine Value:

The Iodine value of a substance is the weight of iodine absorbed by 100 part by weight of the substance, when determined by one of the following methods:-

Iodine Flasks—The Iodine flasks have a nominal capacity of 250 ml.

- A. Iodine Monochloride Method**—Place the substance accurately weighed, in dry iodine flask, add 10 ml of *carbon tetrachloride*, and dissolve. Add 20 ml of iodine monochloride solution, insert the stopper, previously moistened with solution of potassium iodine and allow to stand in a dark place at a temperature of about 17° or thirty minutes. Add 15 ml of solution of potassium iodine and 100 ml water; shake, and titrate with 0.1 N sodium thiosulphate, using solution of starch as indicator. Note the number of ml required (a). At the same time carry out the operation in exactly the same manner, but without the substance being tested, and note the number of ml of 0.1 N sodium thiosulphate required (b).

Calculate the iodine value from the formula:-

$$\text{Iodine Value} = \frac{(b-a) \times 0.01269 \times 100}{W}$$

Where 'W' is the weight in g of the substance taken.

The approximate weight, in g, of the substance to be taken may be calculated by dividing 20 by the highest expected iodine value. If more than half the available halogen is absorbed, the test must be repeated, a smaller quantity of the substance being used.

Iodine Monochloride Solution: The solution may be prepared by either of the two following methods:

- (1) Dissolve 13 g of iodine in a mixture of 300 ml of carbon tetrachloride and 700 ml of glacial acetic acid. To 20 ml of this solution, add 15 ml of *solution of potassium iodide* and 100 ml of *water*, and titrate the solution with 0.1 N sodium thiosulphate. Pass chlorine, washed and dried, through the remainder of the iodine solution until the amount of 0.1 N sodium thiosulphate required for the titration is approximately, but more than, doubled.

- | | | |
|-----|--|---------|
| (2) | Iodine trichloride | 8 g |
| | Iodine | 9 g |
| | Carbon tetrachloride | 300 ml |
| | Glacial acetic acid, sufficient to produce | 1000 ml |

Dissolve the iodine trichloride in about 200 ml of glacial acetic acid, dissolve the iodine in the carbon tetrachloride, mix the two solutions, and add sufficient glacial acetic acid to produce 1000 ml. Iodine Monochloride Solution should be kept in a stoppered bottle, protected from light and stored in a cool place.

B. Pyridine Bromide Method—Place the substance, accurately weighed, in a dry iodine flask, add 10 ml of *carbon tetrachloride* and dissolve. Add 25 ml of pyridine bromide solution, allow to stand for ten minutes in a dark place and complete the determination described under iodine monochloride method, beginning with the words. Add 15 ml.

The approximate weight in gram, of the substance to be taken may be calculated by dividing 12.5 by the highest expected iodine value. If more than half the available halogen is absorbed the test must be repeated, a small quantity of the substance being used.

Pyridine bromide Solution: Dissolve 8 g pyridine and 10 g of *sulphuric acid* in 20 ml of *glacial acetic acid*, keeping the mixture cool. Add 8 g of *bromine* dissolved in 20 ml of *glacial acetic acid* and dilute to 100 ml with *glacial acetic acid*.

Pyridine bromide Solution should be freshly prepared.

3.12. Determination of Acid Value:

The acid value is the number of mg of *potassium hydroxide* required to neutralize the free acids in 1 g of the substance, when determined by the following method:

Weigh accurately about 10 g of the substance (1 to 5) in the case of a resin into a 250 ml flask and add 50 ml of a mixture of equal volumes of alcohol and solvent ether, which has been neutralized after the addition of 1 ml of solution of phenolphthalein. Heat gently on a water-bath, if necessary until the substance has completely melted, titrate with *0.1 N potassium hydroxide*, shaking constantly until a pink colour which persists for fifteen seconds is obtained. Note the number of ml required. Calculate the acid value from the following formula:

$$\text{Acid Value} = \frac{a \times 0.00561 \times 1000}{W}$$

Where 'a' is the number of ml of *0.1 N potassium hydroxide* required and 'W' is the weight in g of the substance taken.

3.13. Determination of Peroxide Value:

The peroxide value is the number of milliequivalents of active oxygen that expresses the amount of peroxide contained in 1000 g of the substance.

Method

Unless otherwise specified in the individual monograph, weigh 5 g of the substance being examined, accurately weighed, into a 250-ml glass-stoppered conical flask, add 30 ml of a mixture of 3 volumes

of *glacial acetic acid* and 2 volumes of *chloroform*, swirl until dissolved and add 0.5ml volumes of saturated *potassium iodide solution*. Allow to stand for exactly 1 minute, with occasional shaking, add 30 ml of *water* and titrate gradually, with continuous and vigorous shaking, with *0.01M sodium thiosulphate* until the yellow colour almost disappears. Add 0.5 ml of *starch solution* and continue the titration, shaking vigorously until the blue colour just disappears (a ml). Repeat the operation omitting the substance being examined (b ml). The volume of *0.01M sodium thiosulphate* in the blank determination must not exceed 0.1 ml.

Calculate the peroxide value from the expression

$$\text{Peroxide Value} = \frac{10 (a - b)}{W}$$

Where *W* = weight, in g, of the substance.

3.14. Determination of Unsaponifiable Matter:

The unsaponifiable matter consists of substances present in oils and fats, which are not saponifiable by alkali hydroxides and are determined by extraction with an organic solvent of a solution of the saponified substance being examined.

Method

Unless otherwise specified in the individual monograph, introduce about 5 g of the substance being examined, accurately weighed, into a 250-ml flask fitted with a reflux condenser. Add a solution of 2 g of *potassium hydroxide* in 40 ml of *ethanol (95per cent)* and heat on a water-bath for 1 hour, shaking frequently. Transfer the contents of the flask to a separating funnel with the aid of 100 ml of hot water and, while the liquid is still warm, shake very carefully with three quantities, each of 100 ml, of *peroxide-free ether*. Combine the ether extracts in a second separating funnel containing 40 ml of water, swirl gently for a few minute, allow to separate and reject the lower layer. Wash the ether extract with two quantities, each of 40 ml, of water and with three quantities, each of 40 ml, of a 3 per cent w/v solution of *potassium hydroxide*, each treatment being followed by a washing with 40 ml of water. Finally, wash the ether layer with successive quantities, each of 40 ml, of water until the aqueous layer is not alkaline to *phenolphthalein solution*. Transfer the ether layer to a weighed flask, washing out the separating funnel with *peroxide-free ether*. Distil off the ether and add to the residue 6 ml of *acetone*. Remove the solvent completely from the flask with the aid of a gentle current of air. Dry at 100⁰ to 105⁰ for 30 minutes. Cool in a desiccator and weigh the residue. Calculate the unsaponifiable matter as per cent w/w.

Dissolve the residue in 20 ml of *ethanol (95per cent)*, previously neutralised to *phenolphthalein solution* and titrate with *0.1M ethanolic potassium hydroxide*. If the volume of *0.1M ethanolic potassium hydroxide* exceeds 0.2 ml, the amount weighed cannot be taken as the unsaponifiable matter and the test must be repeated.

3.15. Detection of Mineral Oil (Holde's Test):

Take 22 ml of the alcoholic potassium hydroxide solution in a conical flask and add 1ml of the sample of the oil to be tested. Boil in a water bath using an air or water cooled condenser till the solution becomes clear and no oily drops are found on the sides of the flask. Take out the flask from the water bath, transfer the contents to a wide mouthed warm test tube and carefully add 25ml of boiling distilled water along the side of the test tube. Continue shaking the tube lightly from side to side during the addition. The turbidity indicates presence of mineral oil, the depth of turbidity depends on the percentage of mineral oil present.

3.16. Rancidity Test (Kreis Test):

The test depends upon the formation of a red colour when oxidized fat is treated with conc. *hydrochloric acid* and a solution of phloroglucinol in ether. The compound in rancid fats responsible for the colour reaction is epihydrin aldehyde. All oxidized fats respond to the Kreis test and the intensity of the colour produced is roughly proportional to the degree of oxidative rancidity.

Procedure

Mix 1 ml of melted fat and 1 ml of conc. *hydrochloric acid* in a test tube. Add 1 ml of a 1 per cent solution of phloroglucinol in *diethyl ether* and mix thoroughly with the fat-acid mixture. A pink colour formation indicates that the fat is slightly oxidized while a red colour indicates that the fat is definitely oxidized.

3.17. Determination of Alcohol Content:

The ethanol content of a liquid is expressed as the number of volumes of ethanol contained in 100 volumes of the liquid, the volumes being measured at 24.9⁰ to 25.1⁰. This is known as the "percentage of ethanol by volume". The content may also be expressed in g of ethanol per 100 g of the liquid. This is known as the "percentage of ethanol by weight".

Use Method I or Method II, as appropriate, unless otherwise specified in the individual monograph.

Method I

Carry out the method for gas chromatography, using the following solutions. Solution (1) contains 5.0 per cent v/v of ethanol and 5.0 per cent v/v of 1-propanol (internal standard). For solution (2) dilute a volume of the preparation being examined with water to contain between 4.0 and 6.0 per cent v/v of ethanol. Prepare solution (3) in the same manner as solution (2) but adding sufficient of the internal standard to produce a final concentration of 5.0 per cent v/v.

The chromatographic procedure may be carried out using a column (1.5 m x 4 mm) packed with porous polymer beads (100 to 120 mesh) and maintained at 150⁰, with both the inlet port and the detector at 170⁰, and nitrogen as the carrier gas.

Calculate the percentage content of ethanol from the areas of the peaks due to ethanol in the chromatogram obtained with solutions (1) and (3).

Method II

For preparations where the use of Industrial Methylated Spirit is permitted in the monograph, determine the content of ethanol as described in Method I but using as solution (2) a volume of the preparation being examined diluted with water to contain between 4.0 and 6.0 per cent v/v of total ethanol and methanol.

Determine the concentration of methanol in the following manner. Carry out the chromatographic procedure described under Method I but using the following solutions. Solution (1) contains 0.25 per cent v/v of methanol and 0.25 per cent v/v of 1-propanol (internal standard). For solution (2) dilute a volume of the preparation being examined with water to contain between 0.2 per cent and 0.3 per cent v/v of methanol. Prepare solution (3) in the same manner as solution (2) but adding sufficient of the internal standard to produce a final concentration of 0.25 per cent v/v.

The sum of the contents of ethanol and methanol is within the range specified in the individual monograph and the ration of the content of methanol to that of ethanol is commensurate with Industrial Methylated Spirit having been used.

Method III

This method is intended only for certain liquid preparations containing ethanol. Where the preparation contains dissolved substances that may distil along with ethanol Method III B or III C must be followed.

Apparatus

The apparatus (see Fig. 3) consists of a round-bottomed flask (A) fitted with a distillation head (B) with a steam trap and attached to a vertical condenser (C). A tube is fitted to the lower part of the condenser and carries the distillate into the lower part of a 100-ml or 250-ml volumetric flask (D). The volumetric flask is immersed in a beaker (E) containing a mixture of ice and water during the distillation. A disc with a circular aperture, 6 cm in diameter, is placed under the distillation flask (A) to reduce the risk of charring of any dissolved substances.

Method III A

Transfer 25 ml of the preparation being examined, accurately measured at 24.9⁰ to 25.1⁰, to the distillation flask. Dilute with 150 ml of water and add a little pumice powder. Attach the distillation head and condenser. Distil and collect not less than 90 ml of the distillate into a 100-ml volumetric flask. Adjust the temperature to 24.9⁰ to 25.1⁰ and dilute to volume with distilled water at 24.9⁰ to 25.1⁰. Determine the relative density at 24.9⁰ to 25.1⁰. The values indicated in column 2 of Table 17 are multiplied by 4 in order to obtain the percentage of ethanol by volume contained in the preparation. If the specific gravity is found to be between two values, the percentage of ethanol should be obtained by interpolation. After calculation of the ethanol content, report the result to one decimal place.

NOTE – (1) If excessive frothing is encountered during distillation, render the solution strongly acid with phosphoric acid or treat with a small amount of liquid paraffin or silicone oil.

(2) The distillate should be clear or not more than slightly cloudy. If it is turbid or contains oily drops, follow Method IIIC. When steam-volatile acids are present, make the solution just alkaline with 1M sodium hydroxide using solid phenolphthalein as indicator before distillation.

Method III B

Follow this method or the following one if the preparation being examined contains appreciable proportions of volatile materials other than ethanol and water.

Mix 25 ml of the preparation, accurately measured at 24⁰ to 25.1⁰, with about 100 ml of water in a separating funnel. Saturate this mixture with sodium chloride, add about 100 ml of hexane and shake vigorously for 2 to 3 minutes. Allow the mixture to stand for 15 to 20 minutes. Run the lower layer into the distillation flask, wash the hexane layer in the separating funnel by shaking vigorously with about 25 ml of sodium chloride solution, allow to separate and run the wash liquor into the first saline solution. Make the mixed solutions just alkaline with 1M sodium hydroxide using solid phenolphthalein as indicator, add a little pumice powder and 100 ml of water, distil 90 ml and determine the percentage v/v of ethanol by Method IIIA beginning at the words “Adjust the temperature...”.

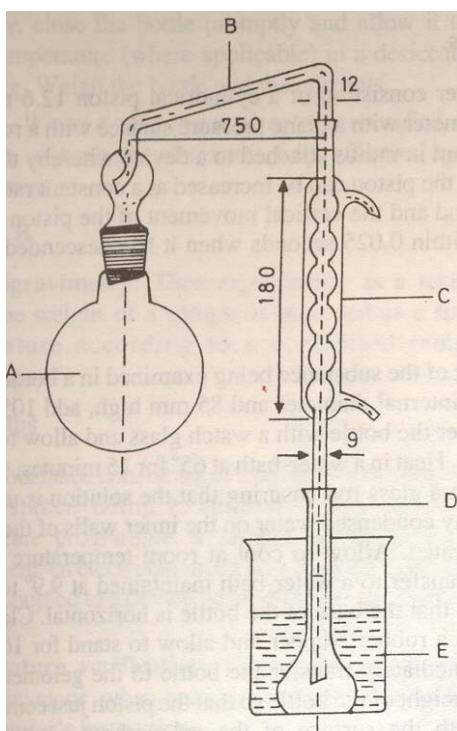


Fig.3 Apparatus for Determination of Ethanol by Distillation Method

Table-17

Specific gravity at 25⁰	Ethanol content*
1.0000	0
0.9985	1
0.9970	2
0.9956	3
0.9941	4
0.9927	5
0.9914	6
0.9901	7
0.9888	8
0.9875	9
0.9862	10
0.9850	11
0.9838	12
0.9826	13
0.9814	14
0.9802	15
0.9790	16
0.9778	17
0.9767	18
0.9756	19
0.9744	20
0.9733	21
0.9721	22
0.9710	23
0.9698	24
0.9685	25

* per cent v/v at 15.56⁰

Method III C

Transfer 25 ml of the preparation, accurately measured at 24.9⁰ to 25.1⁰, to the distillation flask. Dilute with 150 ml of water and add a little pumice powder. Attach the distillation head and condenser. Distil and collect about 100 ml. Transfer to a separating funnel and determine the percentage v/v of ethanol by Method III B beginning at the words “Saturate this mixture...”.

3.18 Disintegration test for tablets:

This test determines whether tablets disintegrate within a prescribed time when placed in a liquid medium under the prescribed experimental conditions.

Disintegration is defined as that state in which no residue of the tablet remains on the screen of the apparatus or, if a residue remains, it consists of fragments of insoluble coating of the tablets. If discs have been used with tablets, any residue remaining on the lower surfaces of the discs consists only of fragments of shells.

Apparatus:

- a) A rigid basket-rack assembly supporting six cylindrical glass tubes, 77.5 ± 2.5 mm long, 21.5 mm in internal diameter and with a wall thickness of about 2 mm.
- b) The tubes are held vertically by two superimposed transparent plastic plates, 90 mm in diameter and 6 mm thick, perforated by six holes having the same diameter as the tubes. The holes are equidistant from the centre of the plate and are equally spaced from one another. Attached to the under side of the lower plate is a piece of woven gauze made from stainless steel wire 635 μm in diameter and having nominal mesh apertures of 2.00 mm. The upper plate is covered with a stainless steel disc perforated by six holes, each about 22 mm in diameter, which fits over the tubes and holds them between the plastic plates. The holes coincide with those of the upper plastic plate and the upper open ends of the glass tubes.
- c) The plates held rigidly in position and 77.5 mm apart by vertical metal rods at the periphery and a metal rod is also fixed to the centre of the upper plate to enable the assembly to be attached to a mechanical device capable of raising and lowering it smoothly at a constant frequency of between 28 and 32 cycles per minutes through a distance of 50 to 60 mm. The design of the basket rack assembly may be somewhat different provided specification for the glass tubes and the screen mesh size are unchanged.
- d) A cylindrical disc for each tube, each 20.7 ± 0.15 mm in a diameter and 9.5 ± 0.15 mm thick, made of transparent plastic with a relative density of 1.18 to 1.20 and pierced with five holes each 2 mm in diameter, one in the centre and the other four spaced equally on a circle of radius 6 mm from the centre of the disc. Four equally spaced grooves are cut in the lateral surface of the disc in such a way that at the upper surface of the disc they are 9.5 mm wide and 2.55 mm deep and at the lower surface 1.6 mm square.
- e) The assembly is suspended in the liquid medium in a suitable vessel, preferably a 1000 ml beaker. The volume of liquid is such that the wire mesh at its highest point is at least 25 mm

below the surface of the liquid, and at its lower point is at least 25 mm above the bottom of the beaker.

- f) A thermostatic arrangement for heating the liquid and maintaining the temperature at $37^{\circ} \pm 2^{\circ}$.

Method:

Introduce one tablet into each tube and add a disc to each tube. Suspend the assembly in the beaker containing the specified liquid and operate the apparatus for the specified liquid and operate the apparatus for the specified time. Remove the assembly from the liquid. The tablets pass the test if all of them have disintegrated.

If 1 or 2 tablets fail to disintegrate, repeat the test on 12 additional tablets; not less than 16 of the total of 18 tablets tested disintegrate.

If the tablets adhere to the disc and the preparation being examined fails to comply, repeat the test omitting the discs. The preparation complies with the test if all the tablets in the repeat test disintegrate.

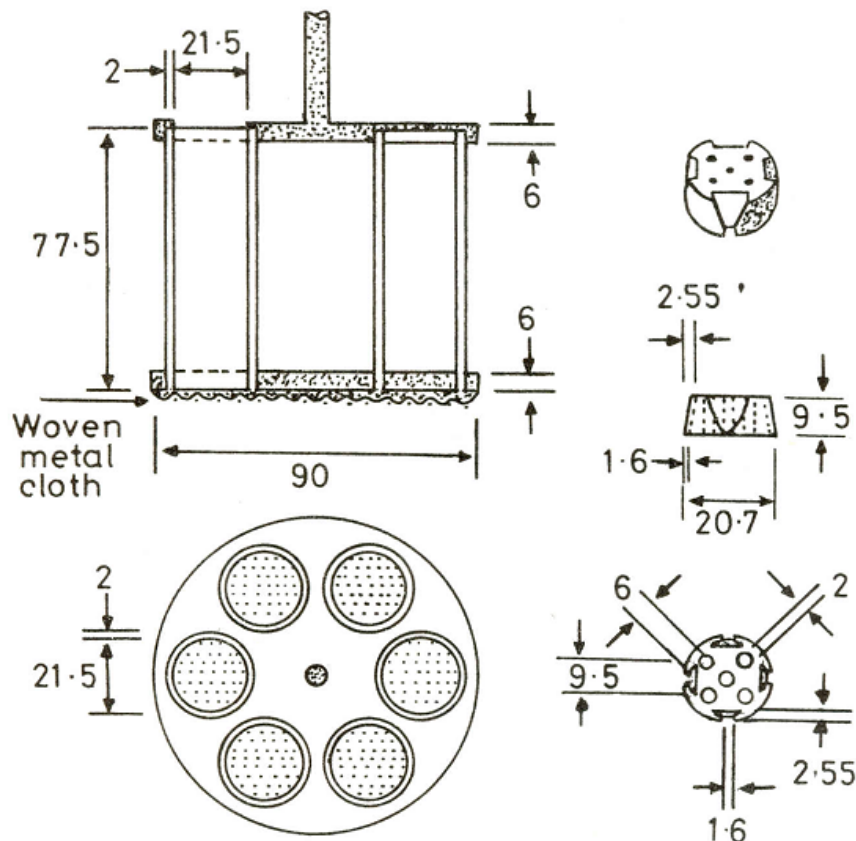


Fig.4. APPARATUS FOR DISINTEGRATION OF TABLETS

3.19 Uniformity of Weight of Single Dose Preparations

Weigh individually 20 units selected at random or, for single dose preparations in individual containers, the contents of 20 units, and calculate the average weight. Not more than two of the individual weights deviate from the average weight by more than the percentage shown in the table and none deviates by more than twice that percentage.

Table-18

Dosage form	Average weight	Percentage deviation
Uncoated and film-coated tablets	80 mg or less	10
	More than 80 mg but less than 250 mg	7.5
	250 mg or more	5

APPENDIX - 4

REAGENTS AND SOLUTIONS

Acetic Acid – Contains approximately 33 per cent w/v of $C_2H_4O_2$. Dilute 315 ml of glacial acetic acid to 1000 ml with *water*.

Acetic Acid, Glacial – $CH_3COOH = 60.05$.

Contains not less than 99.0 per cent w/w of $C_2H_4O_2$. About 17.5 N in strength.

Description – At temperature above its freezing point a clear colourless liquid, odour, pungent and characteristic; crystallises when cooled to about 10^0 and does not completely re-melt until warmed to about 15^0 .

Solubility – Miscible with *water*, with *glycerin* and most fixed and volatile oils.

Boiling range – Between 117^0 and 119^0 .

Congealing temperature – Not lower than 14.8^0 .

Wt. per ml – At 25^0 about 1.047 g.

Heavy metals – Evaporate 5 ml to dryness in a porcelain dish on water-bath, warm the residue with 2 ml of 0.1 N *hydrochloric acid* and water to make 25 ml; the limit of heavy metals is 10 parts per million, Appendix 2.3.3.

Chloride – 5 ml complies with the limit test for chlorides, Appendix 2.3.2.

Sulphate – 5 ml complies with the limit test for sulphates,

Certain aldehydic substances – To 5 ml add 10 ml of *mercuric chloride solution* and make alkaline with *sodium hydroxide solution*, allow to stand for five minutes and acidify with dilute *sulphuric acid*; the solution does not show more than a faint turbidity.

Formic acid and oxidisable impurities – Dilute 5 ml with 10 ml of water, to 5 ml of this solution add 2.0 ml of 0.1 N potassium dichromate and 6 ml of sulphuric acid, and allow to stand for one minute, add 25 ml of water, cool to 15^0 , and add 1 ml of freshly prepared potassium iodide solution and titrate the liberated iodine with 0.1 N sodium thiosulphate, using starch solution as indicator. Not less than 1 ml of 0.1 N *sodium thiosulphate* is required.

Odorless impurities – Neutralise 1.5 ml with *sodium hydroxide solution*; the solution has no odour other than a faint acetous odour.

Readily oxidisable impurities – To 5 ml of the solution prepared for the test for Formic Acid and Oxidisable Impurities, add 20 ml of water and 0.5 ml of 0.1 *N* potassium permanganate; the pink colour does not entirely disappear within half a minute.

Non-volatile matter – Leaves not more than 0.01 per cent w/w of residue when evaporated to dryness and dried to constant weight at 105⁰.

Assay – Weigh accurately about 1 g into a stoppered flask containing 50 ml of water and titrate with *N* sodium hydroxide, using phenolphthalein solution as indicator. Each ml of sodium hydroxide is equivalent to 0.06005 g of C₂H₄O₂.

Acetic Acid, Lead-Free – Acetic acid which complies with following additional test, boil 25 ml until the volume is reduced to about 15 ml, cool make alkaline with lead-free ammonia solution, add 1 ml of lead free potassium cyanide solution, dilute to 50 ml with water, add 2 drops of sodium sulphide solution; no darkening is produced.

Acetone – Propan-2-one; (CH₃)₂CO = 58.08

Description – Clear, colourless, mobile and volatile liquid; taste, pungent and sweetish; odour characteristic; flammable.

Solubility – Miscible with water, with alcohol, with solvent ether, and with chloroform, forming clear solutions.

Distillation range – Not less than 96.0 per cent distils between 55.5⁰ and 57⁰.

Acidity – 10 ml diluted with 10 ml of freshly boiled and cooled water; does not require for neutralisation more than 0.2 ml of 0.1 *N* sodium hydroxide, using phenolphthalein solution as indicator.

Alkalinity – 10 ml diluted with 10 ml of freshly boiled and cooled water, is not alkaline to litmus solution.

Methyl alcohol – Dilute 10 ml with water to 100 ml. To 1 ml of the solution add 1 ml of water and 2 ml of potassium permanganate and phosphoric acid solution. Allow to stand for ten minutes and add 2 ml of oxalic acid and sulphuric acid solution; to the colourless solution add 5 ml of decolorised magenta solution and set aside for thirty minutes between 15⁰ and 30⁰; no colour is produced.

Oxidisable substances – To 20 ml add 0.1 ml of 0.1 *N* potassium permanganate, and allow to stand for fifteen minutes; the solution is not completely decolorised.

Water – Shake 10 ml with 40 ml of carbon disulphide; a clear solution is produced.

Non-volatile matter – When evaporated on a water-bath and dried to constant weight at 105⁰, leaves not more than 0.01 per cent w/v residue.

Acetone Solution, Standard – A 0.05 per cent v/v solution of acetone in water.

Alcohol –

Description – Clear, colourless, mobile, volatile liquid, odour, characteristic and spirituous; taste, burning, readily volatilised even at low temperature, and boils at about 78⁰, flammable. Alcohol containing not less than 94.85 per cent v/v and not more than 95.2 per cent v/v of C₂H₅OH at 15.56⁰.

Solubility – Miscible in all proportions with *water*, with *chloroform* and with *solvent ether*.

Acidity or alkalinity – To 20 ml add five drops of *phenolphthalein solution*; the solution remains colourless and requires not more than 2.0 ml of 0.1N *sodium hydroxide* to produce a pink colour.

Specific gravity – Between 0.8084 and 0.8104 at 25⁰.

Clarity of solution – Dilute 5 ml to 100 ml with *water* in glass cylinder; the solution remains clear when examined against a black background. Cool to 10⁰ for thirty minutes; the solution remains clear.

Methanol – To one drop add one of *water*, one drop of *dilute phosphoric acid*, and one drop of *potassium permanganate solution*. Mix, allow to stand for one minute and add sodium bisulphite solution dropwise, until the permanganate colour is discharged. If a brown colour remains, add one drop of *dilute phosphoric acid*. To the colourless solution add 5 ml of freshly prepared *chromotropic acid* solution and heat on a water-bath at 60⁰ for ten minutes; no violet colour is produced.

Foreign organic substances – Clean a glass-stoppered cylinder thoroughly with *hydrochloric acid*, rinse with *water* and finally rinse with the alcohol under examination. Put 20 ml in the cylinder, cool to about 15⁰ and then add from a carefully cleaned pipette 0.1 ml 0.1 N *potassium permanganate*. Mix at once by inverting the stoppered cylinder and allow to stand at 15⁰ for five minutes; the pink colour does not entirely disappear.

Isopropyl alcohol and t-butyl alcohol – To 1 ml add 2 ml of *water* and 10 ml of *mercuric sulphate solution* and heat in a boiling water-bath; no precipitate is formed within three minutes.

Aldehydes and ketones – Heat 100 ml of *hydroxylamine hydrochloride solution* in a loosely stoppered flask on a water-bath for thirty minutes, cool, and if necessary, add sufficient 0.05 N *sodium hydroxide* to restore the green colour. To 50 ml of this solution add 25 ml of the alcohol and heat on a water bath for ten minutes in a loosely stoppered flask. Cool, transfer to a *Nessler cylinder*, and titrate with 0.05 N *sodium hydroxide* until the colour matches that of the remainder of the *hydroxylamine hydrochloride solution* contained in a similar cylinder, both solutions being viewed down the axis of the cylinder. Not more than 0.9 ml of 0.05 N *sodium hydroxide* is required.

Fusel oil constituents – Mix 10 ml with 5 ml of *water* and 1 ml of *glycerin* and allow the mixture to evaporate spontaneously from clean, odourless absorbent paper; no foreign odour is perceptible at any stage of the evaporation.

Non-volatile matter – Evaporate 40 ml in a tared dish on a water-bath and dry the residue at 105⁰ for one hour; the weight of the residue does not exceed 1 mg.

Storage – Store in tightly-closed containers, away from fire.

Labelling – The label on the container states “Flammable”.

Alcohol, Aldehyde-free. –Alcohol which complies with the following additional test :

Aldehyde – To 25 ml, contained in 300 ml flask, add 75 ml of *dinitrophenyl hydrazine solution*, heat on a water bath under a reflux condenser for twenty four hours, remove the alcohol by distillation, dilute to 200 ml with a 2 per cent v/v solution of sulphuric acid, and set aside for twenty four hours; no crystals are produced.

Alcohol, Sulphate-free. –Shake alcohol with an excess of anion exchange resin for thirty minutes and filter.

Anisaldehyde-sulphuric acid reagent(AS) - Mix 0.5 ml Anisaldehyde with 10 ml glacial acetic acid, followed by 85 ml *methanol* and 5 ml concentrated sulphuric acid.

Ammonia, XN. –Solutions of any normality xN may be prepared by diluting 75 x ml of strong ammonia solution to 1000 ml with water.

Ammonia Solution, Iron-free –Dilute ammonia solution which complies with the following additional test :-

Evaporate 5 ml nearly to dryness on a water-bath add 40 ml of water, 2 ml of 20 per cent w/v *solution of iron free citric acid* and 2 drops of *thioglycollic acid*, mix, make alkaline with *iron-free ammonia solution* and dilute to 50 ml with water, no pink colour is produced.

Ammonium Chloride Solution –A 10.0 per cent w/v solution of *ammonium chloride* in water.

Ammonium molybdate- $\text{NH}_4\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}$ = 1235.86

Analytical reagent grade of commerce.

White crystal or crystalline masses, sometimes with a yellowish or green tint.

Ammonium Thiocyanate – NH_4SCN = 76.12.

Description –Colourless crystals.

Solubility – Very soluble in water, forming a clear solution, readily soluble in alcohol.

Chloride –Dissolve 1 g in 30 ml of solution of hydrogen peroxide, add 1 g of *sodium hydroxide*, warm gently, rotate the flask until a vigorous reaction commences and allow to stand until the reaction is complete; add a further 30 ml of *hydrogen peroxide solution* boil for two minutes, cool, and add 10 ml of *dilute nitric acid* and 1 ml of *silver nitrate solution*; any opalescence produced is not greater than that obtained by treating 0.2 ml of 0.01 N *hydrochloric acid* in the same manner.

Sulphated ash –Moisten 1 g with *sulphuric acid* and ignite gently, again moisten with sulphuric acid and ignite; the residue weighs not more than 2.0 mg.

Ammonium Thiocyanate, 0.1N – $\text{NH}_4\text{SCN} = 76.12$; 7.612 in 1000 ml. Dissolve about 8 g of *ammonium thiocyanate* in 1000 ml of water and standardise the solution as follows :

Pipette 30 ml of standardised 0.1 N *silver nitrate* into a glass stoppered flask, dilute with 50 ml of *water* then add 2 ml of *nitric acid* and 2 ml of *ferric ammonium sulphate solution* and titrate with the *ammonium thiocyanate solution* to the first appearance of a red brown colour. Each ml of 0.1N *silver nitrate* is equivalent to 0.007612 g of NH_4SCN .

Ammonium Thiocyanate Solution – A 10.0 per cent w/v solution of *ammonium thiocyanate solution*.

Anisaldehyde-Sulphuric Acid Reagent – 0.5 ml *anisaldehyde* is mixed with 10 ml *glacial acetic acid*, followed by 85 ml methanol and 5 ml concentrated *sulphuric acid* in that order.

The reagent has only limited stability and is no longer usable when the colour has turned to redviolet.

Arsenomolybdic Acid Reagent- 250 mg of ammonium molybdate was dissolved in 45 ml of distilled water. To this, 2.1 ml of concentrated H_2SO_4 was added and mixed well. To this solution, 3mg of $\text{Na}_2\text{AsO}_4 \cdot 7 \text{H}_2\text{O}$ dissolved in 25 ml of distilled water, mixed well and placed in incubator maintained at 37°C for 24 h.

Borax - Sodium Tetraborate, $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O} = 381.37$.

Contains not less than 99.0 per cent and not more than the equivalent of 103.0 per cent of $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10 \text{H}_2\text{O}$

Description –Transparent, colourless crystals, or a white, crystalline powder; odourless, taste, saline and alkaline. Effloresces in dry air, and on ignition, loses all its water of crystallisation.

Solubility –Soluble in water, practically insoluble in alcohol.

Alkalinity –A solution is alkaline to litmus solution.

Heavy metals – Dissolve 1 g in 16 ml of water and 6 ml of *N hydrochloric acid* and add *water* to make 25 ml; the limit of heavy metals is 20 parts per million, Appendix 2.3.3.

Iron –0.5 g complies with the *limit test for iron*, Appendix 2.3.4.

Chlorides –1 g complies with the *limit test for chlorides*, Appendix 2.3.2.

Sulphates –1 g complies with the *limit test for sulphates*, Appendix 2.3.6.

Assay –Weigh accurately about 3 g and dissolve in 75 ml of *water* and titrate with 0.5 N *hydrochloric acid*, using *methyl red solution* as indicator. Each ml of 0.5 N hydrochloric acid is equivalent to 0.09534 g of $\text{Na}_2\text{B}_4\text{O}_7 \cdot 10\text{H}_2\text{O}$.

Storage – Preserve Borax in well-closed container.

Bromine – Br₂ = 159.80.

Description – Reddish-brown, fuming, corrosive liquid.

Solubility – Slightly soluble in *water*, soluble in most organic solvents.

Iodine – Boil 0.2 ml with 20 ml of *water*, 0.2 ml of *N sulphuric acid* and a small piece of marble until the liquid is almost colourless. Cool, add one drop of *liquefied phenol*, allow to stand for two minutes, and then add 0.2 g of *potassium iodide* and 1 ml of *starch solution*; no blue colour is produced.

Sulphate – Shake 3 ml with 30 ml of *dilute ammonia solution* and evaporate to dryness on a water bath, the residue complies with the *limit test for sulphates*, Appendix 2.3.6.

Bromine Solution – Dissolve 9.6 ml of *bromine* and 30 g of *potassium bromide* in sufficient *water* to produce 100 ml.

Canada Balsam Reagent – General reagent grade of commerce.

Carbon Tetrachloride – CCl₄ = 153.82

Description – Clear, colourless, volatile, liquid; odour, characteristic.

Solubility – Practically insoluble in *water*; miscible with ethyl alcohol, and with solvent ether.

Distillation range – Not less than 95 per cent distils between 76⁰ and 77⁰.

Wt. per ml – At 20⁰, 1.592 to 1.595 g.

Chloride, free acid – Shake 20 ml with 20 ml of freshly boiled and cooled *water* for three minutes and allow separation to take place; the aqueous layer complies with the following test :

Chloride – To 10 ml add one drop of nitric acid and 0.2 ml of *silver nitrate solution*; no opalescence is produced.

Free acid – To 10 ml add a few drops of *bromocresol purple solution*; the colour produced does not indicate more acidity than that indicated by the addition of the same quantity of the indicator to 10 ml of freshly boiled and cooled *water*.

Free chlorine – Shake 10 ml with 5 ml of *cadmium iodide solution* and 1 ml of *starch solution*, no blue colour is produced.

Oxidisable impurities – Shake 20 ml for five minutes with a cold mixture of 10 ml of *sulphuric acid* and 10 ml of 0.1 *N potassium dichromate*, dilute with 100 ml of *water* and add 3 g of *potassium iodide* : the liberated iodine requires for decolourisation not less than 9 ml of 0.1 *N sodium thiosulphate*.

Non-volatile matter – Leaves on evaporation on a water-bath and drying to constant weight at 105⁰ not more than 0.002 per cent w/v of residue.

Caustic Alkali Solution, 5 per cent – Dissolve 5 g of *potassium or sodium hydroxide* in water and dilute to 100 ml.

Charcoal, Decolourising –General purpose grade complying with the following test.

Decolourising powder –Add 0.10 g to 50 ml of 0.006 per cent w/v solution of *bromophenol blue* in ethanol (20 per cent) contained in a 250 ml flask, and mix. Allow to stand for five minutes, and filter; the colour of the filtrate is not deeper than that of a solution prepared by diluting 1 ml of the *bromophenol blue solution* with *ethanol* (20 per cent) to 50 ml.

Chloral Hydrate – $\text{CCl}_3\cdot\text{CH}(\text{OH})_2 = 165.40$.

Description –Colourless, transparent crystals, odour, pungent but not acrid; taste, pungent and slightly bitter, volatilises slowly on exposure to air.

Solubility –Very soluble in *water*, freely soluble in *alcohol*, in chloroform and in *solvent ether*.

Chloral alcoholate – Warm 1 g with 6 ml of *water* and 0.5 ml of *sodium hydroxide solution* : filter, add sufficient 0.1 N *iodine* to impart a deep brown colour, and set aside for one hour; no yellow crystalline precipitate is produced and no smell of iodoform is perceptible.

Chloride – 3 g complies with the limit test for chlorides, Appendix 2.3.2.

Assay – Weigh accurately about 4 g and dissolve in 10 ml of *water* and add 30 ml of N *sodium hydroxide*. Allow the mixture to stand for two minutes, and then titrate with N *sulphuric acid* using *phenolphthalein solution* as indicator. Titrate the neutralised liquid with 0.1 N *silver nitrate* using solution of *potassium chromate* as indicator. Add two-fifteenth of the amount of 0.1 N *silver nitrate* used to the amount of N *sulphuric acid* used in the first titration and deduct the figure so obtained from the amount of N *sodium hydroxide* added. Each ml of N *sodium hydroxide*, obtained as difference; is equivalent to 0.1654 g of $\text{C}_2\text{H}_3\text{Cl}_3\text{O}_2$.

Storage – Store in tightly closed, light resistant containers in a cool place.

Chloral Hydrate Solution –Dissolve 20 g of *chloral hydrate* in 5 ml of water with warming and add 5 ml of *glycerin*.

Chloral Iodine Solution –Add an excess of crystalline *iodine* with shaking to the *chloral hydrate solution*, so that crystals of undissolved iodine remain on the bottom of bottle. Shake before use as the iodine dissolves, and crystals of the iodine to the solution. Store in a bottle of amber glass in a place protected from light.

Chloroform – $\text{CHCl}_3 = 119.38$

Description –Colourless, volatile liquid; odour, characteristic. Taste, sweet and burning.

Solubility –Slightly soluble in water; freely miscible with ethyl alcohol and with solvent ether.

Wt. per ml. : Between 1.474 and 1.478 g.

Boiling range – A variable fraction, not exceeding 5 per cent v/v, distils below 60⁰ and the remainder distils between 50⁰ to 62⁰.

Acidity – Shake 10 ml with 20 ml of freshly boiled and cooled water for three minutes, and allow to separate. To a 5 ml portion of the aqueous layer add 0.1 ml of *litmus solution*; the colour produced is not different from that produced on adding 0.1 ml of *litmus solution* to 5 ml of freshly boiled and cooled water.

Chloride – To another 5 ml portion of the aqueous layer obtained in the test for Acidity, add 5 ml of water and 0.2 ml of *silver nitrate solution*; no opalescence is produced.

Free chlorine – To another 10 ml portion of the aqueous layer, obtained in the test for Acidity, add 1 ml of *cadmium iodide solution* and two drops of starch solution; no blue colour is produced.

Aldehyde – Shake 5 ml with 5 ml of water and 0.2 ml of *alkaline potassium mercuri-iodide solution* in a stoppered bottle and set aside in the dark for fifteen minutes; not more than a pale yellow colour is produced.

Decomposition products – Place 20 ml of the *chloroform* in a glass-stoppered flask, previously rinsed with *sulphuric acid*, add 15 ml of *sulphuric acid* and four drops of *formaldehyde solution*, and shake the mixture frequently during half an hour and set aside for further half an hour, the flask being protected from light during the test; the acid layer is not more than slightly coloured.

Foreign organic matter – Shake 20 ml with 10 ml of *sulphuric acid* in a stoppered vessel previously rinsed with *sulphuric acid* for five minutes and set aside in the dark for thirty minutes, both the acid and chloroform layers remain colourless. To 2 ml of the acid layer add 5 ml of water; the liquid remains colourless and clear, and has no unpleasant odour. Add a further 10 ml of water and 0.2 ml of *silver nitrate solution*; no opalescence is produced.

Foreign odour – Allow 10 ml to evaporate from a large piece of filter paper placed on a warm plate; no foreign odour is detectable at any stage of the evaporation.

Non volatile matter – Not more than 0.004 per cent w/v determined on 25 ml by evaporation and drying at 105⁰.

Storage : Store in tightly-closed, glass-stoppered, light-resistant bottles.

Copper Sulphate – $\text{CuSO}_4 \cdot 5\text{H}_2\text{O} = 249.68$

Contains not less than 98.5 per cent and not more than the equivalent of 101.0 per cent of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$.

Description – Blue triclinic prisms or a blue, crystalline powder.

Solubility – Soluble in *water*, very soluble in boiling water, almost insoluble in *alcohol*; very slowly soluble in glycerin.

Acidity and clarity of solution – 1 g, dissolved in 20 ml of water, forms a clear blue solution, which becomes green on the addition of 0.1 ml of *methyl orange solution*.

Iron – To 5 g, add 25 ml of water, and 2 ml of nitric acid, boil and cool. Add excess of *strong ammonia solution*, filter, and wash the residue with *dilute ammonia solution* mixed with four times its volumes of water. Dissolve the residue, if any, on the filter with 2 ml of *hydrochloric acid*, diluted with 10 ml of water; to the acid solutions add *dilute ammonia solution* till the precipitation is complete; filter and wash; the residue after ignition weighs not more than 7 mg.

Copper Sulphate, Anhydrous – $\text{CuSO}_4 = 159.6$

Prepared by heating copper sulphate to constant weight at about 230° .

Copper Sulphate Solution – A 10.0 per cent w/v solution of *copper sulphate* in water.

Cresol Red – 4,4',-(3H-2, 1-Benzoxathiol-3 ylidene) di-O-cresol SS-dioxide; $\text{C}_{12}\text{H}_8\text{O}_5\text{S} = 382.4$.

Gives a red colour in very strongly acid solutions, a yellow colour in less strongly acid and neutral solutions, and a red colour in moderately alkaline solutions (pH ranges, 0.2 to 1.8, and 7.2 to 8.8).

Cresol Red Solution – Warm 50 ml of cresol red with 2.65 ml of 0.05 M sodium hydroxide and 5 ml of ethanol (90 per cent); after solution is effected, add sufficient ethanol (20 per cent) to produce 250 ml.

Sensitivity – A mixture of 0.1 ml of the solution and 100 ml of *carbon dioxide-free water* to which 0.15 ml of 0.02 M *sodium hydroxide* has been added is purplish-red. Not more than 0.15 ml of 0.02 M *hydrochloric acid* is required to change the colour to yellow.

Disodium Ethylenediamine tetraacetate – (Disodium Acetate) $\text{C}_{10}\text{H}_{14}\text{N}_2\text{Na}_2\text{O}_8 \cdot 2\text{H}_2\text{O} = 372.2$, Analytical reagent grade.

Dragendorff Reagent –

Solution 1 – Dissolve 0.85 g of *bismuth oxy nitrate* in 40 ml of water and 10 ml of acetic acid.

Solution 2 – Dissolve 8 g of *potassium iodide* in 20 ml of water.

Mix equal volumes of solution 1 and 2, and to 10 ml of the resultant mixture add 100 ml of water and 20 ml of acetic acid.

Dragendorffs reagent with tartaric acid:

Solution A: 17 g bismuth sub-nitrate and 200 g tartaric acid in 800 ml water

Solution B: 160 g potassium iodide in 400 ml water.

Stock solution: solution A & solution B

Spray reagent: 50 ml stock solution + 500 ml water + 100 g tartaric acid

Dithizone; 1,5-Diphenylthiocarbazone; Diphenylthiocarbazone; $C_6H_5N:NCSNHNHC_6H_5 = 56.32$

Analytical Reagent grade of commerce.

Almost black powder; mp, about 168^0 , with decomposition.

Store in light-resistant containers.

Eosin – Acid Red 87; Tetrabromofluorescein disodium salt; $C_{20}H_6O_5Br_4Na_2 = 691.86$.

Description – Red powder, dissolves in water to yield a yellow to *purplish-red* solution with a greenish-yellow fluorescence.

Solubility – Soluble in *water* and in alcohol.

Chloride – Dissolve 50 mg in 25 ml of *water*, add 1 ml of *nitric acid*, and filter; the filtrate complies with *the limit test for chlorides*, Appendix 2.3.2.

Sulphated ash – Not more than 24.0 per cent, calculated with reference to the substance dried at 110^0 for two hours, Appendix 2.2.6.

Eosin Solution – A 0.5 per cent w/v solution of eosin in water.

Eriochrome Black T – Mordant Black 11; Sodium 2(1-hydroxy-2-naphthylazo) 5-nitro-2-naphthol-4-sulphonate; $C_{20}H_{12}N_3NaO_7S = 461.38$.

Brownish black powder having a faint, metallic sheen, soluble in alcohol, in *methyl alcohol* and in hot water.

Ethyl Acetate – $CH_3CO_2C_2H_5 = 88.11$.

Analytical reagent grade.

A colourless liquid with a fruity odour; boiling point, about 77^0 ; weight per ml about 0.90g.

Ethyl Alcohol – $C_2H_5OH = 46.07$.

Absolute Alcohol; Dehydrated Alcohol.

Description – Clear, colourless, mobile, volatile liquid; odour, characteristic and spirituous; taste, burning; hygroscopic. Readily volatilisable even at low temperature and boils at 78^0 and is flammable.

Solubility – Miscible with water, with solvent ether and with chloroform.

Contains not less than 99.5 per cent w/w or 99.7 per cent v/v of C_2H_5OH .

Identification – Acidity or Alkalinity: Clarity of Solution; Methanol; Foreign organic substances; Isopropyl alcohol and butyl alcohol; Aldehydes and ketones; fusel oil constituents; Non-volatile matter; complies with the requirements described under Alcohol.

Specific gravity –Between 0.7871 and 0.7902, at 25⁰.

Storage –Store in tightly closed containers in a cool place away from fire and protected from moisture.

Labelling –The label on the container states “Flammable”.

Fehlings Solution –

- A. Dissolve 69.278 g of CuSO₄. 5H₂O in water and make the volume up to 1 litre
- B. Dissolve 100 g of sodium hydroxide and 340 g of Sodium potassium tartarate in water and make the volume to 1 litre.

Mix equal volumes of A and B before the experiment.

Formaldehyde Solution –Formalin; HCHO =30.03

Formaldehyde Solution is a solution of formaldehyde in water with *methyl alcohol* added to prevent polymerisation. It contains not less than 34.0 per cent w/w and not more than 38.0 per cent w/w of CH₂O.

Description – Colourless liquid; odour, characteristic, pungent and irritating; taste, burning. A slight white cloudy deposit is formed on long standing, especially in the cold, due to the separation of paraformaldehyde. This white deposit disappears on warming the solution.

Solubility –Miscible with *water*, and with *alcohol*.

Acidity –To 10 ml add 10 ml of *carbon dioxide free water* and titrate with 0.1 N *sodium hydroxide* using *bromothymol blue solutions* as indicator; not more than 5 ml of 0.1 N *sodium hydroxide* is required.

Wt. per ml – At 20⁰, 1.079 to 1.094 g

Assay –Weigh accurately about 3 g and add to a mixture of 50 ml of *hydrogen peroxide solution* and 50 ml of N *sodium hydroxide*, warm on a water-bath until effervescence ceases and titrate the excess of alkali with N *sulphuric acid* using *phenolphthalein solution* as indicator. Repeat the experiment with the same quantities of the same reagents in the same manner omitting the formaldehyde solution. The difference between the titrations represents the sodium hydroxide required to neutralise the formic acid produced by the oxidation of the formaldehyde. Each ml of N sodium hydroxide is equivalent to 0.03003 g of CH₂O.

Storage–Preserve Formaldehyde Solution in well-closed container preferably at a temperature not below 15⁰.

Formaldehyde Solution, Dilute –

Dilute 34 ml of *formaldehyde solution* with sufficient water to produce 100 ml.

Folin Ciocalteu reagent- Dilute commercially available Folin-Ciocalteu reagent (2N) with an equal volume of distilled water. Transfer it in a brown bottle and store in a refrigerator (4⁰). It should be goldern in colour. Do not use it if it turns olive green.

Formic acid- HCOOH = 46.03

Description:- Colourless liquid, odour, very pungent, highly corrosive; wt per ml. about 1.20 g, contains about 90.0 per cent of HCOOH and is about 23.6 M in strength.

Assay:- Weigh accurately, a conical flask containing 10ml of water, quickly add about 1ml of the reagent being examined and weigh again. Add 50ml of water and titrate with 1M sodium hydroxide using 0.5 ml of *phenolphthalein solution* as indicator. Each ml of 1M sodium hydroxide is equivalent to 0.04603 g of HCOOH.

Gallic acid

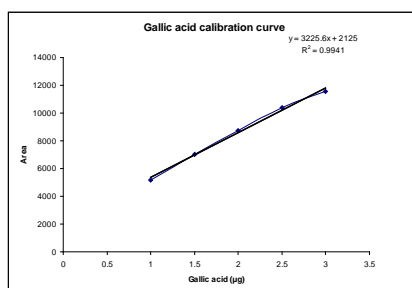
Category: Astringent, styptic

Description: Off white powder; Needles from absolute methanol or chloroform.

Solubility: 1 g dissolves in 87 ml water, 3 ml boiling water, 6 ml alcohol, 100 ml ether, 10 ml glycerol, 5 ml acetone. [Merck Index, 2001, 13th Edn., pp. 722]

Loss on Drying: NMT 10 %

Calibration Curve: Weigh 25 mg of gallic acid and dissolve in 25 ml of methanol. Perform serial dilutions to make the concentrations of 100 µg/ml, 200 µg/ml, 300 µg/ml, 400 µg/ml, 500 µg/ml and 600 µg/ml. Apply 5 µl of each concentration in triplicate on precoated thin layer chromatographic plate of 0.2 mm thickness. Develop the plate in twin trough TLC chamber (Solvent system: Toluene: Ethyl Acetate: Acetic Acid; 5:4:1 % v/v) upto 8 cm. Spray the plate with Natural Products reagent. Dry the plate in a current of cold air and visualize the plate at 366 nm. Scan the plate densitometrically at 366 nm. Record the peak area under curve and plot the calibration curve for gallic acid.



Assay : Contains not less than 0.008 per cent w/v of gallic acid and ethyl gallate, when assayed by the following method:

Glycerine – $C_3H_8O_3 = 82.09$.

Description – Clear, colorless, liquid of syrupy consistency; odourless, taste sweet followed by a sensation of warmth. It is hygroscopic.

Solubility – Miscible with water and with *alcohol*; practically insoluble in chloroform, in solvent ether and in fixed oils.

Acidity – To 50 ml of a 50 per cent w/v solution add 0.2 ml of *dilute phenolphthalein solution*; not more than 0.2 ml of 0.1 N *sodium hydroxide* is required to produce a pink colour.

Wt. per ml – Between 1.252 g and 1.257 g, corresponding to between 98.0 per cent and 100.0 per cent w/w of $C_3H_8O_3$.

Refractive index – Between 1.470 and 1.475 determined at 20°.

Arsenic – Not more than 2 parts per million, Appendix 2.3.1.

Copper – To 10 ml add 30 ml of *water*, and 1 ml of *dilute hydrochloric acid*, and 10 ml of *hydrogen sulphide solution*; no colour is produced.

Iron – 10 g complies with the *limit test* for iron, Appendix 2.3.4.

Heavy metals – Not more than 5 parts per million, determined by Method A on a solution of 4 g in 2 ml of 0.1 N *hydrochloric acid* and sufficient water to produce 25 ml, Appendix 2.3.3.

Sulphate – 1 ml complies with the *limit test* for sulphates, Appendix 2.3.6.

Chloride – 1 ml complies with the *limit test* for chloride, Appendix 2.3.2.

Acetaldehyde and glucose – Heat strongly; it assumes not more than a faint yellow, and not a pink colour. Heat further; it burns with little or no charring and with no odour of burnt sugar.

Aldehydes and related substances – To 12.5 ml of a 50 per cent w/v solution in a glass-stoppered flask add 2.5 ml of *water* and 1 ml of *decolorised magenta solution*. Close the flask and allow to stand for one hour. Any violet colour produced is not more intense than that produced by mixing 1.6 ml of 0.1 N *potassium permanganate* and 250 ml of *water*.

Sugar – Heat 5 g with 1 ml of *dilute sulphuric acid* for five minutes on a water-bath. Add 2 ml of *dilute sodium hydroxide solution* and 1 ml of *copper sulphate solution*. A clear, blue coloured solution is produced. Continue heating on the water-bath for five minutes. The solution remains blue and no precipitate is formed.

Fatty acids and esters – Mix 50 ml with 50 ml of freshly boiled *water* and 50.0 ml of 0.5N *sodium hydroxide*, boil the mixture for five minutes. Cool, add a few drops of *phenolphthalein solution* and titrate the excess alkali with 0.5 N *hydrochloric acid*. Perform a blank determination, not more than 1 ml of 0.5 N *sodium hydroxide* is consumed.

Sulphated ash – Not more than 0.01 per cent, Appendix 2.2.6

Storage –Store in tightly-closed containers.

Glycerin Solution –Dilute 33 ml of *glycerin* to 100 ml with water and add a small piece of camphor or liquid phenol.

n- Hexane:- C_6H_{14} = 86.18

Analytical reagent grade of commerce containing not less than 90.05 of *n-Hexane*

Colourless, mobile, highly flammable liquid, bp 68⁰; wt per ml, about 0.674 g

Hydrochloric Acid – HCl = 36.46

Concentrated Hydrochloric Acid

Description –Clear, colourless, fuming liquid; odour, pungent.

Arsenic –Not more than 1 part per million, Appendix 2.3.1.

Heavy metals –Not more than 5 parts per million, determined by Method A on a solution prepared in the following manner : Evaporate 3.5 ml to dryness on a water-bath, add 2 ml of *dilute acetic acid* to the residue, and add water to make 25 ml, Appendix 2.3.3.

Bromide and iodide –Dilute 5 ml with 10 ml of *water*, add 1 ml of *chloroform*, and add drop by drop, with constant shaking, *chlorinated lime solution*; the chloroform layer does not become brown or violet.

Sulphite –Dilute 1 ml with 10 ml of water, and add 5 drops of *barium chloride solution* and 0.5 ml of 0.001 *N iodine*; the colour of the iodine is not completely discharged.

Sulphate –To 5 ml add 10 mg of sodium bicarbonate and evaporate to dryness on a water bath; the residue, dissolved in *water*; complies with the *limit test for sulphates*, Appendix. 2.3.7.

Free chlorine –Dilute 5 ml with 10 ml of freshly boiled and cooled *water*, add 1 ml of cadmium *iodide solution*, and shake with 1 ml of *chloroform*; the chloroform layer does not become violet within one minute.

Sulphated ash –Not more than 0.01 per cent, Appendix 2.2.6.

Assay –Weigh accurately about 4 g into a stoppered flask containing 40 ml of water, and titrate with *N sodium hydroxide*, using *methyl orange solution* as indicator. Each ml of *N sodium hydroxide* is equivalent to 0.03646 g of HCl.

Storage –Store in glass-stoppered containers at a temperature not exceeding 30⁰.

Hydrochloric Acid, x N –Solution of any normality x N may be prepared by diluting 84 x ml of *hydrochloric acid* to 1000 ml with *water*.

Hydrochloric Acid –(1 per cent w/v) Dilute 1 g of *hydrochloric acid* to 100 ml with *water*.

Dilute Hydrochloric Acid –

Description –Colourless liquid.

Arsenic, Heavy metals bromoide and iodide, Sulphate, free chlorine –Complies with the tests described under Hydrochloric Acid, when three times the quantity is taken for each test.

Assay –Weigh accurately about 10 g and carry out the Assay described under Hydrochloric Acid.

Storage –Store in stoppered containers of glass or other inert material, at temperature below 30⁰.

Hydrochloric Acid, N – HCl = 36.460

36.46 g in 1000 ml

Dilute 85 ml of hydrochloric acid with water to 1000 ml and standardise the solution as follows:

Weigh accurately about 1.5 g of anhydrous sodium carbonate, previously heated at about 270⁰ for one hour. Dissolve it in 100 ml of *water* and add two drops of *methyl red solution*. Add the acid slowly from a burette with constant stirring, until the solution becomes faintly pink. Heat again to boiling and titrate further as necessary until the faint pink colour no longer affected by continued boiling. Each 0.5299 g of *anhydrous* sodium carbonate is equivalent to 1 ml of N hydrochloric acid.

Hydrochloric Acid, Iron-Free –Hydrochloric acid, which complies with the following additional test. Evaporate 5 ml on a water-bath nearly to dryness, add 40 ml of water, 2 ml of a 20 per cent w/v solution of citric acid and two drops of thioglycollic acid, mix, make alkaline with *dilute ammonia solution*, and dilute to 50 ml with water; no pink colour is produced.

Hydrogen Peroxide Solution – (20 Vol.) H₂O₂ = 34.02

Analytical reagent grade of commerce or *hydrogen peroxide solution* (100 Vol.) diluted with 4 volumes of water.

A colourless liquid containing about 6 per cent w/v of H₂O₂; weight per ml, about 1.02 g

Hydroxylamine Hydrochloride; Hydroxylammonium Chloride – NH₂OH.HCl = 69.49.

Contains not less than 97.0 per cent w/w of NH₂OH. HCl

Description – Colourless crystals, or a white, crystalline powder

Solubility –Very soluble in water; soluble in alcohol

Free acid –Dissolve 1.0 g in 50 ml of *alcohol*, add 3 drops of *dimethyl yellow solution* and titrate to the full yellow colour with *N sodium hydroxide*; not more than 0.5 ml of *N sodium hydroxide* is required.

Sulphated ash –Not more than 0.2 per cent, Appendix 2.2.6

Assay –Weigh accurately about 0.1 g and dissolve in 20 ml of water, add 5 g of ferric ammonium sulphate dissolve in 20 ml of water, and 15 ml of *dilute sulphuric acid*, boil for five minutes, dilute with 200 ml of water, and titrate with 0.1 N *potassium permanganate*. Each ml of 0.1 N *potassium permanganate* is equivalent to 0.003475 g of NH₂OH. HCl.

Hydroxylamine Hydrochloride Solution –Dissolve 1 g of *hydroxylamine hydrochloride* in 50 ml of *water* and add 50 ml of *alcohol*, 1 ml of *bromophenol blue solution* and 0.1 N *sodium hydroxide* until the solution becomes green.

Liebermann Burchard reagent –Add carefully 5 ml *acetic anhydride* and 5 ml *conc sulphuric acid* to 50 ml *absolute ethanol*, while cooling in ice. Spray the plate and heat at 105⁰ C for about 10 min. Use freshly prepared reagent.

Mercuric Chloride –HgCl₂ =271.50.

Contains not less than 99.5 per cent of HgCl₂;

Description –Heavy, colourless or white, crystalline masses, or a white crystalline powder.

Solubility –Soluble in *water*; freely soluble in *alcohol*.

Non-volatile matter –When volatilised, leaves not more than 0.1 per cent of residue.

Assay –Weigh accurately about 0.3 g and dissolve in 85 ml of *water* in a stoppered-flask, add 10 ml of *calcium chloride solution*, 10 ml of *potassium iodide solution*, 3 ml of *formaldehyde solution* and 15 ml of *sodium hydroxide solution*, and shake continuously for two minutes. Add 20 ml of acetic acid and 35 ml of 0.1 N *iodine*. Shake continuously for about ten minutes, or until the precipitated mercury is completely redissolved, and titrate the excess of iodine with 0.1 N *sodium thiosulphate*. Each ml of 0.1 N *iodine* is equivalent to 0.01357 g of HgCl₂.

Mercuric Chloride, 0.2 M – Dissolve 54.30 g of *mercuric chloride* in sufficient water to produce 1000 ml.

Mercuric Chloride Solution –A 5.0 per cent w/v solution of *mercuric chloride* in water.

Mercuric Potassium Iodide Solution – See Potassium - Mercuric Iodide solution.

Methyl Alcohol : Methanol : CH₃OH = 32.04.

Description –Clear, Colourless liquid with a characteristic odour.

Solubility –Miscible with water, forming a clear colourless liquid.

Specific Gravity – At 25⁰, not more than 0.791.

Distillation range – Not less than 95 per cent distils between 64.5⁰ and 65.5⁰.

Refractive Index –At 20⁰, 1.328 to 1.329.

Acetone –Place 1 ml in a *Nessler cylinder*, add 19 ml of water, 2 ml of a 1 per cent w/v solution of *2-nitrobenzaldehyde* in *alcohol (50 per cent)*, 1 ml of 30 per cent w/v solution of *sodium hydroxide* and allow to stand in the dark for fifteen minutes. The colour developed does not exceed that produced by mixing 1 ml of standard acetone solution, 19 ml of water, 2 ml of the solution of *2-nitrobenzaldehyde* and 1 ml of the solution of *sodium hydroxide* and allowing to stand in the dark for fifteen minutes.

Acidity –To 5 ml add 5 ml of *carbon dioxide-free water*, and titrate with 0.1 *N sodium hydroxide*, using *bromothymol blue solution* as indicator; not more than 0.1 ml is required.

Non-volatile matter – When evaporated on a water-bath and dried to constant weight at 105n, leaves not more than 0.005 per cent w/v of residue.

Methyl Alcohol, Dehydrated –Methyl alcohol, which complies with the following additional requirement.

Water –Not more than 0.1 per cent w/w.

Methyl Orange – Sodium-*p*-di methylamineazobenzene sulphate, $C_{14}H_{14}O_3N_3SNa$.

An orange-yellow powder or crystalline scales, slightly soluble in cold water; insoluble in alcohol; readily soluble in hot water.

Methyl Orange Solution –Dissolve 0.1 g of methyl orange in 80 ml of water and dilute to 100 ml with alcohol.

Test for sensitivity –A mixture of 0.1 ml of the methyl orange solution and 100 ml freshly boiled and cooled water is yellow. Not more than 0.1 ml of 0.1 *N hydrochloric acid* is required to change the colour to red.

Colour change – pH 3.0 (red) to pH 4.4 (yellow).

Methyl Red – *p*-Dimethylaminoazobenzene-*O*-carboxylic acid, $C_{15}H_{15}O_2N_3$.

A dark red powder or violet crystals, sparingly soluble in *water*; soluble in alcohol.

Methyl red solution –Dissolve 100 mg in 1.86 ml of 0.1 *N sodium hydroxide* and 50 ml of *alcohol* and dilute to 100 ml with water.

Test for sensitivity –A mixture of 0.1 ml of the *methyl red solution* and 100 ml of freshly boiled and cooled *water* to which 0.05 ml of 0.02 *N hydrochloric acid* has been added is red. Not more than 0.01 ml of 0.02 *N sodium hydroxide* is required to change the colour to yellow.

Colour change – pH 4.4 (red) to pH 6.0 (yellow).

Molish's Reagent –Prepare two solutions in separate bottles, with ground glass stoppers:

(a) Dissolve 2 g of α -naphthol in 95 per cent alcohol and make upto 10 ml with alcohol (α -naphthol can be replaced by thymol or resorcinol). Store in a place protected from light. The solution can be used for only a short period.

(b) *Concentrated sulphuric acid.*

Natural Product Reagent (Natural Product-Polyethylene Reagent) –

Solution A-1% ethyl acetate diphenylboric acid-Bethylaminoester(NP),

Solution B-5%ethyl acetatepolyethylene glycol 4000 (PEG)

Spray the plate with 10 ml solution A and 8ml solution B respectively.

Ninhydrin Regent

30mg. Ninhydrin in 10 ml. of n-butanol and 0.3 ml. of glacial acetic acid.

Nitric Acid –Contains 70.0 per cent w/w of HNO_3 (limits, 69.0 to 71.0). About 16 N in strength.

Description –Clear, colourless, fuming liquid.

Wt. per ml. – At 20⁰, 1.41 to 1.42 g.

Copper and Zinc –Dilute 1 ml with 20 ml of water, and add a slight excess of dilute ammonia solution; the mixture does not become blue. Pass hydrogen sulphide; a precipitate is not produced.

Iron –0.5 ml of complies with the limit test for iron, Appendix 2.3.4.

Lead –Not more than 2 parts per million, Appendix 2.3.5.

Chloride –5 ml neutralised with dilute ammonia solution, complies with the limit test for chlorides, Appendix 2.3.2.

Sulphates –To 2.5 ml add 10 mg of sodium bicarbonate and evaporate to dryness on a water-bath, the residue dissolved in water, complies with the limit test for sulphates, Appendix 2.3.7.

Sulphated ash – Not more than 0.01 per cent w/w, Appendix 2.2.6.

Assay –Weigh accurately about 4 g into a stoppered flask containing 40 ml of water, and titrate with N Sodium hydroxide, using methyl orange solution as indicator. Each ml of N sodium hydroxide is equivalent to 0.06301 g of HNO_3 .

Nitric Acid, xN –Solutions of any normality XN may be prepared by diluting 63x ml of nitric acid to 1000 ml with water.

Nitric Acid, Dilute –Contains approximately 10 per cent w/w of HNO_3 . Dilute 106 ml of nitric acid to 1000 ml with water.

Petroleum Light – Petroleum Spirit.

Description – Colourless, very volatile, highly flammable liquid obtained from petroleum, consisting of a mixture of the lower members of the paraffin series of hydrocarbons and complying with one or other of the following definitions :

Light Petroleum –(Boiling range, 30⁰ to 40⁰).

Wt. per ml. –At 20⁰, 0.620 to 0.630 g.

Light Petroleum –(Boiling range, 40⁰ to 60⁰).

Wt. per ml –At 20⁰, 0.630 to 0.650 g.

Light Petroleum –(Boiling range, 60⁰ to 80⁰).

Wt. per ml. –At 20⁰, 0.670 to 0.690.

Light Petroleum –(Boiling range, 80⁰ to 100⁰).

Wt. per ml. –At 20⁰, 0.700 to 0.720

Light Petroleum –(Boiling range, 100⁰ to 120⁰).

Wt. per ml –At 20⁰, 0.720 to 0.740 g.

Light Petroleum –(Boiling range, 120⁰ to 160⁰).

Wt. per ml –At 20⁰, about 0.75 g.

Non-volatile matter –When evaporated on a water-bath and dried at 105⁰, leaves not more than 0.002 per cent w/v of residue.

Phenolphthalein –C₂₀H₁₄O₄.

A white to yellowish-white powder, practically insoluble in water, soluble in alcohol.

Phenolphthalein Solution –Dissolve 0.10 g in 80 ml of *alcohol* and dilute to 100 ml with water.

Test for sensitivity –To 0.1 ml of the *phenolphthalein solution* add 100 ml of freshly boiled and cooled water, the solution is colourless. Not more than 0.2 ml of 0.02 *N sodium hydroxide* is required to change the colour to pink.

Colour change –pH 8.2 (colourless) to pH 10.0 (red)

Phloroglucinol – 1, 3, 5 – Trihydroxybenzene, C₆H₃(OH)₃ . 2H₂O.

Description – White or yellowish crystals or a crystalline powder.

Solubility –Slightly soluble in water; soluble in *alcohol*, and in *solvent ether*.

Melting range –After drying at 110⁰ for one hour, 215⁰ to 219⁰.

Sulphated ash – Not more than 0.1 per cent, Appendix 2.2.6.

Phloroglucinol should be kept protected from light.

Phosphoric Acid – $\text{H}_3\text{PO}_4 = 98.00$.

(Orthophosphoric Acid; Concentrated Phosphoric Acid).

Description – Clear and colourless syrupy liquid, corrosive.

Solubility – Miscible with water and with alcohol.

Phosphoric Acid, x N –

Solutions of any normality, x N may be prepared by diluting 49 x g of *phosphoric acid* with water to 1000 ml.

Phosphoric Acid, Dilute –

Contains approximately 10 per cent w/v of H_3PO_4 .

Dilute 69 ml of *phosphoric acid* to 1000 ml with water.

Potassium Chloride – $\text{KCl} = 74.55$

Analytical reagent grade

Potassium Chromate – $\text{K}_2\text{CrO}_4 = 194.2$

Analytical reagent grade

Potassium Chromate Solution – A 5.0 per cent w/v solution of potassium chromate.

Gives a red precipitate with *silver nitrate* in neutral solutions.

Potassium Cupri-Tartrate Solution – Cupric Tartrate Alkaline Solution: Fehling's Solution.

(1) *Copper Solution* – Dissolve 34.66 g of carefully selected small crystals of *copper sulphate*, showing no trace of efflorescence or of adhering moisture, in sufficient water to make 500 ml. Keep this solution in small, well-stoppered bottles.

(2) *Alkaline Tartrate Solution* – Dissolve 176 g of sodium *potassium tartrate* and 77 g of *sodium hydroxide* in sufficient water to produce 500 ml.

Mix equal volumes of the solutions No. 1 and No. 2 at the time of using.

Potassium Dichromate – $\text{K}_2\text{Cr}_2\text{O}_7 = 294.18$.

Contains not less than 99.8 per cent of $\text{K}_2\text{Cr}_2\text{O}_7$.

Description – Orange-red crystals or a crystalline powder.

Solubility – Soluble in water

Chloride –To 20 ml of a 5 per cent w/v solution in *water* and 10 ml *nitric acid*, warm to about 50° and add a few drops of *silver nitrate solution*; not more than a faint opalescence is produced.

Assay –Carry out the assay described under Potassium Chromate, using 2 g. Each ml of 0.1 *N sodium thiosulphate* is equivalent to 0.004904 g of $K_2Cr_2O_7$.

Potassium Dichromate Solution – A 7.0 per cent w/v solution of *potassium dichromate* in *water*.

Potassium Dichromate, Solution 0.1N – $K_2Cr_2O_7 = 294.18$, 4.903 g in 1000 ml.

Weigh accurately 4.903 g of *potassium dichromate* and dissolve in sufficient *water* to produce 1000 ml.

Potassium Dihydrogen Phosphate - $KH_2PO_4 = 136.1$

Analytical reagent grade of commerce.

Potassium Ferrocyanide – $K_4Fe(CN)_6 \cdot 3H_2O = 422.39$.

Contains not less than 99.0 per cent of $K_4Fe(CN)_6 \cdot 3H_2O$.

Description –Yellow, crystalline powder.

Solubility –Soluble in *water*.

Acidity or Alkalinity –A 10 per cent w/v solution in *water* is neutral to litmus paper.

Assay –Weigh accurately about 1g and dissolve in 200 ml of *water*, add 10 ml of *sulphuric acid* and titrate with 0.1 *N potassium permanganate*. Each ml of 0.1 *N potassium permanganate* is equivalent to 0.04224 g of $K_4Fe(CN)_6 \cdot 3H_2O$.

Potassium Ferrocyanide Solution –A 5.0 per cent w/v solution of *potassium ferrocyanide* in *water*.

Potassium Hydrogen Phthalate – $CO_2H \cdot C_6H_4 \cdot CO_2K = 204.22$.

Contains not less than 99.9 per cent and not more than the equivalent of 100.1 per cent of $C_8H_5O_4K$ calculated with reference to the substance dried at 110° for one hour.

Description –White, crystalline powder.

Solubility –Slowly soluble in *water*, forming clear, colourless solution.

Acidity –A 2.0 per cent w/v solution in carbon dioxide free *water* gives with *bromophenol blue solution* the grey colour indicative of pH 4.0.

Assay –Weigh accurately about 9 g, dissolve in 100 ml of *water* and titrate with *N sodium hydroxide* using *phenolphthalein solution* as indicator. Each ml of *N Sodium hydroxide* is equivalent to 0.2042 g of $C_8H_5O_4K$.

Potassium Hydrogen Phthalate, 0.02 M – Dissolve 4.084 g of *Potassium hydrogen phthalate* in sufficient *water* to produce 1000 ml.

Potassium Hydrogen Phthalate, 0.2 M – Dissolve 40.84 g of *potassium hydrogen phthalate* in sufficient *water* to produce 1000 ml.

Potassium Hydroxide –Caustic Potash : KOH = 56.11

Contains not less than 85.0 per cent of total alkali, calculated as KOH and not more than 4.0 per cent of K_2CO_3 .

Description – Dry white sticks, pellets or fused mass; hard, brittle and showing a crystalline fracture; very deliquescent; strongly alkaline and corrosive.

Solubility –Freely soluble in water, in alcohol and in glycerin; very soluble in boiling *ethyl alcohol*.

Aluminium, iron and matter insoluble in *hydrochloric acid* –Boil 5 g with 40 ml of dilute *hydrochloric acid*, cool, make alkaline with dilute ammonia solution, boil, filter and wash the residue with a 2.5 per cent w/v solution of ammonium nitrate; the insoluble residue, after ignition to constant weight, weighs not more than 5 mg.

Chloride –0.5 g dissolved in water with the addition of 1.6 ml of nitric acid, complies with the limit test for chlorides, Appendix 2.3.2.

Heavy metals –Dissolve 1 g in a mixture of 5 ml of water and 7 ml of dilute hydrochloric acid. Heat to boiling, add 1 drop of phenolphthalein solution and dilute ammonia solution dropwise to produce a faint pink colour. Add 2 ml of acetic acid and water to make 25 ml; the limit of heavy metals is 30 parts per million, Appendix 2.3.3.

Sulphate –Dissolve 1 g in water with the addition of 4.5 ml of hydrochloric acid; the solution complies with the limit test for sulphates, Appendix 2.3.6.

Sodium –To 3 ml of a 10 per cent w/v solution add 1 ml of water, 1.5 ml of alcohol, and 3 ml of potassium antimonate solution and allow to stand; no white crystalline precipitate or sediment is visible to the naked eye within fifteen minutes.

Assay –Weigh accurately about 2 g, and dissolve in 25 ml of water, add 5 ml of barium chloride solution, and titrate with N hydrochloric acid, using phenolphthalein solution as indicator. To the solution in the flask add bromophenol blue solution, and continue the titration with N hydrochloric acid. Each ml of N hydrochloric acid, used in the second titration is equivalent to 0.06911 g of K_2CO_3 . Each ml of N hydrochloric acid, used in the combined titration is equivalent to 0.05611 g of total alkali, calculated as KOH.

Storage –Potassium Hydroxide should be kept in a well-closed container.

Potassium Hydroxide, xN – Solution of any normality, x N, may be prepared by dissolving 56.11x g of potassium hydroxide in water and diluting to 1000 ml.

Potassium Hydroxide Solution –Solution of Potash.

An aqueous solution of potassium hydroxide containing 5.0 per cent w/v of total alkali, calculated as KOH (limits, 4.75 to 5.25).

Assay –Titrate 20 ml with N sulphuric acid, using solution of methyl orange as indicator. Each ml of N sulphuric acid is equivalent to 0.05611 g of total alkali, calculated as KOH.

Storage –Potassium hydroxide solution should be kept in a well-closed container of lead-free glass or of a suitable plastic.

Potassium Iodide –KI = 166.00

Description – Colourless crystals or white powder; odourless, taste, saline and slightly bitter.

Solubility –Very soluble in water and in glycerin; soluble in alcohol.

Arsenic –Not more than 2 parts per million, Appendix 2.3.1.

Heavy metals –Not more than 10 parts per million, determined on 2.0 g by Method A, Appendix 2.3.3.

Barium –Dissolve 0.5 g in 10 ml of water and add 1 ml of dilute sulphuric acid; no turbidity develops within one minute.

Cyanides –Dissolve 0.5 g in 5 ml of warm water, add one drop of ferrous sulphate solution and 0.5 ml of sodium hydroxide solution and acidify with hydrochloric acid; no blue colour is produced.

Iodates –Dissolve 0.5 g in 10 ml of freshly boiled and cooled water, and add 2 drops of dilute sulphuric acid and a drop of starch solution; no blue colour is produced within two minutes.

Assay –Weigh accurately about 0.5 g, dissolve in about 10 ml of water and add 35 ml of hydrochloric acid and 5 ml of chloroform. Titrate with 0.05 M potassium iodate until the purple colour of iodine disappears from the chloroform. Add the last portion of the iodate solution drop-wise and agitate vigorously and continuously. Allow to stand for five minutes. If any colour develops in the chloroform layer continue the titration. Each ml of 0.05 M potassium iodate is equivalent to 0.0166 mg of KI.

Storage –Store in well-closed containers.

Potassium Iodide, M –Dissolve 166.00 g of potassium iodide in sufficient water to produce 1000 ml.

Potassium Iodide and Starch Solution –Dissolve 10 g of potassium iodide in sufficient water to produce 95 ml and add 5 ml of starch solution.

Potassium Iodide and Starch solution must be recently prepared.

Potassium Iodide Solution –A 10 per cent w/v solution of potassium iodide in water.

Potassium Iodobismuthate Solution –Dissolve 100 g of tartaric acid in 400 ml of water and 8.5 g of bismuth oxynitrate. Shake during one hour, add 200 ml of a 40 per cent w/v

Potassium Iodobismuthate Solution, Dilute –Dissolve 100 g of tartaric acid in 500 ml of water and add 50 ml of potassium iodobismuthate solution.

Potassium Mercuric-Iodide Solution –Mayer's Reagent.

Add 1.36 g of mercuric chloride dissolved in 60 ml of water to a solution of 5 g of potassium iodide in 20 ml of water, mix and add sufficient water to produce 100 ml.

Potassium Mercuri-Iodide Solution, Alkaline (Nessler's Reagent)

To 3.5 g of potassium iodide add 1.25 g of mercuric chloride dissolved in 80 ml of water, add a cold saturated solution of mercuric chloride in water, with constant stirring until a slight red precipitate remains. Dissolve 12 g of sodium hydroxide in the solution, add a little more of the cold saturated solution of mercuric chloride and sufficient water to produce 100 ml. Allow to stand and decant the clear liquid.

Potassium Permanganate – $\text{KMnO}_4 = 158.03$

Description –Dark purple, slender, prismatic crystals, having a metallic lustre, odourless; taste, sweet and astringent.

Solubility –Soluble in *water*; freely soluble in *boiling water*.

Chloride and Sulphate –Dissolve 1 g in 50 ml of *boiling water*, heat on a water-bath, and add gradually 4 ml or a sufficient quantity of *alcohol* until the meniscus is colour-less; filter. A 20 ml portion of the filtrate complies with the limit test for *chloride*, Appendix 2.3.2., and another 20 ml portion of the filtrate complies with the limit test for *sulphates*, Appendix 2.3.7.

Assay –Weigh accurately about 0.8 g, dissolve in water and dilute to 250 ml. Titrate with this solution 25.0 ml of 0.1 *N oxalic acid* mixed with 25 ml of *water* and 5 ml of *sulphuric acid*. Keep the temperature at about 70° throughout the entire titration. Each ml of 0.1 *N oxalic acid* is equivalent to 0.00316 g of KMnO_4 .

Storage –Store in well-closed containers.

Caution –Great care should be observed in handling *potassium permanganate*, as dangerous explosions are liable to occur if it is brought into contact with organic or other readily oxidisable substance, either in solution or in the dry condition.

Potassium Permanganate Solution – A 1.0 per cent w/v solution of *potassium permanganate* in water.

Potassium Permanganate, 0.1 N Solution –158.03. 3.161 g in 1000 ml

Dissolve about 3.3. g of *potassium permanganate* in 1000 ml of *water*, heat on a water-bath for one hour and allow to stand for two days. Filter through glass wool and standardise the solution as follows:

To an accurately measured volume of about 25 ml of the solution in a glass stoppered flask add 2 g of *potassium iodide* followed by 10 ml of *N sulphuric acid*. Titrate the liberated *iodine* with standardised 0.1 *N sodium thiosulphate*, adding 3 ml of *starch solution* as the end point is approached. Correct for a blank run on the same quantities of the same reagents. Each ml of 0.1 *N sodium thiosulphate* is equivalent to 0.003161 g of KMnO_4 .

Potassium Tellurite: $\text{K}_2 \text{TeO}_3$ (approx)

General reagent grade of commerce.

Purified Water – $\text{H}_2\text{O} = 18.02$.

Description –Clear, colourless liquid, odourless, tasteless.

Purified water is prepared from potable water by distillation, ion-exchange treatment, reverse osmosis or any other suitable process. It contains no added substances.

pH – Between 4.5 and 7.0 determined in a solution prepared by adding 0.3 ml of a saturated solution of *potassium chloride* to 100 ml of the liquid being examined.

Carbon dioxide –To 25 ml add 25 ml of *calcium hydroxide solution*, no turbidity is produced.

Chloride –To 10 ml add 1 ml of *dilute nitric acid* and 0.2 ml of *silver nitrate solution*; no opalescence is produced, Appendix 2.3.2.

Sulphate –To 10 ml add 0.1 ml of *dilute hydrochloric acid* and 0.1 ml of *barium chloride*, Appendix 2.3.6.

Solution : the solution remains clear for an hour.

Nitrates and Nitrites –To 50 ml add 18 ml of *acetic acid* and 2 ml of *naphthylamine-sulphanilic acid* reagent. Add 0.12 g of *zinc reducing mixture* and shake several times. No pink colour develops within fifteen minutes.

Ammonium – To 20 ml add 1 ml of *alkaline potassium mercuric-iodide solution* and after five minutes view in a Nessler cylinder placed on a white tile; the colour is not more intense than that given on adding 1 ml of *alkaline potassium mercuric-iodide solution* to a solution containing 2.5 ml of *dilute ammonium chloride solution (Nessler's)* 7.5 ml of the liquid being examined.

Calcium –To 10 ml add 0.2 ml of *dilute ammonia solution* and 0.2 ml of *ammonium oxalate solution*; the solution remains clear for an hour.

Heavy metals –Adjust the pH of 40 ml to between 3.0 and 4.0 with *dilute acetic acid*, add 10 ml of freshly prepared *hydrogen sulphide solution* and allow to stand for ten minutes; the colour of the solution is not more than that of a mixture of 50 ml of the liquid being examined and the same amount of *dilute acetic acid* added to the sample, Appendix 2.3.3.

Oxidisable matter –To 100 ml add 10 ml of *dilute sulphuric acid* and 0.1 ml of 0.1 N *potassium permanganate* and boil for five minutes. The solution remains faintly pink.

Total Solids –Not more than 0.001 per cent w/v determined on 100 ml by evaporating on a water bath and drying in an oven at 105⁰ for one hour.

Storage –Store in tightly closed containers.

Silver Nitrate Solution –

A freshly prepared 5.0 per cent w/v solution of silver nitrate in water.

Silver Nitrate, 0.1 N– AgNO₃ = 169. 87; 16.99 g in 1000 ml. Dissolve about 17 g in sufficient *water* to produce 1000 ml and standardise the solution as follows:

Weigh accurately about 0.1 g of *sodium chloride* previously dried at 110⁰ for two hours and dissolve in 5 ml of *water*. Add 5 ml of *acetic acid*, 50 ml of *methyl alcohol* and three drops of *eosin solution* is equivalent to 1 ml of 0.1 N *silver nitrate*.

Sodium Bicarbonate – NaHCO₃ =84.01

Description –White, crystalline powder or small, opaque, monoclinic crystals; odourless; taste, saline.

Solubility –Freely soluble in *water*; practically insoluble in *alcohol*.

Carbonate –pH of a freshly prepared 5.0 per cent w/v solution in *carbon dioxide-free water*, not more than 8.6.

Aluminium, calcium and insoluble matter –Boil 10 g with 50 ml of *water* and 20 ml of *dilute ammonia solution*, filter, and wash the residue with *water*; the residue, after ignition to constant weight, not more than 1 mg.

Arsenic –Not more than 2 parts per million, Appendix 2.3.1.

Iron –Dissolve 2.5 g in 20 ml of *water* and 4 ml of *iron-free hydrochloric acid*, and *dilute* to 40 ml with *water*; the solution complies with the *limit test for iron*, Appendix 2.3.4.

Heavy metals –Not more than 5 parts per million, determined by Method A on a solution prepared in the following manner:

Mix 4.0 g with 5 ml of *water* and 10 ml of *dilute hydrochloric acid*, heat to boiling, and maintain the temperature for one minute. Add one drop of *phenolphthalein solution* and sufficient *ammonia solution* drop wise to give the solution a faint pink colour. Cool and dilute to 25 ml with *water*, Appendix 2.3.3.

Chlorides –Dissolve 1.0 g in *water* with the addition of 2 ml of *nitric acid*; the solution complies with the *limit test for chlorides*, Appendix 2.3.2.

Sulphates –Dissolve 2 g in *water* with the addition of 2 ml of *hydrochloric acid*; the solution complies with the limit test for *sulphates*, Appendix 2.3.6.

Ammonium compounds –1 g warmed with 10 ml of *sodium hydroxide solution* does not evolve ammonia.

Assay –Weigh accurately about 1 g, dissolve in 20 ml of *water*, and titrate with 0.5 N *sulphuric acid* using *methyl orange solutions* as indicator. Each ml of 0.5 N *sulphuric acid* is equivalent to 0.042 g of NaHCO_3 .

Storage –Store in well-closed containers.

Sodium Bicarbonate Solution –A 5 per cent w/v solution of *sodium bicarbonate* in *water*.

Sodium Carbonate – $\text{Na}_2\text{CO}_3 \cdot 10\text{H}_2\text{O} = 286.2$.

Analytical reagent grade

Sodium Chloride – $\text{NaCl} = 58.44$

Analytical reagent grade

Sodium Hydroxide – $\text{NaOH} = 40.00$

Description –White sticks, pellets, fused masses, or scales; dry, hard brittle and showing a crystalline fracture, very deliquescent; strongly alkaline and corrosive.

Solubility –Freely soluble in *water* and in *alcohol*.

Aluminium, iron and matter insoluble in hydrochloric acid –Boil 5 g with 50 ml of dilute hydrochloric acid, cool, make alkaline with *dilute ammonia solution*, boil, filter, and wash with a 2.5 per cent w/v solution of *ammonium nitrate*; the insoluble residue after ignition to constant weight weighs not more than 5 mg.

Arsenic –Not more than 4 parts per million, Appendix 2.3.1.

Heavy metals –Not more than 30 parts per million, determined by Method A, Appendix 2.3.3. in a solution prepared by dissolving 0.67 g in 5 ml of *water* and 7 ml of 3 N *hydrochloric acid*. Heat to boiling, cool and dilute to 25 ml with *water*.

Potassium –Acidify 5 ml of a 5 per cent w/v solution with *acetic acid* and add 3 drops of *sodium cobaltnitrite solution*; no precipitate is formed.

Chloride – 0.5 g dissolved in *water* with the addition of 1.8 ml of *nitric acid*, complies with the limit test for *chlorides*, Appendix 2.3.2.

Sulphates –1 g dissolved in *water* with the addition of 3.5 ml of *hydrochloric acid* complies with the limit test for *sulphates*, Appendix 2.3.6.

Assay –Weigh accurately about 1.5 g and dissolve in about 40 ml of *carbon dioxide-free water*. Cool and titrate with *N sulphuric acid* using *phenolphthalein solution* as indicator. When the pink colour of the solution is discharged, record the volume of acid solution required, add *methyl orange solution* and continue the titration until a persistent pink colour is produced. Each ml of *N sulphuric acid* is equivalent to 0.040 g of total alkali calculated as NaOH and each ml of acid consumed in the titration with *methyl orange* is equivalent to 0.106 g of Na₂CO₃.

Storage –Store in tightly closed containers.

Sodium Hydroxide, xN – Solutions of any normality, xN may be prepared by dissolving 40 x g of *sodium hydroxide* in *water* and diluting to 1000 ml.

Sodium Hydroxide Solution – A 20.0 per cent w/v solution of *sodium hydroxide* in *water*.

Sodium Hydroxide Solution, Dilute –

A 5.0 per cent w/v solution of *sodium hydroxide* in *water*.

Sodium Potassium Tartrate –Rochelle Salt COONa.CH(OH).CH(OH), COOK. 4H₂O = 282.17

Contains not less than 99.0 per cent and not more than the equivalent of 104.0 per cent of C₄H₄O₆KNa. 4H₂O.

Description –Colourless crystals or a white, crystalline powder; odourless; taste saline and cooling. It effloresces slightly in warm, dry air, the crystals are often coated with a white powder.

Solubility –Soluble in *water*; practically insoluble in alcohol.

Acidity or Alkalinity –Dissolve 1 g in 10 ml of recently boiled and cooled *water*, the solution requires for neutralisation not more than 0.1 ml of 0.1 *N sodium hydroxide* or of 0.1 *N hydrochloric acid*, using *phenolphthalein solution* as indicator.

Iron –0.5 g complies with the *limit test for iron*, Appendix 2.3.4.

Chloride –0.5 g complies with the *limit test for chlorides*, Appendix 2.3.2.

Sulphate –0.5 g complies with the *limit test for sulphate*, Appendix 2.3.6.

Assay –Weigh accurately about 2 g and heat until carbonised, cool, and boil the residue with 50 ml of *water* and 50 ml of 0.5 N *sulphuric acid*; filter, and wash the filter with *water*; titrate the excess of acid in the filtrate and washings with 0.5 N *sodium hydroxide*, using *methyl orange solution* as indicator. Each ml of 0.5 N *sulphuric acid* is equivalent to 0.07056 g of $C_4H_4O_6KNa \cdot 4H_2O$.

Sodium Sulphate (anhydrous) – $Na_2SO_4 = 142.04$

Analytical reagent grade of commerce

White, crystalline powder of granules; hygroscopic

Sodium Thiosulphate – $Na_2S_2O_3 \cdot 5H_2O = 248.17$.

Description – Large colourless crystals or coarse, crystalline powder; odourless; taste, saline, deliquescent in moist air and effloresces in dry air at temperature above 33° .

Solubility – Very soluble in *water*; insoluble in *alcohol*.

pH –Between 6.0 and 8.4, determined in a 10 per cent w/v solution.

Arsenic –Not more than 2 parts per million, Appendix 2.3.1.

Heavy metals –Not more than 20 parts per million, determined by Method A, Appendix 2.3.3. in a solution prepared in the following manner : Dissolve 1 g in 10 ml of *water*, slowly add 5 ml of *dilute hydrochloric acid* and evaporate the mixture to dryness on a water-bath. Gently boil the residue with 15 ml of *water* for two minutes, and filter. Heat the filtrate to boiling, and add sufficient *bromine solution* to the hot filtrate to produce a clear solution and add a slight excess of *bromine solution*. Boil the solution to expel the *bromine* completely, cool to room temperature, then add a drop of *phenolphthalein solution* and *sodium hydroxide solution* until a slight pink colour is produced. Add 2 ml of *dilute acetic acid* and dilute with *water* to 25 ml.

Calcium –Dissolve 1 g in 20 ml of *water*, and add a few ml of *ammonium oxalate solution*; no turbidity is produced.

Chloride –Dissolve 0.25 g in 15 ml of 2N *nitric acid* and boil gently for three to four minutes, cool and filter; the filtrate complies with the *limit test for chlorides*, Appendix 2.3.2.

Sulphate and Sulphite –Dissolve 0.25 g in 10 ml of *water*, to 3 ml of this solution add 2 ml of *iodine solution*, and gradually add more *iodine solution*, dropwise until a very faint-persistent yellow colour is produced; the resulting solution complies with the limit test for sulphates, Appendix 2.3.7.

Sulphide –Dissolve 1 g in 10 ml of *water* and 10.00 ml of a freshly prepared 5 per cent w/v solution of *sodium nitroprusside*; the solution does not become violet.

Assay –Weigh accurately about 0.8 g and dissolve in 30 ml of *water*. Titrate with 0.1 *N iodine*, using 3 ml of *starch solution* as indicator as the end-point is approached. Each ml of 0.1 iodine is equivalent to 0.02482 g of $\text{Na}_2\text{S}_2\text{O}_3 \cdot 5\text{H}_2\text{O}$.

Storage –Store in tightly-closed containers.

Sodium Thiosulphate 0.1 N – $\text{Na}_2\text{S}_2\text{O}_3 \cdot 5\text{H}_2\text{O}$. = 248.17, 24.82 g in 1000 ml.

Dissolve about 26 g of *sodium thiosulphate* and 0.2 g of *sodium carbonate* in *carbon dioxide-free water* and dilute to 1000 ml with the same solvent. Standardise the solution as follows :

Dissolve 0.300 g of *potassium bromate* in sufficient *water* to produce 250 ml. To 50 ml of this solution, add 2 g of *potassium iodide* and 3 ml of 2 *N hydrochloric acid* and titrate with the *sodium-thiosulphate solution* using *starch solution*, added towards the end of the titration, as indicator until the blue colour is discharged. Each 0.002784 g of *potassium bromate* is equivalent to 1 ml of 0.1*N sodium thiosulphate*. Note: –Re-standardise 0.1 *N sodium thiosulphate* frequently.

Stannous Chloride – $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$ =225.63.

Contains not less than 97.0 per cent of $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$.

Description –Colourless crystals.

Solubility –Soluble in *dilute hydrochloric acid*.

Arsenic- Dissolve 5.0 g in 10 ml of *hydrochloric acid*, heat to boiling and allow to stand for one hour; the solution shows no darkening when compared with a freshly prepared solution of 5.0 g in 10 ml of *hydrochloric acid*.

Sulphate –5.0 g with the addition of 2 ml of *dilute hydrochloric acid*, complies with the *limit test for sulphates*, Appendix 2.3.7.

Assay –Weigh accurately about 1.0 g and dissolve in 30 ml of *hydrochloric acid* in a stoppered flask. Add 20 ml of *water* and 5 ml of *chloroform* and titrate rapidly with 0.05 *M potassium iodate* until the *chloroform* layer is colourless. Each ml of 0.05 *M potassium iodate* is equivalent to 0.02256 g of $\text{SnCl}_2 \cdot 2\text{H}_2\text{O}$.

Stannous Chloride Solution – May be prepared by either of the two methods given below :

Dissolve 330 g of stannous *chloride* in 100 ml of *hydrochloric acid* and add sufficient *water* to produce 1000 ml.

Dilute 60 ml of *hydrochloric acid* with 20 ml of *water*, add 20 g of tin and heat gently until gas ceases to be evolved; add sufficient *water* to produce 100 ml, allowing the undissolved tin to remain in the solution.

Starch Soluble – Starch, which has been treated with *hydrochloric acid* until after being washed, it forms an almost clear liquid solution in hot water.

Description –Fine, white powder.

Solubility –Soluble in hot *water*, usually forming a slightly turbid *solution*.

Acidity or Alkalinity –Shake 2 g with 20 ml of *water* for three minutes and filter; the filtrate is not alkaline or more than faintly acid to litmus paper.

Sensitivity –Mix 1 g with a little cold *water* and add 200 ml *boiling water*. Add 5 ml of this solution to 100 ml of *water* and add 0.05 ml of 0.1 *N iodine*. The deep blue colour is discharged by 0.05 ml of 0.1 *N sodium thiosulphate*.

Ash – Not more than 0.3 per cent, Appendix 2.3.

Starch Solution –Triturate 0.5 g of *soluble starch*, with 5 ml of *water*, and add this, with constant stirring, to sufficient water to produce about 100 ml. Boil for a few minutes, cool, and filter.

Solution of *starch must be recently prepared*.

Sulphamic Acid – $\text{NH}_2\text{SO}_3\text{H}$ =97.09.

Contains not less than 98.0 per cent of $\text{H}_3\text{NO}_3\text{S}$.

Description –White crystals or a white crystalline powder.

Solubility –Readily soluble in water. Melting Range – 203^0 to 205^0 , with decomposition.

Sulphuric Acid – H_2SO_4 = 98.08.

When no molarity is indicated use analytical reagent grade of commerce containing about 98 per cent w/w of *sulphuric acid*. An oily, corrosive liquid weighing about 1.84 g per ml and about 18 M in strength.

When solutions of molarity xM are required, they should be prepared by carefully adding 54 ml of sulphuric acid to an equal volume of water and diluting with water to 1000 ml.

Solutions of sulphuric acid contain about 10 per cent w/v of H_2SO_4 per g mol.

Sulphuric Acid, Dilute –Contains approximately 10 per cent w/w of H_2SO_4 .

Dilute 57 ml of sulphuric acid to 1000 ml with water.

Sulphuric Acid, Chlorine-free –Sulphuric acid which complies with the following additional test:

Chloride –Mix 2 ml with 50 ml of water and add 1 ml of solution of *silver nitrate*, no opalescence is produced.

Sulphuric Acid, Nitrogen-free–Sulphuric acid which contains not less than 98.0 per cent w/w of H_2SO_4 and complies with the following additional test :

Nitrate –Mix 45 ml with 5 ml of *water*, cool and add 8 mg of *diphenyl benezidine*; the solution is colourless or not more than very pale blue.

Sulphuric acid, methanolic- 10 per cent v/v solution of Sulphuric acid in ice cooled methanol

Tartaric Acid – (CHOH. COOH)₂ =150.1

Analytical reagent grade

Thioglycollic Acid – Mercapto acetic acid, – HS. CH₂COOH =92.11.

Contains not less than 89.0 per cent w/w of C₂H₄O₂S, as determined by both parts of the Assay described below :

Description –Colourless or nearly colourless liquid; odour strong and upleasant.

Iron –Mix 0.1 ml with 50 ml of water and render alkaline with *strong ammonia solution*; no pink colour is produced.

Assay – Weigh accurately about 0.4 g and dissolve in 20 ml of *water* and titrate with 0.1 *N sodium hydroxide* using *cresol red solution* as indicator. Each ml of 0.1 *N sodium hydroxide* is equivalent to 0.009212 g of C₂H₄O₂S.

To the above neutralised solution and 2 g of *sodium bicarbonate* and titrate with 0.1 *N iodine*. Each ml of 0.1 *N iodine* is equivalent to 0.009212 g of C₂H₄O₂S.

Triethanolamine -

Toluene :-Methyl benzene,C₆H₅.CH₃ =102.14.

Analytical grade reagent of commerce.

Clear, colourless liquid, odour, characteristic; bp about 110⁰, wt per ml, about 0.870 g.

Water –See purified water.

Water, Ammonia-free –Water, which has been boiled vigorously for a few minutes and protected from the atomosphere during cooling and storage.

Xylenol Orange – [3H-2,1-Benzoxathiol–3-ylidene bis – (6-hydroxy-5-methyl-m-phenylene) methylenenitrilo] tetra acetic acid SS-dioxide or its tetra sodium salt.

Gives a reddish-purple colour with mercury, lead, zinc and contain other metal ions in acid solution. When metal ions are absent, for example, in the presence of an excess of *disodium ethylenediamine tetraacetate*, this solution is yellow.

Xylenol Orange Solution –Dissolve 0.1 g of *xylenol orange* with 100 ml of *water* and filter, if necessary.

Zinc, Acetate – analytical grade reagent of commerce

APPENDIX - 5

ASSAYS AND CHEMICAL TESTS

5.1. Estimation of Sugars

The method of Lane and Eyonon by reduction of Fehling's solution is the most generally applied volumetric method, the use of methylene blue as an internal indicator increasing the accuracy of the process. If strict attention be given to the details of the procedure and the table-18 are used to calculate the results, a high degree of accuracy can be obtained.

Prepare a solution of the sugar of such concentration that more than 15 ml and less than 50 ml. will be required to reduce all the copper in 10 ml. or 25 ml. of Fehling's solution (0.1 to 0.8 g per 100 ml of dextrose and 0.13 to 1.1 g per 100 ml. of lactose). For preliminary titration, measure accurately 10 ml. or 25 ml. of Fehling's solution into a conical flask of about 300 ml. capacity. From a burette add 15 ml. of the sugar solution and heat to boiling over asbestos covered wire gauze. Continue adding the sugar solution in fairly large portions at fifteen second intervals, until from the colour of the mixture, the copper appears to be nearly reduced; then boil for a minute or two, add 3 to 5 drops of 1 percent methylene blue solution and continue the titration until the blue colour is discharged. Repeat the titration, adding before heating, almost the full amount of sugar solution required to reduce all the copper. After boiling has commenced maintain a moderate degree of ebullition for two minutes and without removing the flame, add 3 to 5 drops of indicator and continue the titration so that it is just complete in a total boiling time of exactly three minutes. The end point is clearly indicated by the disappearance of the blue colour, the solution becoming orange. The flask must not be removed from the gauze at any stage of the titration. The proportion of the various sugars, equivalent to 10 ml of Fehling's solution are given in the table-19

Fehling's solution No. 1 contains 34.64 g of $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ and 0.5 ml of H_2SO_4 in water to 500 ml. No. 2 contains 176 g. of sodium potassium tartrate and 77 g of sodium hydroxide in water to 500 ml. Mix equal volumes of No. 1 and No. 2 solutions immediately before use.

Clarifying agents may be required for dark or turbid solutions, the general reagents being a slight excess of basic or neutral lead acetate or alumina cream, added before adjusting the volume. Alumina cream is prepared by adding a slight excess of ammonia to a saturated solution of alum and washing the precipitate by decantation until almost free from sulphates. Special reagents are used for definite processes and should always be employed where directed. Excess of lead may be removed by addition of anhydrous sodium carbonate to the filtered solution.

Table No. 19

Total reducing sugar required for complete reduction of 10 ml Soxhlet soln to be used in connection with Lane-Eynon general volumetric method

ml of sugar solution required	Solution containing besides invert sugar:-									
	no sucrose		1 g sucrose per 100 ml		5 g sucrose per 100 ml		10 g sucrose per 100 ml		25 g sucrose per 100 ml	
	Invert sugar factor	mg invert sugar per 100 ml	Invert sugar factor	mg invert sugar per 100 ml	Invert sugar factor	mg invert sugar per 100 ml	Invert sugar factor	mg invert sugar per 100 ml	Invert sugar factor	mg invert sugar per 100 ml
15	50.5	336	49.9	333	47.6	317	46.1	307	43.4	289
16	50.6	316	50.0	312	47.6	297	46.1	288	43.4	271
17	50.7	298	50.1	295	47.6	280	46.1	271	43.4	255
18	50.8	282	50.1	278	47.6	264	46.1	256	43.3	240
19	50.8	267	50.2	264	47.6	250	46.1	243	43.3	227
20	50.9	254.5	50.2	251.0	47.6	238.0	46.1	230.5	43.2	216
21	51.0	542.9	50.2	239.0	47.6	226.7	46.1	219.5	43.2	206
22	51.0	231.8	50.3	228.2	47.6	216.4	46.1	209.5	43.1	196
23	51.1	222.2	50.3	218.7	47.6	207.0	46.1	200.4	43.1	187
24	51.2	213.3	50.3	209.8	47.6	198.3	46.1	192.1	42.9	179
25	51.2	204.8	50.4	201.6	47.6	190.4	46.0	184.0	42.8	171
26	51.3	197.4	50.4	193.8	47.6	183.1	46.0	176.9	42.8	164
27	51.4	190.4	50.4	186.7	47.6	176.4	46.0	170.4	42.7	158
28	51.4	183.7	50.5	180.2	47.7	170.3	46.0	164.3	42.7	152
29	51.5	177.6	50.5	174.1	47.7	164.5	46.0	158.6	42.6	147
30	51.5	171.7	50.5	168.3	47.7	159.0	46.0	153.3	42.5	142
31	51.6	166.3	50.6	163.1	47.7	153.9	45.9	148.1	42.5	137
32	51.6	161.2	50.6	158.1	47.7	149.1	45.9	143.4	42.4	132
33	51.7	156.6	50.6	153.3	47.7	144.5	45.9	139.1	42.3	128
34	51.7	152.2	50.6	148.9	47.7	140.3	45.8	134.9	42.2	124
35	51.8	147.9	50.7	144.7	47.7	136.2	45.8	130.8	42.2	121
36	51.8	143.9	50.7	140.7	47.7	132.5	45.8	127.1	42.1	117
37	51.9	140.2	50.7	137.0	47.7	128.9	45.7	123.5	42.0	114
38	51.9	136.6	50.7	133.5	47.7	125.5	45.7	120.3	42.0	111
39	52.0	133.3	50.8	130.5	47.7	122.3	45.7	117.1	41.9	107
40	52.0	130.1	50.8	127.0	47.7	119.2	45.6	114.1	41.8	104
41	52.1	127.1	50.8	123.0	47.7	116.3	45.6	111.2	41.8	102
42	52.1	124.2	50.8	121.0	47.7	113.5	45.6	108.5	41.7	99
43	52.2	121.4	50.8	118.2	47.7	110.9	45.5	105.8	41.6	97
44	52.2	118.7	50.9	115.6	47.7	108.4	45.5	103.4	41.5	94
45	52.3	116.1	50.9	113.1	47.7	106.0	45.4	101.0	41.4	92
46	52.3	113.7	50.9	110.6	47.7	103.7	45.4	98.7	41.4	90
47	52.4	111.4	50.9	108.2	47.7	101.5	45.3	96.4	41.3	88
48	52.4	109.2	50.9	106.0	47.7	99.4	45.3	94.3	41.2	86
49	52.5	107.1	51.0	104.0	47.7	97.4	45.2	92.3	41.4	84
50	52.5	105.1	51.0	102.0	47.7	95.4	45.2	90.4	41.0	82

5.2. Determination of Aluminum:

Solutions:

10 per cent sodium hydroxide solution – Dissolve 10 g *sodium hydroxide* in 100 ml purified water.

EDTA solution 0.05 M – Dissolve 18.6120 g of sodium salt of EDTA in purified water and make up to 1000 ml.

Zinc acetate solution 0.05M:- Dissolve 10.9690 g of *zinc acetate* in 50 ml *purified water* and few drops of *glacial acetic acid* and dilute to 1000 ml.

Acetate buffer 5.5 pH – Dissolve 21.5 g of *sodium acetate* (AR) in 300 ml *purified water* containing 2 ml *glacial acetic acid* and dilute to 1000 ml

Xylenol orange indicator –Dissolve 0.2 g of *xylenol orange indicator* in 100 ml *purified water* with 2 ml *acetic acid*.

Procedure:

Take suitable aliquot from the stock solution in 250 ml beaker. Take 50 ml of 10 per cent *sodium hydroxide solution* in another beaker. Neutralize the aliquot with *sodium hydroxide solution*. Transfer the 10 per cent *sodium hydroxide solution* to aliquot with constant stirring. Add a pinch of *sodium carbonate* into the solution. Boil the content on burner. Cool and filter through Whatman 40 No. filter paper with pulp in 600 ml beaker. Wash the precipitate with hot water 6-8 times. Acidify the filtrate with *dil. hydrochloric acid* and adjust pH 5.5. Add, in excess normally 25 ml 0.05M EDTA solution. Add 25 ml *acetate buffer solution*. Boil the solution; cool and again adjust pH 5 – 5.5. Add 5-6 drops of *xylenol orange indicator*. The colour changes from golden yellow to orange red at the end point. Take 25 ml 0.05 M EDTA solution and run a blank. Each of 1M EDTA is equivalent to 0.05098 g of Al_2O_3 .

5.3. Determination of Borax:

Powder 5-6 g of drug and incinerated at 450° for 3 hours to get it ash. Dissolve the ash in 20 ml. of *purified water* and left for 15 minutes, filter, wash the residue with 80 ml of *purified water* for 4-5 washings. If necessary, shake the contents and titrate with *0.5N hydrochloric acid* using solution of *methyl orange* as an indicator. Each ml of *0.5N hydrochloric acid* is equivalent to 0.09536 g of $Na_2B_4O_7 \cdot 10H_2O$.

5.4. Determination of Calcium:

Solutions:

20 per cent Potassium hydroxide solution – Dissolve 200 g *potassium hydroxide* in *purified water* and make up to 1000 ml.

Ammonia buffer solutions 9.5 pH – Dissolve 67.5 g *ammonium chloride* in 300 ml *purified water*, add 570 ml *ammonia solution* and dilute to 1000 ml.

EDTA (Ethylene Diethyl Tetra Acetic acid) solution 0.05 M – Dissolve 18.6120 g of solution salt of EDTA and in water and make up to 1000 ml.

Triethanolamine 20per cent Solution – 200 ml of triethanolamine, adds 800 ml water and make up to 1000 ml.

Eriochrome Black T indicator 0.1per cent solution – Dissolve 0.10 g indicator in 100 ml of Methanol.

Patterns & Reeders indicators 0.1per cent solution – Dissolve 0.01g indicator in 100 ml of Methanol.

Procedure:

Take one part of filtrate reserved from Iron (Fe) estimation. Add 5 ml Triethanolamine 20 per cent solution. Add a pinch of *Hydroxylamine hydrochloride*. Add 25-30 ml *potassium hydroxide* 20 per cent solution. Add 4-5 drops of Patterns and Reeders indicator, which imparts rose red colour. Titrate the solution against standard EDTA solution. The colour changes from rose red to Prussian blue mark end point.

Each ml of 1M EDTA solution is equivalent to 0.04008 g Calcium.

5.5. Determination of Copper:

Solutions:

Standard 0.1 N sodium thiosulphate solutions

Potassium iodide

Starch 1per cent solution - Dissolve 1 g in water, boil and make up 100ml.

Procedure:

Take suitable aliquot from the stock solution in a beaker. Add approx. 1.0 g sodium fluoride. Add *ammonia solution* and precipitate solution. Add *acetic acid* to dissolve the precipitate. Boil and cool in water bath. Add approx 1.0 g *potassium iodide*. Titrate the liberated iodine against 0.1 N sodium thioisulphate (hypo) solutions by adding *starch solution* as indicator. The liberated iodine colour blackish brown changes to white at the end point. Calculate copper value against 1 ml of hypo solution titrating against standard 1000 ppm copper solution.

Each ml of 1N $\text{Na}_2\text{S}_2\text{O}_3$ solution is equivalent to 0.06357 g of Copper

5.6. Determination of Iron :

Preparation of sample solution:

Ignite a suitable quantity of the sample (in the presence of organic matter) in a crucible in a muffle furnace at 500-550⁰ until the residue is free from organic matter. Moisten with 5-10 ml of

hydrochloric acid, boil for two min, add 30 ml of water, heat on the water bath for few min, filter and wash thoroughly the residue with water and make up to volume in a volumetric flask.

Solutions:

Stannous chloride solution – Dissolve 5 g *stannous chloride* (A.R) in 25 ml Conc. *hydrochloric acid* and dilute to 100 ml (5 per cent solution).

Mercuric chloride – saturated solution in water.

Sulphuric acid + ortho phosphoric acid mixture – take 60 ml water, add 15 ml conc. *sulphuric acid* and 15 ml H_3PO_4 cool and dilute to 1000ml.

Diphenylamine barium sulphonate – Dissolve 0.25 g in 100 ml water.

0.1 N *Standard potassium dichromate solution*. Dissolve 4.9035 g AR grade in water and dilute to 1000 ml.

Procedure:

Take /withdraw a suitable aliquot from the stock solution in 250 ml in duplicate. Dilute to about 100 ml with distilled water. Add 1-2 drops of *methyl red* indicator. Add 1-2 g *ammonium chloride*. Add dil. Ammonium solution till brown precipitate appears. Boil the solution with ppt. for 4-5 minutes. Cool the content and filter through Whatman 41 no. filter paper. Wash the residue with hot water 4-6 times. Dissolve the residue in dil. HCl in 250 ml beaker. Wash with hot water and make the volume to 100 ml approx. Boil the solution on burner. Reduce the Fe^{3+} to Fe^{2+} by adding *stannous chloride solution* drop wise till solution becomes colourless.

Add 1-2 drops of *stannous chloride solution* in excess. Cool the content in water. Add 10-15 ml 10 percent solution of *mercuric chloride*. Add 25 ml acid mixture. Add 2-3 drops of *diphenylamine barium sulphonate indicator*. Add distilled water, if required. Titrate against standard *potassium dichromate solution*. Appearance of violet colour show end point.

Each ml of 1N $K_2Cr_2O_7$ solution is equivalent to 0.05585 g Iron

Each ml of 1N $K_2Cr_2O_7$ solution is equivalent to 0.7985 g Fe_2O_3

5.7. Determination of Magnesium:

Take another part of filtrate reserved from Fe estimation. Add 5 ml *triethanolamine 20 per cent solution*. Add a pinch of *hydroxylamine hydrochloride*. Add 25-30 ml *ammonia buffer 9.5 pH*. Add 4-5 drops of *eriochrome black T* indicator. The colour changes from rose red to blue marks the end point.

Each ml of 1M EDTA solution is equivalent to 0.0409 g of MgO.

5.8. Determination of Mercury:

Powder 0.5 g drug and treat with 7 ml of conc. *nitric acid* and 15 ml of conc. *sulphuric acid* in a kjeldahl flask; heat under reflux gently at first then strongly for 30 minutes. Cool and add 50 ml conc. *nitric acid* boil so as to remove the brown fumes. Continue the addition of *nitric acid* and boiling until the liquid is colourless; cool, wash the condenser with 100 ml of water, remove the flask and add 1.0 per cent *potassium permanganate* solution drop wise until pink colour persists. Decolourize it by adding 6.0 per cent *hydrogen peroxide* drop wise to remove excess of *potassium permanganate* followed by 3.0 ml of conc. *nitric acid* and titrate with 0.1N *ammonium thiocyanate solutions* using *ferric alum* as indicator.

Each ml. of 0.1N NH_4SCN solution is equivalent to 0.01003 g Mercury.

5.9. Determination of Silica (SiO_2)

Weigh 0.5 g (in case of high silica) or 1.0 g (low silica) finely powdered and dried sample in a platinum crucible (W_1). Add 4-5 g *anhydrous sodium carbonate* into the crucible. Mix thoroughly and cover the crucible with lid, if necessary. Place the crucible in muffle furnace. Allow the temperature to rise gradually to reach $900-950^\circ$ and keep on this temp. for about 1 hour to complete the fusion. Take out the crucible and allow cool at room temperature. Extract the cooled mass in 25-30 ml dil *hydrochloric acid* in 250 ml beaker. Heat on hot plate/burner to dissolve the contents. Wash the crucible with distilled water. Keep the beaker on water bath and allow dry the mass. Dehydrate back and powder the mass. Take out the beaker and allow cooling at room temperature. Add 25-30 ml *hydrochloric acid* dilute to 100 ml distilled water. Boil the content and allow cool. Filter through Whatman no 40. filter paper. Wash the residue with hot water 6-8 times. Place the residue along with filter paper in platinum crucible. Ignite at $900-950^\circ$ for 2-3 min. Allow to cool and weigh as SiO_2 .

5.10. Estimation of Sodium and Potassium by Flame Photometer:

Preparation of Standard solutions

Weigh 2.542 g of AR *sodium chloride* and dissolve in *purified water* and make upto 1000 ml in a volumetric flask. Dilute 1 ml of the stock solution to 100 ml. This gives standard of 1mg of sodium per 100 ml (10 ppm). Prepare 20, 30, 40 and 50 ppm standard solution.

Weigh 1.9115g of AR *potassium chloride* and dissolve in *purified water* and make up to 1000 ml in a volumetric flask. Dilute 1ml of the stock solution to 100ml. This gives standard of 1mg of sodium per 100 ml (10 ppm). Prepare 20, 30, 40 and 50 ppm standard solution.

Preparation of Sample solution

Weigh 10 g of sample in a preweighed silica dish and heat in a muffle furnace for 1hr at 600° . Cool and dissolve the ash in *purified water* and make up to 100ml in a volumetric flask.

Switch on the instrument first and then the pump. Keep distilled water for aspiration and allow it to stand for 15 min (warming time). Open the glass cylinder and ignite the flame. Adjust the instrument to zero.

Introduce the maximum concentration solution and adjust it to 100. Again introduce the purified water so that instrument shows zero. Then introduce the standard solution in ascending concentration. Note down the reading each time. Introduce the purified water for aspiration in between the standard solutions. Introduce the sample solution and if it is within the range take the reading. If it exceeds limit 100 then dilute the solution till the reading is within the range. Plot the curve with concentration in ppm against reading obtained. Find out the concentration of the sample solution. Take two or three readings and find out the average. Find out the concentration of sodium and potassium.

5.11. Determination of Sodium Chloride:

Dissolve about 2-3g accurately weighed drug in 25 ml of *purified water* and left for 30 minutes, filter. Wash the filter paper completely with *purified water* and the filtrate is made 100 ml in volumetric flask, make the solution homogeneous, titrate 25 ml of this solution with 0.1 *N silver nitrate solution* using *potassium chromate* as indicator. The end point shows the light brick red colour.

Each ml. of 0.1 *N Ag NO₃* solution is equivalent to 0.005845 g of NaCl.

5.12. Determination of Sulphur:

Solution:

Carbon tetrachloride saturated with *Bromine*

Barium chloride – 10 per cent solution in water.

Procedure:

Take 0.5 – 1 g powdered sample in 250 ml beaker. Add 10 ml *carbon tetrachloride* saturated with bromine. Keep in cold condition in fume chamber over night. Add 10 – 15 ml conc. *nitric acid*. Digest on water bath. Add 10 ml conc. *hydrochloric acid*, digest it to expel nitrate fumes till syrupy mass. Cool and extract with *hydrochloric acid*, make volume to 100 ml. Boil and filter through Whatman No 40. filter paper. Wash the residue with hot water. Filter through Whatman 41 No. paper in 600 ml beaker. Acidify the filtrate with *hydrochloric acid*. Add 20 ml of 10 per cent *Barium chloride* solution. Stir the solution and digest on burner. Allow to settle BaSO₄ precipitate over night. Filter the precipitate through Whatman No. 42 filter paper. Wash the precipitate with water. Ignite the precipitate in muffle furnace in pre weighed platinum crucible up to 850⁰. Allow to cool and weigh.

Each g of weight of precipitate is equivalent to 0.13734 g of Sulphur.

5.13. Qualitative Reactions :

5.13.1 Sodium

5.13.1.a. Sodium compounds, moistened with hydrochloric acid and introduced on a platinum wire into the flame of a Bunsen burner, give a yellow colour to the flame.

5.13.1.b. Solutions of sodium salts yield, with solution of uranyl zinc acetate, a yellow crystalline precipitate.

5.13.2. Potassium

5.13.2.a. Potassium compounds moistened with hydrochloric acid and introduced on platinum wire into the flame of a Bunsen burner, give a violet colour to the flame.

5.13.2.b. Moderately strong solutions of potassium salts, which have been previously ignited to remove ammonium salts, give a white, crystalline precipitate with perchloric acid.

5.13.2.c. Solutions of potassium salts, which have been previously ignited to free them from ammonium salts and from which iodine has been removed, give a yellow precipitate with solution of sodium cobaltinitrite and acetic acid.

5.13.3. Magnesium

5.13.3.a. Solutions of magnesium salts yield a white precipitate with solution of ammonium carbonate, especially on boiling, but yield no precipitate in the presence of solution of ammonium chloride.

5.13.3.b. Solutions of magnesium salts yield a white crystalline precipitate with solution of sodium phosphate in the presence of ammonium salts and dilute ammonia solution.

5.13.3.c. Solutions of magnesium salts yield with solution of sodium hydroxide a white precipitate insoluble in excess of the reagent, but soluble in solution of ammonium chloride.

5.13.3.d. Solutions of magnesium salts in 1N hydrochloric acid, yield blue precipitate with solutions of 1 drop of magnesium II reagent and 3 ml 1N sodium hydroxide solution.

5.13.4. Carbonates and Bicarbonates

5.13.4.a. Carbonates and bicarbonates effervesce with dilute acids, liberating carbon dioxide; the gas is colourless and produces a white precipitate in solution of calcium hydroxide.

5.13.4.b. Solutions of carbonates produce a brownish-red precipitate with solution of mercuric chloride; Solutions of bicarbonates produce a white precipitate.

5.13.4.c. Solutions of carbonates yield, with solution of silver nitrate, a white precipitate which becomes yellow on the addition of an excess of the reagent and brown on boiling the mixture. The precipitate is soluble in dilute ammonia solution and in dilute nitric acid.

5.13.4.d. Solutions of carbonates produce, at room temperature, a white precipitate with solution of magnesium sulphate. Solutions of bicarbonates yield no precipitate with the reagent at room temperature, but on boiling the mixture a white precipitate is formed.

5.13.4.e. Solutions of bicarbonates, on boiling, liberate carbon dioxide which produces a white precipitate in solution of calcium hydroxide.

5.13.5. Sulphates

5.13.5.a. Solutions of sulphates yield, with solution of barium chloride, a white precipitate insoluble in hydrochloric acid.

5.13.5.b. Solutions of sulphates yield, with solution of lead acetate, a white precipitate soluble in solution of ammonium acetate and in solution of sodium hydroxide.

5.13.6. Chlorides

5.13.6.a. Chlorides, heated with manganese dioxide and sulphuric acid, yield chlorine, recognisable by its odour and by giving a blue colour with potassium iodide and solution of starch.

5.13.6.b. Solutions of chlorides yield, on acidification with dilute nitric acid, a curdy white precipitate with few drops of 5 per cent w/v silver nitrate solution.

5.13.7. Calcium

Solutions of calcium salts yield, with solution of ammonium carbonate, a white precipitate which after boiling and cooling the mixture, is insoluble in solution of ammonium chloride.

5.13.8. Sulphides

Solutions of sulphides yield, with solution of 5 per cent w/v silver nitrate solution, a black precipitate.

Dissolve 1 g of sample in 10 ml of deionised water and filter. Add 4 ml of 5 per cent w/v silver nitrate solution to the filtrate. A black precipitate appears.

5.13.9. Test for Mercury :

Weight about 0.1 g of the formulation, dissolve in hydrochloric acid and filter. Pass hydrogen sulphide gas to the filtrate, a precipitate appears. Filter the solution and wash the precipitate with water and redissolve in dilute hydrochloric acid/nitric acid. The presence of Mercury is indicated by appearance of violet colour when added to dithizone solution.

5.13.10. Test for Boron :

Dissolve 0.1 g of formulation in 10 ml of water and filter. Add 1 ml. of carminic acid (0.5 percent in concentrate sulphuric acid) to 1 ml. of filtrate. Blue colour indicates the presence of Boron.

5.13.11. Test for Sulphur:

Extract 1.0 g of the sample with 10 ml of *carbon disulphide*. Filter the carbon disulphide solution and evaporate the solvent. To the residue add 10 ml of 10 % *alcoholic potash* and boil until the sulphur is dissolved. Dilute with water, oxidize by adding *hydrogen peroxide solution* (~6 per cent w/v of H₂O₂) in excess and heat on a water bath for 30 min. Acidify with *hydrochloric acid*, filter and to the filtrate add *barium chloride solution* (10 % w/v in water). A White precipitate indicates the presence of sulphur.

5.13.12. Test for anthraquinones :

Add 0.5 g of formulation in 50 ml of water, boil until nearly dissolved & cool the solution. Add 0.5 g of Kieslguhr and filter. Apply following test to the filtrate.

5.13.13. Test for Hingu:

Boil 0.2 g of formulation with 2 ml *hydrochloric acid* for about 1 minute, cool, dilute with an equal volume of water and filter into 3 ml of dilute solution of *Ammonia*, if fluorescence is produced.

Heat 5 ml of filtrate with 0.2 g of *borax*, add few drops of this solution to a test tube nearly filled with water a light green fluoresces produced.

APPENDIX-6

AYURVEDIC DEFINITIONS AND METHODS

6.1.Kalpanā Paribhāṣā:

6.1.1. Kalka:

Kalka is the fine paste of macerated fresh plant material.

(*Paribhāṣā Prabandha*)

6.1.2. Kvātha / Kaṣāya:

Kvātha or *Kaṣāya* is the filtered liquid obtained by boiling coarse powder of drug(s) in proportion of 4, 8 or 16 [*Mṛdu Dravya* - 4, *Madhyama Dravya* - 8 and *Kaṭhina Dravya* - 16 respectively] times of water and reduced to one-fourth.

(*Śārṅgadhara saṃhitā* - II - 9/3)

6.1.3. Cūrṇa:

The fine sieved powder of well dried drug(s) is called *Cūrṇa*.

(*Śārṅgadhara saṃhitā* - II - 6/1)

6.1.4. Puṭapāka Svarasa:

It is a kind of procedure, where juice of fresh green herb will be obtained by the process of *Puṭapāka*. Bundle the *Kalka* of green plant material in leaves of *Kāśmarī*, *Vaṭa*, *Jambū* etc., and cover with clay in layers of about 2 cm thickness. Dry and place amidst fire till becomes reddish. Open the bundle and strain the juice from *Kalka* through a *muslin cloth*.

(*Śārṅgadhara saṃhitā* - II - 1/21-23)

6.1.5. Svarasa:

The liquid part of fresh macerated plant material obtained by pressing through a fresh, *muslin cloth* is called as *Svarasa*.

(*Śārṅgadhara saṃhitā* - II - 1/2)

6.1.6. Hima Kaṣāya:

Hima Kaṣāya is the extractive obtained by straining of 48 g [1 part] of powdered drug(s) soaked in 288 ml [6 parts] of water overnight.

(*Śārṅgadhara saṃhitā* - II - 4/1)

6.2. Sāmānya paribhāṣā:

6.2.1. Kajjalī:

Kajjalī is the fine black colored powder obtained by triturating *Gandhaka* (Sulphur) and *Pārada* (Mercury) without adding any liquid.

(*Rasatarāṅgiṇī* - 2/27)

6.2.2. Kāñjika:

Sour liquid prepared with of rice grain etc. is called as *Kāñjika*. Take *ṣaṣṭika śāli* in an earthen vessel, add five parts of water and boil. Shift the preparation into another earthen vessel, add three parts of water and seal the mouth of the vessel tightly. Place the vessel aside for a period of two to three weeks at regulated temperature during which the liquid becomes sour.

(*Paribhāṣā Prabandha*)

6.2.3. Kṣāra Preparation:

- Cut the drug into small pieces, wash, clean and dry well.
- Put few pieces in an earthen pot or iron vessel and ignite. As they burn, add more and more pieces till all the pieces burn completely and reduce to ash. Allow the ash to cool.
- Add *water* to the ash in the ratio of 6: 1, mix well and allow to stand overnight.
- Next day decant the supernatant liquid, strain repeatedly through *muslin cloth* till a clear filtrate is obtained.
- Take the filtrate in a stainless steel vessel and heat it over moderate flame to evaporate the water to obtain a solid salty white residue (*Kṣāra*), at the bottom of the vessel. Allow to cool completely.
- Store it in amber coloured glass containers and pack them air-tight to protect from light and moisture.

6.2.4. Cūrṇodaka:

1	Cūrṇa (Lime powder)	250 mg
2	Water	60 ml

Take 250 mg of lime powder in a stainless steel vessel, add 60 ml of water and keep aside for 9 h. Decant the supernatant layers through a filter paper. The filtrate is known as *Cūrṇodaka*.

(*Rasatarāṅgiṇī* – 11)

6.2.5. Mastu Preparation:

Tie *Godadhi* in a *muslin cloth* and allow the liquid part of it to get separated to obtain the *Mastu*.

6.2.6. Prakṣepa:

Fine powder form of the drug(s), which is added to a *kalpa* such as *Leha*, *Āsava Ariṣṭa* etc. before administration is known as *Prakṣepa*.

6.2.7. Bhāvanā:

Bhāvanā is the process by which powders of drugs are levigated to a soft mass with specified liquids and allowed to dry.

(*Rasataraṅgiṇī* - 2/49)

6.2.8. Śodhana:

Śodhana is the process which removes the impurities to some extent and helps in increasing the therapeutic values of the drugs.

6.2.8.1. Godantī Śodhana:

1	Godantī		1 part
2	Nimbu Svarasa	Fr.	QS
		or	
3	Droṇapuṣpī Svarasa	Pl.	QS

Bundle small pieces of *Godantī* in a cloth, suspend in *Nimbu* or *Droṇapuṣpī svarasa* in a *Dolāyantra*, and boil for 3 h

(*Rasataraṅgiṇī* - 11/239)

6.2.8.2. Gairika Śodhana:

1	Gairika	1 part
2	Godugdha	QS

Fine powder of *Gairika* is to be levigated with Cow's milk.

(*Rasaratnasamuccaya* - 3/49)

or

1	Gairika	1 part
2	Goghṛta	QS

Fry the fine powder of *Gairika* in little amount of *Ghṛta*.

(*Āyurveda prakāśa* - 2/272)

6.2.8.3. Gandhaka Śodhana:

1	Gandhaka	1 part
---	----------	--------

2	Godugdha		QS
		or	
3	Bhṛṅgarāja Svarasa	Pl.	QS

Melt small pieces of *Gandhaka* in an iron pan smeared with *Ghṛta* and pour in to a pot containing *Godugdha* or *Bhṛṅgarāja svarasa*. Collect after cooling. Repeat the process for seven times. At the end of the seventh process, wash and dry the material.

(*Rasāmṛtam* - 2/3)

6.2.8.4. Guggulu Śodhana:

Remove manually the big pieces of sandstone, glass, wood etc. if any from the *Guggulu*. Cut *Guggulu* into small pieces, bundle in a cloth and immerse in *Dolāyantra* containing any one of the following liquids.

Gomūtra
Godugdha
Triphalā kaṣāya
Vāsā kaṣāya / svarasa or
Nirguṇḍī svarasa with Haridrā cūrṇa

Boil till the whole amount of *Guggulu* passes into the liquid through the cloth. Discard the residue present in the bundle if any.

Filter the liquid through *muslin cloth* and heat the mixture till a semi solid mass is obtained.

Dry in sun and store until further use.

6.2.8.5. Ṭaṅkaṇa Śodhana:

Take small pieces of *Ṭaṅkaṇa* in an iron pan, fry till complete dehydration.

(*Āyurveda prakāśa* - 2/244)

6.2.8.6. Tuttha Śodhana:

1	Tuttha		1 part
2	Raktacandana kvātha	Ht. Wd.	QS
3	Mañjiṣṭhā kvātha	Rt.	QS
4	Triphalā kvātha	P.	QS

Prepare fine powder of *Tuttha* and levigate with the individual liquid medias number (ii) to (iv) mentioned above seven times each.

(*Rasāmṛtam* - 3/74)

6.2.8.7. Bhallātaka Śodhana:

1	Bhallātaka	Fr.	1 part
---	------------	-----	--------

2	Gomūtra		QS
3	Godugdha		QS
4	Iṣṭikā cūrṇa		QS
5	Water		QS

Method of Preparation:-Take *Bhallātaka*, remove the attached thalamus and soak in *Gomūtra* for 7 days. Replace *Gomūtra* every 24 h with fresh *Gomūtra*. After 7 days, rinse the *Bhallātaka* twice with water, to wash off the *Gomūtra*. Soak *Bhallātaka* in *Godugdha* for 7 days, replacing *Godugdha* every 24 h with fresh *Godugdha*. After 7 days, rinse the *Bhallātaka* 2 or 3 times with water to wash off the *Godugdha*. Put the *Bhallātaka* in a thick jute bag containing coarse brick powder and rub carefully, with a view to reduce the oil content in *Bhallātaka*. Wash the processed seeds with water and dry.

(*Rasāmṛtam- Pariśiṣṭa*)

6.2.8.8. Manaḥśilā Śodhana:

1	Manaḥśilā		1 part
2	Agastya patra svarasa	Lf.	QS
		or	
3	Śṛṅgavera (Ārdraka) Svarasa	Pl.	QS

Prepare fine powder of *Manaḥśilā* and levigate with any one of the above specified liquid media for seven times.

(*Rasaratnasamuccaya - 3/93*)

6.2.8.9. Vatsanābha Śodhana:

1	Viṣa (Vatsanābha)	Rt. Tr.	1 part
2	Gomūtra		QS

Take small pieces of *Vatsanābha*, bundle in *muslin cloth*, and soak in *Gomūtra* for three days, replacing the later every day. Wash the processed material and dry.

(*Rasāmṛtam- Pariśiṣṭa*)

6.2.8.10. Karavīra Śodhana:

1	Karavīraka (Karavīra)	Rt.	1 part
2	Godugdha		QS

Take small pieces of *Karavīra*, bundle in *muslin cloth*, and perform *Svedana* in *Dolāyantra* with *Godugdha* for 2 h. Wash the processed material and dry.

(*Śārṅgadhara Saṃhitā, Madhyamakhaṇḍa Adhyāya 12:300*)

6.2.8.11. Citraka Śodhana:

1	Rakta Citraka (Citraka)	Rt.	1 part
---	-------------------------	-----	--------

2	Cūrṇodaka		QS
---	-----------	--	----

Take small pieces of *Karavīra*, soak in *Cūrṇodaka*. Wash the processed material and dry.

(*Rasataraṅgiṇī* 24:575)

6.2.8.12. Lāṅgalī Śodhana:

1	Lāṅgalī	Rt.	1 part
2	Gomūtra		QS

Take small pieces of *Karavīra*, soak in *Gomūtra* for 24 h. Wash the processed material and dry.

(*Āyurvedaparakāśa* 6)

6.2.8.13. Śilājatu Śodhana:

1	Śilājatu		2 parts
2	Hot water		4 parts
3	Triphalā Kvātha	P.	1 part

Take powder of *Śilājatu*, add specified amounts of hot water and *Triphalā Kvātha* so as to disengage the soluble matter. Allow to settle down and decant the supernatant layers.

Repeat the process till a clear liquid is obtained.

Concentrate the decanted material to thick paste over moderate heat.

Dry in sun rays and preserve for further purpose.

(*Rasataraṅgiṇī* 22/69-78)

6.2.8.14. Haritāla Śodhana:

1	Haritāla		1 part
2	Kūṣmāṇḍa Toya	Fr.	QS
		or	
3	Tila Kṣāra Jala	Pl.	QS
		or	
4	Cūrṇodaka		QS

Take small pieces of *Haritāla*, bundle in clean muslin cloth, suspend in a *Dolāyantra* containing any one of the above liquid media. Boil for three hours, dry in sun rays and preserve for further purpose.

(*Rasaratnasamuccaya* - 3/70)

6.2.8.15. Hiṅgu Śodhana:

1	Rāmaṭha (Hiṅgu)	Exd.	1 part
2	Ājya (Goghṛta)		QS

Prepare fine powder of *Hiṅgu* and fry it in sufficient amounts of *Goghṛta*, till it becomes crisp.

(*Rasataraṅgiṇī*, 24/578)

6.2.8.16. Vijayā Śodhana:

1	Vijayā	1 part
2	Jala	QS

Put *Vijayā* in a *muslin cloth bag* and wash in water till free from turbidity and dry.

(*Rasāmṛtam, Pariśiṣṭa* 8/147)

6.2.8.17. Kāśīśa Śodhana:

1	Kāśīśa	1 part
2	Bhṛṅga nīra (Bhṛṅgarāja)	QS

Bhāvanā is given with *Bhṛṅgarāja rasa*, 3 times.

(*Rasāmṛtam, Adhyāya* 3/158)

6.2.8.18. Sauvīrāñjana Śodhana:

1	Sauvīrāñjana API	Lead sulphide	1 part
2	Bhṛṅgarājadrava (Bhṛṅgarāja API)	<i>Eclipta alba</i> Pl. juice	QS

Method

Powder *Sauvīrāñjana* and give *Bhāvanā* with *Bhṛṅgarāja svarasa* for 7 days.

(*Rasaratnasamuccaya, Adhyāya* 3; 107)

6.2.8.19. Narasāra Śodhana:

1	Narasāra	API	Sal ammoniac	1 part
2	Jala	API	Potable water	3 parts

Method

Add 3 times Jala to *Narasāra*, make a solution and filter. Boil the solution till water is evaporated.

(*Rasataranṅī, Taraṅga* 14; 3-4.)

6.2.8.20. Pārada Sāmānya Śodhana:

1	Pārada (Mercury)	2 parts
2	Sudhāraja (Lime powder)	2 parts
3	Rasona	2 parts
4	Saindhava Lavaṇa (Rock salt)	1 part

Take equal parts of *Pārada* and *Sudhāraja*, triturate for three days, and filter carefully through a clean cloth.

Add dehusked *Rasona* and *Saindhava lavaṇa* to the *Pārada*, triturate till the paste of *Rasona* becomes black.

Wash with warm water and separate the *Pārada* with caution.

(*Rasatarāṅgiṇī* - 5/ 27-30)

6.2.8.21. Aṣṭasaṃskāra of Pārada

Aṣṭasaṃskāra of *Pārada* have been prescribed in Ayurvedic classics for purification and to increase the therapeutic activities.

6.2.8.21.a. Svedana:

(*Rasaḥṛdayatantra* - 2/3)

1	Pārada (Mercury)		1 part
2	Āsurī (Rājikā)	Sd.	1/16 th part
3	Paṭu (Saindhava Lavaṇa)		1/16 th part
4	Śuṅṭhī	Rz.	1/16 th part
5	Marica	Fr.	1/16 th part
6	Pippalī	Fr	1/16 th part
7	Citraka	Rt.	1/16 th part
8	Ārdraka	Rz.	1/16 th part
9	Mūlaka	Rt. Tr.	1/16 th part
10	Kāñjika		QS

Method:

Take the ingredients numbered 2 to 9 in to wet grinder and grind with sufficient quantity of water to prepare *kalka* (homogeneous blend). Take leaf of *Bhūrja* (*Betula utilis*) or *Kadalī* (*Musa paradisiacal*), place it over four folded cloth, smear with the prepared *Kalka*, and gently place *Pārada* over it. Place the remaining part of *Kalka* if any, over the *Pārada*. Suspend the *poṭṭalī* in a *Dolāyantra* containing *Kāñjika*. Boil for three days. Remove *Pārada* and *Kalka*, wash carefully with warm water and collect *Pārada*.

6.2.8.21.b Mardana:

(*Rasaḥṛdayatantra* - 2/4)

1	Pārada (Mercury)		1 part
2	Guḍa		1/16 th part
3	Dagdhoraṇā		1/16 th part
4	Lavaṇa (Saindhava Lavaṇa)		1/16 th part
5	Mandira dhūma		1/16 th part
6	Iṣṭikā cūrṇa		1/16 th part
7	Āsurī (Rājikā)		1/16 th part
8	Kāñjika		QS

Method:

Take the ingredients numbered 1 to 7 in *Khalvayantra*, add with required amounts of *Kāñjika* and levigate for three days. Remove *Pārada* and *Kalka*, wash carefully with warm water and collect *Pārada*.

6.2.8.21.c. Mūrcchana:

(*Rasahṛdayatantra* - 2/6)

1	Pārada (Mercury)		1 part
2	Gṛhakanyā (Kumārī)	Lf.	1/16 th part
3	Harītakī	P.	1/16 th part
4	Bibhītaka	P.	1/16 th part
5	Āmalakī	P.	1/16 th part
6	Citraka	Rt.	1/16 th part

Method:

Take the ingredients numbered 3 to 6, dry, powder and pass through 180 µm I. S. sieve. Add ingredient number 2 and grind with sufficient quantity of water to prepare *Kalka*. Add *Pārada* to the *Kalka* and triturate for three days.

6.2.8.21.d. Utthāpana:

(*Rasahṛdayatantra* - 2/7)

1	Pārada (Mercury)	1 part
2	Kāñjika	QS

Method:

Collect the *Pārada* at the end of *Mūrcchana* process and subject it to *Utthāpana*, and wash with *Kāñjika* and collect the *Pārada* carefully.

6.2.8.21.e. Pātana:

The process of *Pātana* is again of three types, viz. *Ūrdhvapātana*, *Adhaḥpātana* and *Tiryakpātana*.

Ūrdhvapātana:

(*Āyurveda prakāśa* - 1/68-71)

1	Pārada (Mercury)	3 parts
2	Ravi (Tāmra)	1 part

3 Jambīra svarasa (Nimbu)

QS

Method:

Take *Pārada* and *Tāmra* in the specified ratio and levigate with *Jambīra svarasa* to prepare thick paste. Apply the paste over the lower pot of *±amaru yantra* and apply heat for 12 h. Collect the *Pārada* settled at the upper pot gently.

Adhaḥpātana:

(*Āyurveda prakāśa* - 1/75-77)

1	Pārada (Mercury)		1 part
2	Harītakī	P.	1/16 th part
3	Bibhītaka	P.	1/16 th part
4	Āmalakī	P.	1/16 th part
5	Śigru	St. Bk.	1/16 th part
6	Citraka	Rt.	1/16 th part
7	Saindhava Lavaṇa		1/16 th part
8	Āsurī (Rājikā)	Sd.	1/16 th part
9	Nimbu rasa		QS

Method:

Take the ingredients numbered 2 to 6, dry, powder and pass through 180 µm I. S. sieve. Add the powders to *Pārada* and levigate by adding ingredients numbered 7 to 9 to prepare fine paste. Apply the paste in the *Adhaḥpātana yantra*, subject to heat and collect *Pārada*.

Tiryakpātana:

(*Āyurveda prakāśa* - 1/79-81)

1	Pārada (Mercury)		3 parts
2	Ravi (Tāmra)		1 part
3	Jambīra svarasa (Nimbu)		QS

Method:

Take *Pārada* obtained at the end of *Adhaḥpātana* process, add with *Tāmra* and levigate with *Jambīra svarasa* to prepare thick paste. Apply the paste in the *Tiryakpātana yantra*, subject to heat and collect *Pārada*.

6.2.8.21.f. Rodhana / Bodhana:

(*Rasendracūḍāmaṇi* - 4/88)

1	Pārada (Mercury)		3 parts
2	Saindhava Lavaṇa jala		QS

Method:

Place the *Pārada* in a pot containing *Saindhava lavaṇa jala* and seal the mouth of the pot tightly. Place the pot undisturbed for three days. Decant the water on the fourth day to collect the *Pārada*.

6.2.8.21.g. Niyāmana:

(*Rasahr̥dayatantra* - 2/10)

1	Pārada (Mercury)		1 part
2	Phaṇi (Nāgavallī)	Lf.	1/16 th part
3	Laśuna	Bl.	1/16 th part
4	Ambujā		1/16 th part
5	Karkoṭī		1/16 th part
6	Mārkaḥva (Bhṛṅgarāja)	Pl.	1/16 th part
7	Ciñcikā (Ciñcā)	Lf.	1/16 th part
8	Kāñjī		QS

Method:

Prepare *Kalka* of the ingredients numbered 2 to 7, add with *Pārada* and prepare a *poṭṭalī*. Suspend the *poṭṭalī* in a *Dolāyantra* containing *Kāñjika* and boil. Remove *Pārada* and *Kalka*, wash carefully with warm water and collect *Pārada*.

6.2.8.21. h. Dīpana / Sandīpana:

(*Rasahr̥dayatantra* - 2/11)

1	Pārada (Mercury)		1 part
2	Bhū (Sphaṭikā)		1/16 th part
3	Khaga (Kāśīśa)		1/16 th part
4	Ṭaṅkaṇa		1/16 th part
5	Marica	Fr.	1/16 th part
6	Lavaṇa (Saindhava Lavaṇa)		1/16 th part
7	Āsurī (Rājikā)	Sd.	1/16 th part
8	Śigru		1/16 th part
9	Kāñjika		QS

Method:

Prepare *Kalka* of the ingredients numbered 2 to 8, add with *Pārada* and prepare a *poṭṭalī*. Suspend the *poṭṭalī* in a *Dolāyantra* containing *Kāñjika* and boil for three days. Remove *Pārada* and *kalka*, wash carefully with warm water and collect *Pārada*.

6.2.9. Mūrchanā:

Mūrcchana is the process which removes *Āma doṣa* of *Taila / Ghṛta* and provides good color and fragrance. *Mūrcchana* process is to be followed before any *Sneha* preparation.

6.2.9.1. Mūrcchanā of Eraṇḍa Taila:

(*Bhaiṣajyaratnāvalī, Jvarādhikāra*)

Ingredients:

1	Mañjiṣṭhā API	<i>Rubia cordifolia</i>	P.	12 g
2	Mustā API	<i>Cyperus rotundus</i>	Rz.	12 g
3	Dhānyaka API	<i>Coriandrum sativum</i>	Sd.	12 g
4	Āmalakī API	<i>Emblica officinalis</i>	P.	12 g
5	Harītakī API	<i>Terminalia chebula</i>	P.	12 g
6	Bibhītaka API	<i>Terminalia belerica</i>	P.	12 g
7	Agnimantha API	<i>Clerodendron phlomidis</i> (Official substitute)	Rt.	12 g
8	Hrīvera API	<i>Coleus vettiveroides</i>	Rt.	12 g
9	Kharjūra API	<i>Phoenix sylvestris</i>	Fr.	12 g
10	Vaṭa API	<i>Ficus religiosa</i>	Lf. Bud.	12 g
11	Haridrā API	<i>Curcuma longa</i>	Rz.	12 g
12	Dāruharidrā API	<i>Berberis aristata</i>	St.	12 g.
13	Nalikā (Tvak API)	<i>Cinnamomum tamala</i>	St. Bk.	12 g.
14	Śuṅṭhī API	<i>Zingiber officinale</i>		12 g.
15	Ketakī API	<i>Pandanus odoratissimus</i>	Rt.	12 g.
16	Dadhī API	Curd		1,536 l
17	Kāñjika API			1,536 l
18	Eraṇḍa taila API	Castor oil	Oil	768 ml

Method of preparation:

Take all ingredients of pharmacopoeial quality.

Wash, clean, dry the ingredients numbered 1 to 15 of the formulation composition powder separately and pass through 180 μm I. S. sieve (*Kalka dravyas*).

Transfer the powdered ingredients to wet grinder, grind with sufficient quantity of water to prepare *Kalka* (homogeneous blend).

Take *Eraṇḍa taila* in a stainless steel vessel and heat it mildly.

Add increments of *Kalka*. Stir thoroughly while adding *Dadhī* and *Kāñjika*.

Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.

Start the heating next day and observe the boiling mixture for appearance of froth (*phenodgama*) and constantly check the *Kalka* for formation of *varti* (*madhyama pāka lakṣaṇa*).

Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture. Stop heating when the *Kalka* forms a *varti* and the froth subsides. Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.

Pack it in tightly closed glass containers to protect from light and moisture.

6.2.9.2. Mūrchanā of Ghṛta:

(*Bhaiṣajyaratnāvalī*, *Jvarādhikāra*)

Ingredients:

1	Pathyā (Harītakī API)	<i>Terminalia chebula</i>	P.	48 g
2	Dhātrī (Āmalakī API)	<i>Emblica officinalis</i>	P.	48 g
3	Bibhīta (Bibhītaka API)	<i>Terminalia belerica</i>	P.	48 g
4	Jaladhara (Mustā API)	<i>Cyperus rotundus</i>	Rz.	48 g
5	Rajanī (Haridrā API)	<i>Curcuma longa</i>	Rz.	48 g
6	Mātuluṅga API - drava	<i>Citrus medica</i>	Fr.	48 g
7	Ghṛta (Goghṛta API)	<i>Clarified butter of cow's milk</i>		768 g
8	Jala API	Water		3.072 l

Method of Preparation:

Take all ingredients of pharmacopoeial quality.

Wash, clean, dry the ingredients numbered 1 to 5 of the formulation composition powder separately and pass through 180 μm I. S. sieve (*Kalka dravyas*).

Wash, clean the *Mātuluṅga* and separate the juicy flesh from its rind. Grind and filter through *muslin cloth* to obtain *Svarasa*.

Transfer the *Kalka dravyas* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend.

Take *Ghṛta* in a stainless steel vessel and heat mildly.

Add increments of *Kalka*. Stir thoroughly while adding *Svarasa* and water.

Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.

Start the heating next day and observe the boiling mixture for subsidence of froth (*phenaśānti*) and constantly check the *Kalka* for formation of *varti* (*madhyama pāka lakṣaṇa*).

Expose the *varti* and *Ghṛta* to flame and confirm the absence of crackling sound indicating absence of moisture. Stop heating when the *Kalka* forms a *varti* and the froth subsides. Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.

Pack it in tightly closed glass containers to protect from light and moisture.

Description:

A yellow-coloured, soft, low melting medicated fat, unctuous to touch with odour and taste of *Haridrā*

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.439,	Appendix 3.1
<i>Weight per ml at 40⁰:</i>	0.967,	Appendix 3.2
<i>Saponification value:</i>	229,	Appendix 3.10
<i>Iodine value:</i>	100,	Appendix 3.11
<i>Acid value:</i>	Not more than 0.33,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 1.35,	Appendix 3.13
<i>Congealing point:</i>	28 ⁰ to 18 ⁰ ,	Appendix 3.4.2

6.2.9.3. Mūrccana of Sarṣapa Taila:*(Bhaiṣajyaratnāvalī, Jvarādhikāra)***Ingredients:**

1	Vayaḥsthā (Harītakī API)	<i>Terminalia chebula</i>	P.	12 g
2	Rajani (Haridrā API)	<i>Curcuma longa</i>	Rz.	12 g
3	Musta (Mustā API)	<i>Cyperus rotundus</i>	Rz.	12 g
4	Bilva API	<i>Aegle marmelos</i>	Fr. Pp.	12 g
5	Dāḍima API	<i>Punica granatum</i>	Dr. Sd.	12 g
6	Keśara (Nāgakeśara API)	<i>Mesua ferrea</i>	Stmn.	12 g
7	Kṛṣṇājīraka API	<i>Carum carvi</i>	Fr.	12 g
8	Hrīvera API	<i>Coleus vettiveroides</i>	Rt.	12 g
9	Nalikā (Tvak API)	<i>Cinnamomum tamala</i>	St. Bk.	12 g
10	Bibhītaka API	<i>Terminalia belerica</i>	P.	12 g
11	Aruṇā (Mañjiṣṭhā API)	<i>Rubia cordifolia</i>	Rt.	96 g
12	Toya (Jala API)	Water		3.072 l
13	Kaṭutaila (Sarṣapa Taila API)	Mustard oil		768 ml

Method of Preparation:

Take all ingredients of pharmacopoeial quality.

Wash, clean, dry the ingredients numbered 1 to 10 of the formulation composition powder separately and pass through 180 μm I. S. sieve (*Kalka dravyas*).

Transfer the *Kalka dravyas* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend.

Take *Sarṣapa taila* in a stainless steel vessel and heat mildly.

Add increments of *Kalka*. Stir thoroughly while adding water.

Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.

Start the heating next day and observe the boiling mixture for appearance of froth (*phenodgama*) and constantly check the *Kalka* for formation of *varti* (*madhyama pāka lakṣaṇa*).

Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture. Stop heating when the *Kalka* forms a *varti* and the froth subsides. Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.

Pack it in tightly closed glass containers to protect from light and moisture.

Physico-chemical parameters:

<i>Refractive index at 40⁰:</i>	1.471,	Appendix 3.1
<i>Saponification value:</i>	159,	Appendix 3.10
<i>Iodine value:</i>	70,	Appendix 3.11
<i>Acid value:</i>	Not more than 0.40,	Appendix 3.12
<i>Peroxide value:</i>	Not more than 3.5,	Appendix 3.13

6.2.9.4. Mūrçhana of Tila Taila:

(*Bhaiṣajyaratnāvalī*, *Jvarādhikāra*)

Ingredients:

1	Mañjiṣṭhā API	<i>Rubia cordifolia</i>	Rt.	96 g
2	Harītakī API	<i>Terminalia chebula</i>	P.	24 g
3	Bibhītaka API	<i>Terminalia belerica</i>	P.	24 g
4	Āmalakī API	<i>Emblica officinalis</i>	P.	24 g
5	Hrīvera API	<i>Coleus vetiveroides</i>	Rt.	24 g
6	Haridrā API	<i>Curcuma longa</i>	Rz.	24 g
7	Jaladhara (Mustā API)	<i>Cyperus rotundus</i>	Rz.	24 g
8	Lodhra API	<i>Symplocos racemosa</i>	St. Bk.	24 g
9	Sūcīpuṣpa (Ketakī API)	<i>Pandanus odoratissimus</i>	Rt.	24 g
10	Vaṭāṅkura (Nyagrodha API)	<i>Ficus bengalensis</i>	Lt. Bd.	24 g
11	Nalikā (Tvak API)	<i>Cinnamomum tamala</i>	St. Bk.	24 g
12	Taila (Tila taila API)	Sesame oil		1.536 l
13	Jala API	Water		6.144 l

Method of preparation:

Take all ingredients of pharmacopoeial quality.

Wash, clean, dry the ingredients numbered 1 to 11 of the formulation composition powder separately and pass through 180 μm I. S. sieve (*Kalka dravyas*).

Transfer the *Kalka dravyas* to the wet grinder and grind with sufficient quantity of water to prepare homogeneous blend.

Take *Sarṣapa taila* in a stainless steel vessel and heat mildly.

Add increments of *Kalka*. Stir thoroughly while adding water.

Heat for 3 h with constant stirring maintaining the temperature between 50⁰ and 90⁰ during the first hour of heating. Stop heating and allow to stand overnight.

Start the heating next day and observe the boiling mixture for appearance of froth (*phenodgama*) and constantly check the *Kalka* for formation of *varti* (*madhyama pāka lakṣaṇa*).

Expose the *varti* and oil to flame and confirm the absence of crackling sound indicating absence of moisture. Stop heating when the *Kalka* forms a *varti* and the froth subsides. Filter while hot (about 80⁰) through a *muslin cloth* and allow to cool.

Pack it in tightly closed glass containers to protect from light and moisture.

6.3. Yantra Paribhāṣā:

6.3.1. Khalva yantra:

Khalva yantra is an instrument made up of good quality of stone in different sizes and shapes, useful for trituration and levigation processes. It resembles with mortar and pestle.

(*Rasataraṅgiṇī* - 4/53)

6.3.2. Tiryak pātana yantra:

Tiryak pātana yantra is an instrument prepared for distillation of *Pārada* with the delivery tank weld approximately at an angle of 45⁰.

(*Āurveda prakāśa* - 1/79)

6.3.3. Ḍamaru yantra:

Ḍamaru yantra is a contravenes of shape resembling *Ḍamaru* for sublimation prepared by sealing two pots with their mouths one telescoping the other sealing joint securely.

(*Rasataraṅgiṇī* - 4/41)

6.3.4. Dolāyantra:

Dolāyantra consists of a pot half filled with specified liquid with a horizontal rod placed on the rim from which the bundle of material to be treated will be immersed and heated.

(*Rasaratnasamuccaya* - 9/3-4)

APPENDIX - 7

WEIGHTS AND MEASURES

7.1. Metric Equivalents of Classical Weights and Measures

Weights and measures described in Ayurvedic classics and their metric equivalents adopted by the Ayurvedic Pharmacopoeia Committee

The following table of metric equivalents of weights and measures, linear measures and measurement of time used in the Ayurvedic classics have been approved by the Ayurvedic Pharmacopoeia committee in consultation with Indian Standards Institution.

I. WEIGHTS AND MEASURES			II. LINEAR MEASURES		
Classical Unit	Metric Equivalent		Classical Unit	Inches	Metric Equivalent
1 Rattī or Guñjā	= 125 mg		Yavodara	1/8 of 3/4"	0.24 cm
8 Rattī or Guñjā = 1 Māṣa	= 1 g		Aṅgula	3/4"	1.95 cm
12 Māṣas = 1 Karṣa (Tolā)	= 12 g		Vitasti	9"	22.86 cm
2 Karṣas = 1 Śukti	= 24 g		Aratni	10 1/2"	41.91 cm
2 Śukti = 1 Palam	= 48 g		Hasta	18"	45.72 cm
2 Palas = 1 Prasṛti	= 96 g		Nṛpahasta (Rājahasta)	22"	55.88 cm
2 Prasṛtis = 1 Kuḍava	= 192 g		Vyāma	72"	182.88 cm
2 Kuḍavas = 1 Mānikā	= 384 g		III. MEASUREMENT OF TIME		
2 Mānikās = 1 Prastha	= 768 g		Unit		Equivalent (in hours, minutes & in seconds)
4 Prasthas = 1 Āḍhaka	= 3 kg 72 g		2 Kṣaṇas = 1 Lava		
4 Āḍhakas = 1 Droṇa	= 12 kg 288 g		2 Lavas = 1 Nimeṣa		
2 Droṇas = 1 Śūrpa	= 24 kg 576 g		3 Nimeṣas = 1 Kāṣṭhā	= 4.66 seconds	
2 Śūrpas = 1 Droṇī (Vāhī)	= 49 kg 152 g		1 Ghaṭīś = 24 minutes		
4 Droṇīs = 1 Khārī	= 196 kg 608 g		30 Kāṣṭhās = 1 Kalā	= 2 minutes	
100 Palas = 1 Tulā	= 4 kg 800 g			20 seconds	
20 Tulās = 1 Bhāra	= 96 kg		20 Kalās + 3 Kāṣṭhās = 1 Muhūrta	= 48 minutes	
In case of liquids, the metric equivalents would be the corresponding litre and milliliter.			30 Muhūrtas = 1 Ahorātra	= 24 hours	
			15 Ahorātras = 1 Pakṣa	= 15 days	
			2 Pakṣas = 1 Māsa	= 30 days/1 month	
			2 Māsas = 1 Ṛtu	= 60 days/ Two Months	
			3 Ṛtus = 1 Ayana	= 6 Months	
			2 Ayanas = 1 Saṃvatsara	= 12 months/1 Year	
			5 Saṃvatsara = 1 Yuga	= 5 Years	
			1 Ahorātra of Devas	= 1 Year	
			1 Ahorātra of Pitaras	= 1 Month	

7.2. Metric System

Measure of Mass (Weights)

- 1 Kilogram (Kg) – is the mass of the International Prototype Kilogram.
- 1 Gramme (g) – the 1000th part of 1 Kilogram
- 1 Milligram (mg) – the 1000th part of 1 gramme
- 1 Microgram (μ g) – the 1000th part of 1 milligram

Measures of capacity (Volumes)

1 Litre (l) is the volume occupied at its temperature of maximum density by a quantity of water having a mass of 1 Kilogram.

1 Millilitre (ml) is the 1000th part of 1 litre.

The accepted relation between the litre and the cubic centimetre is 1 litre – 1000.027 cubic centimeters.

Relation of capacity of Weight (Metric)

One litre of water at 20^o weighs 997.18 grams when weighed in air of density 0.0012 gram per millilitre against brass weights of density 84 grams per millilitre.

Measures of Length

- 1 Metre (m) is the length of the International Prototype Metre at 0.
- 1 Centimetre (cm) – the 100th part of 1 metre.
- 1 Millimetre (mm) – the 1000th part of 1 metre.
- 1 Micron (μ) – the 1000th part of 1 millimetre
- 1 Millimicron (m μ) – the 1000th part of micron.

APPENDIX - 8

CLASSICAL AYURVEDIC REFERENCES

2 : 3 JAṬĀMĀMSYĀRKA (जटामांस्यर्क) (AFI, Part-I)

(Arkaprakāśa, Śataka 4;: 22 1/2)

सुगन्धवीरणं बाला जटामांसी मुराघनः ।
सटीकचूर्णकाङ्गी सुगन्धोऽयं गुणोत्तमः ॥२२॥
विधिनिष्कासितो योऽर्को रुच्यः पाचनदीपनः ।
(अर्कप्रकाश, शतक ४; २२१/२)

2 : 2 YAVĀNYARKA (यवान्यर्क) (AFI, Part-II)

(Arkaprakāśa, śataka : 3:7)

यवान्याः पाचनो रुच्यो दीपनस्त्रिकशूलहृत् ॥
अजमोदोद्भवो वातकफहा बस्तिशोधनः ॥७॥
(अर्कप्रकाश, शतक ३; ७)

3 : 31 HARIDRĀ KHANḌA (हरिद्रा खण्ड) (AFI, Part-I)

(Bhīṣajyaratnāvalī, Śītapittodardakoṭhādhikāra : 12-16)

हरिद्रायाः पलान्यष्टौ षट्पलं हविषस्तथा ।
क्षीराढकेन संयुक्तं खण्डस्यार्द्धशतं तथा ॥१२॥
पचेन्मृद्वग्निना वैद्यो भाजने मृण्मये दृढे ।
त्रिकटुश्च त्रिजातञ्च कृमिघ्नं त्रिवृता तथा ॥१३॥
त्रिफला केशरं मुस्तं लौहं प्रति पलं पलम् ।
सञ्चूर्ण्य प्रक्षिपेत्तत्र तोलकार्द्धन्तु भक्षयेत् ॥१४॥
कण्डूविस्फोटदद्रूणां नाशनं परमौषधम् ।
प्रतप्तकाञ्चनाभासो देहो भवति नान्यथा ॥१५॥
शीतपित्तोदरद्वकोठान् सप्ताहादेव नाशयेत् ।

हरिद्रानामतः खण्डः कण्डूनां परमौषधम् ॥१६॥
(भैषज्यरत्नावली, शीतपित्तोदरकोठाधिकार; १२-१६)

3 : 16 NĀRIKELA KHANḌA (नारिकेल खण्ड) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Śularogādhikāra : 168-169)

कुडवमितमिह स्यान्नारिकेलं सुपिष्टं पलपरिमितसर्पिः पाचितं खण्डतुल्यम् ।
निजपयसि तदेतत् प्रस्थमात्रे विपक्वं गुडवदथ सुशीते शाणभागान् क्षिपेच्च ॥१६८॥
धन्याकपिप्पलिपयोदतुगाद्विजीरान् शाणं त्रिजातमिभकेशरवद्विचूर्ण्य ।
हन्त्यम्लपित्तमरुचिं क्षतमस्रपित्तं शूलं वमिं सकलपौरुषकारि हरि ॥१६९॥
(भैषज्यरत्नावली, शूलरोगाधिकार; १६८-१६९)

7 : 11 CITRAKĀDI CŪRṆA (चित्रकादि चूर्ण) (AFI, Part-I)

(Śārṅgadharasamhitā, Madhyamakhaṇḍa, Adhyāya 6 : 108-110)

चित्रकं नागरं हिङ्गु पिप्पली पिप्पलीजटा ।
चव्याजमोदा मरिचं प्रत्येकं कर्षसम्मितम् ॥१०८॥
स्वर्जिका च यवक्षारः सिन्धुः सौवर्चलं विडम् ।
सामुद्रकं रोमकं च कोलमात्राणि कारयेत् ॥१०९॥
एकीकृत्याखिलं चूर्णं भावयेन्मातुलुङ्गजैः ।
रसैर्दाडिमजैर्वापि शोषयेदातपेन च ॥११०॥
एतच्चूर्णं जयेद् गुल्मं ग्रहणीमामजां रुजम् ।
अग्निं च कुरुते दीप्तं रुचिकृत् कफनाशनम् ॥१११॥
(शाङ्गधरसंहिता, मध्यमखण्ड, अध्याय ६; १०८-११०)

5:3 SAPTĀṅGA GUGGULU (सप्ताङ्ग गुग्गुलु) (AFI, Part-III)

(Cakradatta, Bhagandaracikitsāprakaraṇa; 15)

गुग्गुलुस्त्रिफलाव्योषैः समांशैराज्ययोजितः ।

नाडीदुष्टव्रणशूलभगन्दरविनाशनः ॥
(चक्रदत्त, भगन्दरचिकित्साप्रकरण; १५)

5 : 1 VARĀDI GUGGULU (वरादि गुग्गुलु) (AFI, Part-III)

(Bhaiṣajyaratnāvalī, Upadaṃśarōgacikitsāprakaraṇa; 65)
वरानिम्बार्जुनाश्वत्थखदिरासनवासकैः ।
चूर्णितैर्गुग्गुलुसमैर्वटका अक्षसम्मिताः ।
उपदंशानसृग्दोषान् तथा दुष्टव्रणानपि ॥
(भैषज्यरत्नावली, उपदंशरोगचिकित्साप्रकरण; ६५)

5 : 2 VIDĀŅGĀDI GUGGULU (विडङ्गादि गुग्गुलु) (AFI, Part-III)

(Yōgaratnākara, Vraṇaśōdhanarōpaṇavidhi; 1)
विडङ्गत्रिफलाव्योषचूर्णं गुग्गुलुना समम् ।
सर्पिषा वटकान् कुर्यात् खादेद्वा हितभोजनः ।
दुष्टव्रणापचीमेहकुष्ठनाडीविशोधनम् ।
(योगरत्नाकर, व्रणशोधनरोपणविधि; १)

6 : 44 SUKUMĀRA GHṚTA (सुकुमार घृत) (AFI, Part-I)

(Sahasrayoga, Ghṛtaprakaraṇa; 4)
पचेत् पुनर्नवतुलां तथा दशपलाः पृथक् ।
दशमूलपयस्याश्वगन्धैरण्डशतावरीः ॥
द्विदर्भशरकाशेक्षुमूलपोटगलान्विताः ।
वहेऽपामष्टभागस्थे तत्र त्रिंशत्पलाद् गुडात् ॥
प्रस्थमेरण्डतैलस्य द्वौ घृतात्पयसस्तथा ।
आवपेद्द्विपलांशाश्च कृष्णातन्मूलसैन्धवम् ॥
यष्टीमधुकमृद्वीकायवानीनागराणि च ।
तत्सिद्धं सुकुमाराख्यं सुकुमारं रसायनम् ॥
वातातपाध्वभाष्यस्त्रीपरिहार्येष्वयन्त्रणम् ।

प्रयोज्यं सुकुमाराणामीश्वराणां सुखात्मनाम् ॥
नृणां स्त्रीवृन्दभर्तृणामलक्ष्मीकलिनाशनम् ।
सर्वकालोपयोगेन कान्तिलावण्यपुष्टिदम् ॥
वर्धमविद्रधिगुल्मार्शोयोनिशूलानिलार्तिषु ।
शोफोदरखुडप्लीहविड्बन्धेषु चोत्तमम् ॥
(सहस्रयोग, घृतप्रकरण; ४)

8 : 1 ANU TAILA (अणु तैल) (AFI, Part-I)

(Aṣṭāṅgahr̥daya, Sūtrasthāna, Adhyāya 20: 37-38)

जीवन्तीजलदेवदारुजलदत्वक्सेव्यगोपीहिमं
दावीत्वङ्मधुकप्लवागुरुवरीपुण्ड्राह्वबिल्वोत्पलम् ।
धावन्यौ सुरभिः स्थिरे कृमिहरं पत्रं त्रुटिं रेणुकां
किञ्जल्कं कमलाह्वयं शतगुणे दिव्येऽम्भसि क्वाथयेत् ॥३७॥
तैलाद्रसं दशगुणं परिशेष्य तेन तैलं पचेच्च सलिलेन दशैव वारान् ।
पाके क्षिपेच्च दशमे सममाजदुग्धं नस्यं महागुणमुशन्त्यणुतैलमेतत् ॥३८॥
(अष्टाङ्गहृदय, सूत्रस्थान, अध्याय २०; ३७-३८)

8 : 1 APĀMĀRGA KṢĀRA TAILA (अपामार्गक्षार तैल) (AFI, Part-II)

(Bhaiṣajyaratnāvalī, Karṇarogādhikāra: 26)

मार्गक्षारजलेन च तत्कृतकल्केन साधितं तैलम् ।
अपहरति कर्णनादं बाधिर्यञ्चापि पूरणतः ॥२६॥
(भैषज्यरत्नावली, कर्णरोगाधिकार : २६)

8 : 2 ARIMEDĀDI TAILA (अरिमेदादि तैल) (AFI, Part-I)

(Aṣṭāṅgahr̥daya, Uttarasthāna, Adhyāya 22: 90 - 96)

खदिरसाराद् द्वे तुले पचेद्वल्कान्तुलां चारिमेदसः ।
 घटचतुष्के पादशेषेऽस्मिन् पूते पुनः क्वथनाद् घने ॥९०॥
 आक्षिकं क्षिपेत्सुसूक्ष्मं रजः सेव्याम्बुपत्तङ्गैरिकम् ।
 चन्दनद्वयरोध्रपुण्ड्राह्वयष्ट्याह्वलाक्षाञ्जनद्वयम् ॥९१॥
 धातकीकट्फलद्विनिशात्रिफलाचतुर्जातजोङ्गकम् ।
 मुस्तमञ्जिष्ठान्यग्रोधप्ररोहमांसीयवासकम् ॥९२॥
 पद्मकैलेयसमङ्गाश्च शीते तस्मिंस्तथा पालिकां पृथक् ।
 जातिपत्रिकां सजातीफलां सहलवङ्गकङ्कोलिकाम् ॥९३॥
 स्फटिकशुभ्रसुरभिकर्पूरकुडवं च तत्रावपेत्ततः ।
 कारयेद् गुटिकाः सदा चैता धार्या मुखे तद् गदापहाः ॥९४॥
 क्वाथौषधव्यत्यययोजनेन तैलं पचेत्कल्पनयाऽनयैव ।
 सर्वास्यरोगोद्धृतये तदाहर्दन्तस्थिरत्वे त्विदमेव मुख्यम् ॥९५॥
 खदिरेणैता गुटिकास्तैलमिदं चारिमेदसा प्रथितम् ।
 अनुशीलयन् प्रतिदिनं स्वस्थोऽपि दृढद्विजो भवति ॥९६॥
 (अष्टाङ्गहृदय, उत्तरस्थान, अध्याय २२; ९०-९६)

8 : 3 ASANABILVĀDI TAILA (असनबिल्वादि तैल) (AFI, Part-I)

(Sahasrayoga, Tailaparakaraṇa: 45)

असनबिल्वबलामृतपाचिते मधुकनागरकत्रिफलान्विते ।
 पयसि तैलमिदं पयसा पचेन्नयनकर्णशिरोहितमुत्तमम् ॥
 (सहस्रयोग, तैलप्रकरण, ४५)

8 : 33 BALĀ TAILA (बला तैल) (AFI, Part-I)

(Aṣṭāṅgahṛdaya, Cikitsāsthāna, Adhyāya 21 : 72-80)

बलाशतं छिन्नरुहापादं रास्नाऽष्टभागिकम् ॥७२॥
 जलाढकशते पक्त्वा शतभागस्थिते रसे ।
 दधिमस्त्विक्षुनिर्यासशुक्तैस्तैलाढकं समैः ॥७३॥
 पचेत्साजपयोऽर्धाशैः कल्कैरेभिः पलोन्मितैः ।
 शठीसरलदार्वेलामञ्जिष्ठागुरुचन्दनैः ॥७४॥

पद्मकातिबलामुस्ताशूर्पपर्णीहरेणुभिः ।
 यष्ट्याह्वसुरसव्याघ्रनखर्षभकजीवकैः ॥७५॥
 पलाशरसकस्तूरीनीलिकाजातिकोशकैः ।
 स्पृक्काकुङ्कुमशैलेयजातिकाकट्फलाम्बुभिः ॥७६॥
 त्वक्कुन्दरुककपूरतुरुष्कश्रीनिवासकैः ।
 लवङ्गनखकङ्कोलकुष्ठमांसीप्रियङ्गुभिः ॥७७॥
 स्थौणेयतगरध्यामवचामदनकप्लवैः ।
 सनागकेसरैः सिद्धे दद्याच्चाऽत्रावतारिते ॥७८॥
 पत्रकल्कं ततः पूतं विधिना तत्प्रयोजितम् ।
 कासश्वासज्वरच्छर्दिमूर्च्छागुल्मक्षतक्षयान् ॥७९॥
 प्लीहशोषमपस्मारमलक्ष्मीं च प्रणाशयेत् ।
 बलातैलमिदं श्रेष्ठं वातव्याधिविनाशनम् ॥८०॥
 (अष्टाङ्गहृदय, चिकित्सास्थान, अध्याय २१; ७२-८०)

8 : 37 BALĀHATHĀDI TAILA (बलाहठादि तैल) (AFI, Part-I)

(Sahasrayoga, Tailaparakaraṇa : 54)

बलाहठामृतामुद्गमाषक्वाथे तिलोद्भवम् ।
 पक्वं शिरोरुजं हन्ति चन्दनामययष्टिभिः ॥
 (सहस्रयोग, तैलप्रकरण; ५४)

8 : 42 BHRṅGARĀJA TAILA (भृङ्गराज तैल) (AFI, Part-I)

(Bhaiṣajyaranāvalī, Kṣudrarogādhikāra: 91-96)

अनूपदेशसम्भूतं गृहीत्वा मार्कवं शुभम् ।
 सुधौतं जर्जरीकृत्य स्वरसं तस्य चाहरेत् ॥९१॥
 चतुर्गुणेन तेनैव तैलप्रस्थं विपाचयेत् ।
 क्षीरपिष्टैरिमैर्द्रव्यैः संयोज्य मतिमान् भिषक् ॥९२॥
 मञ्जिष्ठा पद्मकं लोभ्रं चन्दनं गैरिकं बला ।
 रजन्यौ केशरञ्चैव प्रियङ्गु मधुयष्टिका ॥९३॥
 प्रपौण्डरीकं गोपी च पलिकान्यत्र दापयेत् ।

सम्यक् पक्वं ततो नीत्वा शुभे भाण्डे निधापयेत् ॥१४॥
 केशपाते शिरोदुष्टे मन्यास्तम्भे गलग्रहे ।
 शिरःकर्णाक्षिरोगेषु नस्येऽभ्यङ्गे च योजयेत् ॥१५॥
 कुञ्चिताग्रानतिस्निग्धान् कचान् कुर्याद् बहूंस्तथा ।
 खालित्यमिन्द्रलुप्तञ्च तैलमेतद् व्यपोहति ॥१६॥
 (भैषज्यरत्नावली, क्षुद्ररोगाधिकार; ९१-९६)

8 : 40 BRĀHAT SAINDHAVĀDYA TAILA (बृहत् सैन्धवाद्य तैल) (AFL, Part-I)

(Bhaiṣajyaratnāvalī, Amavātādhikāra : 157-162.)

सैन्धवं श्रेयसी रास्ना शतपुष्पा यमानिका ।
 सर्जिका मरिचं कुष्ठं शुण्ठी सौवर्चलं विडम् ॥१५७॥
 वचाजमोदा मधुकं जीरकं पौष्करं कणा ।
 एतान्यर्द्धपलांशानि श्लक्ष्णपिष्टानि कारयेत् ॥१५८॥
 प्रस्थमेरण्डतैलस्य प्रस्थाम्बु शतपुष्पजम् ।
 काञ्जिकं द्विगुणं दत्त्वा तथा मस्तुः शनैः पचेत् ॥१५९॥
 सिद्धमेतत्प्रयोक्तव्यमामवातहरं परम् ।
 पानाभ्यञ्जनवस्तौ च कुरुतेऽग्निबलं शुभम् ॥१६०॥
 वातार्तरक्षणे शस्तं कटीजानूरुसन्धिजे ।
 शूले हृत्पार्श्वपृष्ठेषु कृच्छ्राशमरिनिपीडिते ॥१६१॥
 बाह्यायामार्दितानाहे अन्त्रवृद्धिनिपीडिते ।
 अन्यांश्चानिलजान् रोगान् नाशयत्याशु देहिनाम् ॥१६२॥
 (भैषज्यरत्नावली, आमवाताधिकार; १५७-१६२)

8 : 16 CITRAKĀDI TAILA (चित्रकादि तैल) (AFL, Part-I)

(Suśrutasaṃhitā, Bhagandara Cikitsā : 50-50 1/2)

चित्रकार्कौ त्रिवृत्पाठे मलपूं हयमारकम् ।
 सुधां वचां लाङ्गलर्की सप्तपर्णं सुवर्चिकाम् ॥५०॥
 ज्योतिष्मतीं च सम्भृत्य तैलं धीरो विपाचयेत् ।
 (सुश्रुतसंहिता, चिकित्सास्थान, अध्याय ८; ५०-५०१/२)

8 : 18 HINGVĀDI TAILA (हिङ्गवादि तैल) (AFI, Part-II)

(Cakradatta, Karṇarogacikitsā: 16)

हिङ्गुतुम्बुरुशुण्ठीभिः साध्यं तैलन्तु सार्षपम् ।
कर्णशूले प्रधानन्तु पूरणं हितमुच्यते ॥१६॥
(चक्रदत्त कर्णरोगचिकित्सा, १६)

8 : 18 JYOTIṢMATĪ TAILA (ज्योतिष्मती तैल) (AFI, Part-I)

(Yogarātnākara, Kuṣṭhacikitsā: Page 696)

मयूरकक्षारजले सप्तकृत्वः परिशृतम् ।
सिद्धं ज्योतिष्मतीतैलमभ्यङ्गाच्छिवत्रनाशनम् ॥
(योगरत्नाकर, कुष्ठशिवत्रचिकित्सा; ६९६)

8 : 4 KANAKA TAILA (कनक तैल) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Kṣudrarogādhikāra: 59 - 60)

मधुकस्य कषायेण तैलस्य कुडवं पचेत् ।
कल्कैः प्रियङ्गुमज्जिष्ठाचन्दनोत्पलकेशरैः ॥५९॥
कनकं नाम तत्तैलं मुखकान्तिकरं परम् ।
अभीरुनीलिकाव्यङ्गशोधनं परमार्चितम् ॥६०॥
(भैषज्यरत्नावली, क्षुद्ररोगाधिकार; ५९-६०)

8 : 45 MAHĀ NĀRĀYAṆA TAILA (महानारायण तैल) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Vatavyādhyadhikāra : 151-162.)

बिल्वाश्वगन्धाबृहतीश्वदंष्ट्राशयोनाकवाट्यालकपारिभद्रम् ।
क्षुद्राकठिल्लाग्निबलाग्निमन्थं मूलानि चैषां सरणीयुतानाम् ॥१५१॥
मूलं विदध्यादथ पाटलीनां प्रस्थं सपादं विधिनोद्धृतानाम् ।
द्रोणैरपामष्टभिरेव पक्त्वा पादावशेषेण रसेन तेन ॥१५२॥

तैलाढकाभ्यां सममेव दुग्धमाजं निदध्यादथवापि गव्यम् ।
 एकत्र सम्यग्विपचेत्सुबुद्धिर्दद्याद्रसञ्चैव शतावरीणाम् ॥१५३॥
 तैलेन तुल्यं पुनरेव तत्र रास्नाश्वगन्धामिषिदारुकुष्ठम् ।
 पर्णीचतुष्कागरुकेशराणि सिन्धूत्थमांसीरजनीद्वयञ्च ॥१५४॥
 शैलेयकं चन्दनपुष्कराणि एलास्रयष्टीतगराब्दपत्रम् ।
 भृङ्गाष्टवर्गाम्बुवचापलाशं स्थौणेयवृश्चीरकचोरकाख्यम् ॥१५५॥
 एतैः समस्तैर्द्विपलप्रमाणैरालोड्य सर्वं विधिना विपक्वम् ।
 कर्पूरकाशमीरमृगाण्डजानां चूर्णीकृतानां त्रिपलप्रमाणम् ॥१५६॥
 प्रस्वेददौर्गन्ध्यनिवारणाय दद्यात् सुगन्धाय वदन्ति केचित् ।
 नारायणं नाम महच्च तैलं सर्वप्रकारैर्विधिवत्प्रयोज्यम् ॥१५७॥
 आश्वेव पुंसां पवनार्दितानामेकाङ्गहीनार्दितवेपनानाम् ।
 ये पङ्गवः पीठविसर्पिणश्च बाधिर्यशुक्रक्षयपीडिताश्च ॥१५८॥
 मन्याहनुस्तम्भशिरोरुजार्ता मुक्तामयास्ते बलवर्णयुक्ताः ।
 संसेव्य तैलं सहसा भवन्ति वन्ध्या च नारी लभते च पुत्रम् ॥१५९॥
 वीरोपमं सर्वगुणोपपन्नं सुमेधसं श्रीविनयान्वितञ्च ।
 शाखाश्रिते कोष्ठगते च वाते वृद्धौ विधेयं पवनार्दितानाम् ॥१६०॥
 जिह्वानिले दन्तगते च शूले उन्मादकौब्ज्यज्वरकर्षितानाम् ।
 प्राप्नोति लक्ष्मीं प्रमदाप्रियत्वं वपुःप्रकर्षं विजयञ्च नित्यम् ॥१६१॥
 तैलोपसेवी जरयाभिमुक्तो जीवेच्चिराच्चापि भवेद् युवेव ।
 देवासुरे युद्धपरे समीक्ष्य स्नाय्वस्थिभग्नानसुरैः सुरांश्च ।
 नारायणेनापि सुबृंहणार्थं स्वनामतैलं विहितञ्च तेषाम् ॥१६२॥
 (भैषज्यरत्नावली, वातव्याध्यधिकार; १५१-१६२)

8 : 24 NĀLPĀMARĀDI TAILA (नाल्पामरादि तैल) (AFI, Part-I)

(Sahasrayoga, Tailaparakaraṇa, 26)

नाल्पामरं त्रिफलचन्दनसेव्यकुष्ठ
 चोवल्लि चोरमगिलेन्निव कल्कमाविक ।
 पैमञ्जल् पर्पटरसे परिपक्वमेण्ण
 तेविकल् केटुं चोरिचिरंगु विसर्पकुष्ठं ॥
 (सहस्रयोग, तैलप्रकरण; २६)

8 : 26 NĪLĪBHRŅGĀDI TAILA (नीलीभृङ्गादि तैल) (AFI, Part-I)

(Sahasrayoga; Tailaparakaraṇa: 38)

नीलीभृङ्गरसशतक्रतुलताधात्रीफलानां रसे
क्षीरैराजकनालिकेरमहिषीधेनूद्भवैस्साधितम् ।
तैलं तत्पयसैव पिष्टलुलितैर्यष्ट्याह्वगुञ्जाञ्जनैः
केशान् सञ्जनयेत्तलेऽपि करयोरास्तामकेशं शिरः ॥
(सहस्रयोग, तैलप्रकरण; ३८)

8 : 30 PRABHAÑJANA VIMARDANA TAILA(प्रभञ्जनविमर्दन तैल) (AFI, Part-I)

(Sahasrayoga, Tailaparakaraṇa : 5)

बलाशतावरीशिग्रुवरुणार्ककरञ्जकाः ।
एरण्डं चापि कोरण्टो वाजिगन्धा प्रसारिणी ॥
पञ्चमूलं वरिष्ठं च तैस्सर्वैः क्वथिते जले ।
पादावशेषिते दद्यात्तैलस्यार्द्धाढकं भिषक् ॥
क्षीरं तद्विगुणं दद्यात्तत्समे दधिकाञ्जिके ।
तगरामरकाष्ठैलाशुण्ठीसर्षपचोरकाः ॥
शताह्वाकुष्ठसिन्धूत्थरास्नाकालानुसारिकाः ।
वचा सचित्रकं मांसी सरलं कटुरोहिणी ॥
प्रत्येकं कार्षिकोन्मानं प्रतीवापाय योजयेत् ।
सर्वैरेभिस्तु मृद्वग्निसिद्धं तिलजमादरात् ॥
पानाभ्यञ्जननस्येषु विधेयं वस्तिकर्मणि ।
अशीतिं वातजान् रोगान्हन्यादाशु सुदारुणान् ॥
वातगुल्मार्दितं वृद्धिमान्त्रजं वातविद्रधिम् ।
मूढगर्भं तथा शूलान्विविधांश्च विनाशयेत् ।
एतत्तैलं महावीर्यमात्रेयप्रमुखैः पुरा ।
निर्मितं नामतश्चापि प्रभञ्जनविमर्दनम् ॥
(सहस्रयोग, तैलप्रकरण; ५)

8 : 32 PRASĀRIṆĪ TAILA (प्रसारिणी तैल) (AFI, Part-I)

(Śārṅgadharaśaṃhitā, Madhyamakhaṇḍa, Adhyāya 9: 119-123 1/2.)

प्रसारणीपलशतं जलद्रोणेन पाचयेत् ।
पादशिष्टः शृतो ग्राह्यस्तैलं दधि च तत्समम् ॥११९॥
काञ्जिकं च समं तैलात् क्षीरं तैलाच्चतुर्गुणम् ।
तैलात् तथाष्टमांशेन सर्वकल्कानि योजयेत् ॥१२०॥
मधुकं पिप्पलीमूलं चित्रकः सैन्धवं वचा ।
प्रसारिणी देवदारु रास्ना च गजपिप्पली ॥१२१॥
भल्लातः शतपुष्पा च मांसी चैर्भिर्विपाचयेत् ।
एतत् तैलवरं पक्वं वातश्लेष्मामयाञ्जयेत् ॥१२२॥
कौब्ज्यं पङ्गुत्वखञ्जत्वे गृध्रसीमर्दितं तथा ।
हनुपृष्ठशिरोग्रीवाकटिस्तम्भान् विनाशयेत् ॥१२३॥
अन्यांश्च विषमान् वातान् सर्वानाशु व्यपोहति ।
(शाङ्गधरसंहिता, मध्यमखण्ड, अध्याय ९; ११९-१२३ १/२)

8 : 20 TUVARAKA TAILA (तुवरक तैल) (AFI, Part-I)

(Suśrutasaṃhitā, Cikitsāsthāna, Adhyāya 13: 20-23, 29)

वृक्षास्तुवरका ये स्युः पश्चिमार्णवभूमिषु ॥२०॥
वीचीतरङ्गविक्षेपमारुतोद्धूतपल्लवाः ।
तेषां फलानि गृह्णीयात् सुपक्वान्यम्बुदागमे ॥२१॥
मज्जां तेभ्योऽपि संहृत्य शोषयित्वा विचूर्ण्य च ।
तिलवत् पीडयेद् द्रोण्यां स्रावयेद्वा कुसुम्भवत् ॥२२॥
तत्तैलं संहृतं भूयः पचेदातोयसंक्षयात् ।
अवतार्य करीषे च पक्षमात्रं निधापयेत् ॥२३॥
तदेव खदिरक्वाथे त्रिगुणे साधु साधितम् ॥२९॥
(सुश्रुतसंहिता, चिकित्सास्थान, अध्याय १३; २०-२३, २९)

8 : 47 YAṢṬIMADHUKA TAILA (यष्टीमधुक तैल) (AFI, Part-I)

(Śārṅgadharaśaṃhitā, Madhyamakhaṇḍa, Adhyāya 9: 155 1/2)

यष्टीमधुकक्षीराभ्यां नवधात्रीफलैः शृतम् ॥१५५॥
तैलं नस्यकृतं कुर्यात् केशाञ्जमश्रूणि सङ्घशः।
(शाङ्गधरसंहिता, मध्यमखण्ड, अध्याय ९; १५५ १/२)

10 : 2 ARKA VAṬĪ (अर्क वटी) (AFI, Part-II)

(Siddhabheṣajamaṇiṃālā; Agnimāndyādicikitsā; 254)

सौवर्चलं सादरमर्कपुष्पं मरिचमेकत्र समं विमर्द्य।
गुञ्जाप्रमाणा गुटिका विधेयाः कर्षन्ति काश्यं क्रमशः कृशानोः॥ २५४॥

12 : 11 CITRAKĀDI GUTĪKĀ (चित्रकादि गुटिका) (AFI, Part-I)

(Carakasāṃhitā, Cikitsāsthāna, Adhyāya 15: 96-97)

चित्रकं पिप्पलीमूलं द्वौ क्षारौ लवणानि च ।
व्योषं हिङ्गवजमोदां च चव्यं चैकत्र चूर्णयेत् ॥९६॥
गुटिका मातुलुङ्गस्य दाडिमस्य रसेन वा ।
कृता विपाचयत्यामं दीपयत्याशु चानलम् ॥९७॥
(चरकसंहिता, चिकित्सास्थान, अध्याय १५; ९६-९७)

12 : 3 ELĀDI GUTĪKĀ (एलादि गुटिका) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Raktapittādhikāra: 32-35)

एलापत्रत्वचोऽर्द्धाक्षाः पिप्पल्यर्द्धपलं तथा ।
सितामधुकखर्जूरमृद्धीकाश्च पलोन्मिताः ॥३२॥
सञ्चूर्ण्य मधुना युक्ता गुटिकाः कारयेद्भिषक् ।
तोलकाद्धां ततश्चैकां भक्षयेन्ना दिने दिने ॥३३॥
श्वासं कासं ज्वरं हिक्कां छर्दिं मूर्च्छां मदं भ्रमम् ।
रक्तनिष्ठीवनं तृष्णां पार्श्वशूलमरोचकम् ॥३४॥

शोषप्लीहाढ्यवातांश्च स्वरभेदं क्षतक्षयम् ।
गुटिका तर्पणी वृष्या रक्तपित्तं विनाशयेत् ॥३५॥
(भैषज्यरत्नावली, रक्तपित्ताधिकार; ३२-३५)

12 : 27 LAŚUNĀDI VAṬĪ (लशुनादि वटी) (AFI, Part-I)

(Vaidyajīvanam, Kṣayarogādīcikitsā: 13)

लशुनजीरकसैन्धवगन्धकत्रिकटुरामठचूर्णमिदं समम् ।
सपदि निम्बुरसेन विषूचिकां हरति भो रतिभोगविचक्षणे ॥१३॥
(वैद्यजीवनम्, क्षयरोगादिचिकित्सा; १३)

12 : 26 LAVANĠĀDI VAṬĪ (लवङ्गादि वटी) (AFI, Part-I)

(Vaidyajīvanam, Kāsaśvāsacikitsā: 7)

तुल्या लवङ्गमरिचाक्षफलत्वचः स्युः ।
सर्वैः समो निगदितः खदिरस्य सारः ॥
बब्बूलवृक्षजकषाययुतञ्च चूर्णम् ।
कासान्निहन्ति गुटिका घटिकाऽष्टकान्ते ॥७॥
(वैद्यजीवनम्, कासश्वासचिकित्सा, ७)

12 : 17 PLĪHĀRI VAṬIKĀ (प्लीहारि वटिका) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Plīhāyakṛdrogādhikāra: 59)

सहासाराभ्रकासीसलशुनानि समानि च ।
द्रोणपुष्पीरसेनैव मर्दयेत्प्रहरत्रयम् ॥५९॥
वल्लद्वयं प्रदातव्यं प्रदोषे सलिलं ह्यनु ।
प्लीहानं यकृतं गुल्ममग्निमान्द्यं सशोथकम् ॥६०॥
कासं श्वासं तृषां कम्पं दाहं शीतं वमिं भ्रमम् ।
प्लीहारिवटिका ह्येषा नाशयेन्नात्र संशयः ॥६१॥
(भैषज्यरत्नावली, प्लीहयकृद्रोगाधिकार; ५९-६१)

12 : 15 PRABHĀKARA VATĪ (प्रभाकर वटी) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Hṛdrogādhikāra : 67)

माक्षिकं लोहमभ्रञ्च तुगाक्षीरी शिलाजतु ।
क्षिप्त्वा खल्लोदरे पश्चाद् भावयेत् पार्थवारिणा ॥६६॥
गुञ्जाद्वयमितां कुर्याद् वटीं छायाविशोषिताम् ।
प्रभाकरवटी सेयं हद्रोगान् निखिलान् जयेत् ॥६७॥
(भैषज्यरत्नावली, हद्रोगाधिकार; ६६-६७)

12 : 25 RAJAḤPRAVARTINĪ VATĪ (रजःप्रवर्तिनी वटी)(AFI, Part-I)

(Bhaiṣajyaratnāvalī, (Strīrogādhikāra) : 233-235)

कन्यासारं च कासीसं रामठं टङ्कणं तथा ।
समादाय समं सर्वं पेषयेत्कन्यकाद्रवैः ॥२३३॥
निर्मापयेद्भिषग्वर्यो रक्तिद्वयमिता वटीः ।
शीलितेयं तु वटिका विनिहन्ति सुदारुणाम् ॥२३४॥
रजोरोधव्यथां कष्टरजःस्रावव्यथां तथा ।
रजःप्रवर्तिनी ह्येषा नीलकण्ठेन भाषिता ॥२३५॥
(भैषज्यरत्नावली, स्त्रीरोगाधिकार; २३३-२३५)

12 : 35 SAÑJĪVANĪ VATĪ (सञ्जीवनी वटी) (AFI, Part-I)

(Śarṅgadharasamhitā, Madhyamakhaṇḍa. Adhyāya 7: 18-21)

विडङ्गं नागरं कृष्णा पथ्यामलबिभीतकम् ॥१८॥
वचा गुडूची भल्लातं सविषं चात्र योजयेत् ।
एतानि समभागानि गोमूत्रेणैव पेषयेत् ॥१९॥
गुञ्जाभा गुटिका कार्या दद्यादारद्रकजै रसैः ।
एकामजीर्णगुल्मेषु द्वे विषूच्यां प्रदापयेत् ॥२०॥
तिस्रश्च सर्पदष्टे तु चतस्रः सान्निपातके ।
वटी सञ्जीवनी नाम्ना सञ्जीवयति मानवम् ॥२१॥
(शाङ्गधरसंहिता, मध्यमखण्ड, अध्याय ७; १८-२१)

12 : 32 ŚAṆKHA VATĪ (शङ्ख वटी) (AFI, Part-I)

(Bhaiṣajyaratnāvalī, Agnimāndyādhikāra : 182-183)

चिञ्चाक्षारपलं पटुब्रजपलं निम्बूरसे कल्कितम् ।
तस्मिन् शङ्खपलं प्रतप्तमसकृत् संस्थाप्य शीर्णावधि ॥१८२॥
हिङ्गुव्योषपलं रसामृतवली निक्षिप्य निष्कांशिकाः ।
बद्ध्वा शङ्खवटी क्षयग्रहणिकारुक्पक्तिशूलादिषु ॥१८३॥
(भैषज्यरत्नावली, अग्निमान्द्यादिरोगाधिकारः; १८२-१८३)

19 : 1 PUNARNAVĀDI MAṆḌŪRA (पुनर्नवा मण्डूर) (AFI, Part-I)

(Carakasamhitā, Cikitsāsthāna, Adhyāya 16: 93-95)

पुनर्नवा त्रिवृद् व्योषं विडङ्गं दारु चित्रकम् ॥९३॥
कुष्ठं हरिद्रे त्रिफला दन्ती चव्यं कलिङ्गकाः ।
पिप्पली पिप्पलीमूलं मुस्तं चेति पलोन्मितम् ॥९४॥
मण्डूरं द्विगुणं चूर्णाद् गोमूत्रे द्व्याढके पचेत् ।
कोलवद् गुटिकाः कृत्वा तत्रेणालोड्य ना पिबेत् ॥९५॥
ताः पाण्डुरोगान् प्लीहानमर्शांसि विषमज्वरम् ।
श्वयथुं ग्रहणीदोषं हन्युः कुष्ठं क्रिमींस्तथा ॥९६॥
(चरकसंहिता, चिकित्सास्थान, अध्याय १६, ९३-९६)

APPENDIX - 9

LIST OF SINGLE DRUGS USED IN FORMULATIONS

9.1 List of Single Drugs of Animal Origin used in Formulations with equivalent English Names:

Name appearing in Monograph (Samskr̥ta Synonym)	Official Name	English Equivalent	A.P.I. Pt. I Reference (Vol. No.)
Dadhi	Godadhi	Curd from cow milk	
Dadhimastu	Godadhi	Whey from curd of cow milk	
Dhenūdbhava	Godugdha	Cow milk	
Ājadugdha	Ajādugdha	Goat milk''''''''	
Ājaka Kṣīra	Ajādugdha	Goat milk	
Ājapaya	Ajādugdha	Goat milk	
Ājya	Goghṛta	Clarified butter from cow milk	
Ghṛta	Goghṛta	Clarified butter from cow milk	
Godugdha	Godugdha	Cow milk	
Gomūtra	Gomūtra	Cow urine	
Haviṣ	Goghṛta	Clarified butter from cow milk	API, Part-I, Vol-VI
K ṣīra	Godughda	Cow milk	
Lākṣā	Lākṣā	Secretion of Lac insect (<i>Lacca laccifera</i>)	
Madhu	Madhu	Honey	API, Part-I, Vol-VI
Mahiṣī Kṣīra	Mahiṣīdugdha	Buffalo milk	
Mastu	Godadhi	Curd whey from cow milk	
Moma	Madhūcchiṣṭa	Hive of s species (Bees wax)	
Payas	Godugdha	Cow milk	

Sarpi	Goghṛta	Clarified butter from cow milk
Śaṅkha	Śaṅkha bhasma	Calcined Conch

9.2 List of Single Drugs of Mineral and Metal Origin used in Formulations with equivalent English Names:

Name appearing in Monograph (Saṃskṛta Synonym)	Official Name	English Equivalent	A.P.I. Pt. I Reference (Vol. No.)
Abhra	Abhraka bhasma	Calcined Abhraka	
Śilājatu	Śilājatu		
Audbhida Lavaṇa	Audbhida Lavaṇa		
Gairika	Gairika	Red ochre	API, Part-I, Vol-VII
Gandhaka	Gandhaka	Sulphur	
Kāsīśa	Kaśīśa	Green vitreol	API, Part-I, Vol-VII
Lauha	Lauha Bhasma	Calcined Lauha	
Maṇḍūra	Maṇḍūra Bhasma		
Mākṣika	Svarṇamākṣika Bhasma	Calcined Chalcopyrite	
Rasa	Pārada	Mercury	
Romaka Lavaṇa	Romaka Lavaṇa		
Sādara	Narasāra	Sal ammoniac	
Sāmudra	Sāmudra Lavaṇa	Sea salt	API, Part-I, Vol-VII
Sāmudra Lavaṇa	Sāmudra Lavaṇa	Sea salt	API, Part-I, Vol-VII
Sāmudraka	Sāmudra Lavaṇa	Sea salt	API, Part-I, Vol-VII
Saindhava	Saindhava Lavaṇa	Rock salt	
Saindhava Lavaṇa	Saindhava Lavaṇa	Rock salt	
Sarjikā	Sarjīkṣāra	Crude alkaline earth	
Sarjīkṣāra	Sarjīkṣāra	Crude alkaline earth	
Sauvīrāñjana	Sauvīrāñjana	<i>Lead sulphide</i>	

Sauvarcala	Sauvarcala Lavaṇa	Black salt	
Sauvarcala Lavaṇa	Sauvarcala Lavaṇa Saindhava	Black salt Rock salt	
Sindhūttha	Lavaṇa Saindhava	Rock salt	
Sindhu	Lavaṇa		
Suvarcikā	Svarjīkṣāra	Crude alkaline earth	
Svarjikā	Svarkīkṣāra	Crude alkaline earth	
Taṅkaṇa	Taṅkaṇa	Borax	API, Part-I, Vol-VII
Valī	Gandhaka	Sulphur	
Viḍa	Viḍa Lavaṇa		
Viḍa Lavaṇa	Viḍa Lavaṇa		

9.3 List of Single Drugs of Plant Used In Formulation, with Latin Nomenclature:

Name appearing in Monograph (Samskrta Synonym)	Official Name	Botanical name/English Equivalent	A.P.I. Pt. I Reference (Vol. No.)
Śālaparṇī	Śālaparṇī	<i>Desmodium gangeticum</i> DC.	Vol. III, Vol. VI
Aśvagandhā	Aśvagandhā	<i>Withania somnifera</i> (L.) Dunal	Vol. I
Aśvattha	Aśvattha	<i>Ficus religiosa</i> L.	Vol. I
Añjana	Rasāñjana	<i>Berberis aristata</i> DC.	Vol. II, VI
Śaṭhī	Śaṭhī	<i>Hedychium spicatum</i> Buch.Ham.	Vol. I
Śaileya	Śaileya	<i>Parmelia perlata</i> (Huds.) Ach.	Vol. III
Śaileyaka	Śaileya	<i>Parmelia perlata</i> (Huds.) Ach.	
Śaramūla	Śara	<i>Saccharum munja</i> Roxb. (= <i>S. bengalense</i> Retz.)	Vol. III
Śarkarā	Śarkarā	Sugar/Sugar candy / Syrup	
Śatāvarī	Śatāvarī	<i>Asparagus racemosus</i> Willd.	Vol. IV
Śatāvarīrasa	Śatāvarī	<i>Asparagus racemosus</i> Willd.	Vol. IV
Śatakratu latā	Kākatiktā	<i>Cardiospermum halicacabum</i> L.	Vol. V

Śatapuṣpā	Śatāhvā	<i>Anethum sowa</i> Kurz	Vol. II
Śatapuṣpajāmbu	Śatāhvā	<i>Anethum sowa</i> Kurz	Vol. II
Abda	Mustā	<i>Cyperus rotundus</i> L.	Vol. III
Agaru	Agaru	<i>Aquilaria agallocha</i> Roxb.	Vol. IV
Agnimantha	Agnimantha	<i>Clerodendrum phlomidis</i> L. [<i>Premna integrifolia</i> L. (Official substitute)]	Vol. III
Aguru	Agaru	<i>Aquilaria agallocha</i> Roxb.	Vol. IV
Śigru	Śigru	<i>Moringa oleifera</i> Lam.	Vol. II, IV
Śigru	Śigru	<i>Moringa oleifera</i> Lam.	Vol. IV
Śigru	Śigru	<i>Moringa oleifera</i> Lam.	Vol. IV
Śigru	Śigru	<i>Moringa oleifera</i> Lam.	Vol. IV
Śilārasa	Turuṣka	<i>Liquidamber orientalis</i> Mill. [<i>Altingia excelsa</i> (Official substitute)]	
Aileya	Elavāluka	<i>Prunus avium</i> L.	Vol. VI
Aileya	Elavāluka	<i>Prunus avium</i> L.	Vol. V
Ajamodā	Ajamodā	<i>Apium leptophyllum</i> (Pers.) F.V.M. ex Benth.	Vol. I
Akṣaphalatvak	Bibhītaka	<i>Terminalia belerica</i> Roxb.	Vol. I
Akil	Agaru	<i>Aquilaria agallocha</i> Roxb.	Vol. IV
Amṛtā	Guḍūcī	<i>Tinospora cordifolia</i> (Willd.) Miers ex Hook.f. & Thoms.	Vol. I
Amṛta	Vatsanābha	<i>Aconitum chasmanthum</i> Stapf	Vol. II
Amarakāṣṭha	Devadāru	<i>Cedrus deodara</i> (Roxb.) Loud.	Vol. IV
Ambu	Hrīvera	<i>Coleus vettiveroides</i> K.C. Jacob	
Apāmārga	Apāmārga	<i>Achyranthes aspera</i> L.	Vol. II, III
Śrīnivāsaka	Sarala	<i>Pinus roxburghii</i> Sargent.	Vol. III
Śreyasī	Gajapippalī	<i>Scindapsus officinalis</i> Schott.	Vol. II
Arimedavalka	Arimeda	<i>Acacia leucophloea</i> Willd.	Vol. II
Arjuna	Arjuna	<i>Terminalia arjuna</i> (Roxb) W. & A.	Vol. II
Arka	Arka	<i>Calotropis procera</i> (Ait.) R. Br.	Vol. I
Arkamūla	Arka	<i>Calotropis procera</i> (Ait.) R. Br.	Vol. III
Arkapuṣpa	Arka	<i>Calotropis procera</i> (Ait.) R. Br.	Vol. I, Vol. III
Asana	Asana	<i>Pterocarpus marsupium</i> Roxb.	Vol. I
Asana	Asana	<i>Pterocarpus marsupium</i> Roxb.	Vol. III
Asra	Mañjiṣṭhā	<i>Rubia cordifolia</i> L.	Vol. III

Atibalā	Atibalā	<i>Abutilon indicum</i> (L.) Sw.	Vol. I
Śuṅṭhī	Śuṅṭhī	<i>Zingiber officinale</i> Rosc.	Vol. II
Śuṅṭhī	Śuṅṭhī	<i>Zingiber officinale</i> Rosc.	Vol. I
Śukta	Kāñjika	Sour gruel	
Śvadamṣṭrā	Gokṣura	<i>Tribulus terrestris</i> L.	Vol. VI
Śvadamṣṭrā	Gokṣura	<i>Tribulus terrestris</i> L.	Vol. I
Śveta candana	Śveta candana	<i>Santalum album</i> L.	Vol. III
Śveta jīraka	Śveta jīraka	<i>Cuminum cyminum</i> L.	Vol. I
Śyonāka	Śyonāka	<i>Oroxylum indicum</i> Vent.	Vol. III
Bṛhatī	Bṛhatī	<i>Solanum indicum</i> Lam.	Vol. II
Babbūlavṛkṣaja	Babbūla	<i>Acacia arabica</i> Willd. (= <i>Acacia nilotica</i> (L.) Willd. ex Del. sp. indica (Benth.)	Vol. I
Balā	Balā	<i>Sida cordifolia</i> L.	
Bhṛṅga	Tvak	<i>Cinnamomum zeylanicum</i> Breyn.	Vol. I, Vol. VI
Bhṛṅgarasa	Bhṛṅgarāja	<i>Eclipta alba</i> (L.) Hassk.	Vol. II
Bhallāta	Bhallātaka	<i>Semecarpus anacardium</i> L.f	Vol. II
Bhallātaka	Bhallātaka	<i>Semecarpus anacardium</i> L.f	Vol. II
Bibhītaka	Bibhītaka	<i>Terminalia bellerica</i> Roxb.	Vol. I
Bilva	Bilva	<i>Aegle marmelos</i> Corr.ex Roxb.	Vol. I
Bilva	Bilva	<i>Aegle marmelos</i> Corr.ex Roxb.	Vol. IV
Bilva	Bilva	<i>Aegle marmelos</i> Corr.ex Roxb.	Vol. III
Brāhmī	Brāhmī	<i>Bacopa monnieri</i> (L.) Penn.	Vol. II
Candana	Rakta candana	<i>Pterocarpus santalinus</i> L.f.	Vol. III
Cavya	Cavya	<i>Piper chaba</i> Hunter	
Chinnaruhā	Guḍūcī	<i>Tinospora cordifolia</i> (Willd.) Miers ex Hook.f. & Thoms.	Vol. I
Ciñcā kṣāra	Ciñcā -kṣāra	<i>Tamarindus indica</i> L.	Vol. IV
Citraka	Citraka	<i>Plumbago zeylanica</i> L.	Vol. I
Coraka	Coraka	<i>Angelica glauca</i> Edgew	Vol. V
Coram	Coram	<i>Kaempferia galanga</i> L.	
Covalli	Mañjiṣṭhā	<i>Rubia cordifolia</i> L.	Vol. III
Dāru	Devadāru	<i>Cedrus deodara</i> (Roxb.) Loud.	Vol. IV
Dāruharidrā	Dāruharidrā	<i>Berberis aristata</i> DC	Vol. II
Dārvī	Dāruharidrā	<i>Berberis aristata</i> DC	Vol. VI
Darbha	Darbha	<i>Imperata cylindrica</i> (L.) Beauv	Vol. V
Devadāru	Devadāru	<i>Cedrus deodara</i> (Roxb.) Loud.	Vol. IV

Dhātakī	Dhātakī	<i>Woodfordia fruticosa</i> Kurz	Vol. I
Dhātrī Phala	Āmalakī	<i>Emblīca officinalis</i> Gaertn.	Vol. I
Dhanyāka	Dhānyaka	<i>Coriandrum sativum</i> L.	Vol. I
Dhyāma	Rohiṣa	<i>Cymbopogon martini</i> (Roxb.) Wats.	Vol. V
Drākṣā	Drākṣā	<i>Vitis vinifera</i> L.	Vol. III
Droṇapuṣpī - Svarasa	Droṇapuṣpī	<i>Leucas cephalotes</i> Spreng.	Vol. II
Eṇṇa	Tila Taila	Sesame oil	
Elā	Sūkṣmailā	<i>Elettaria cardamomum</i> Maton	Vol. I
Āmala	Āmalakī	<i>Phyllanthus emblica</i> L.	
Āmalakī	Āmalakī	<i>Emblīca officinalis</i> Gaertn.	Vol. I
Āmaya	Kuṣṭha	<i>Saussurea lappa</i> C.B. Clarke	Vol. I
Eraṇḍa	Eraṇḍa	<i>Ricinus communis</i> L.	Vol. III
Eraṇḍa	Eraṇḍa	<i>Ricinus communis</i> L.	Vol. I
Eraṇḍa taila	Eraṇḍa taila	<i>Ricinus communis</i> L. oil	Vol. I, Vol. III
Gajppalī	Gajapippalī	<i>Scindapsus officinalis</i> Schott.	Vol. II
Gambhārī	Gambhārī	<i>Gmelina arborea</i> Roxb.	Vol. II
Gambhārī	Gambhārī	<i>Gmelina arborea</i> Roxb.	Vol. I, Vol. III, Vol. IV
Gambhārī	Gambhārī	<i>Gmelina arborea</i> Roxb.	
Gambhārī	Gambhārī	<i>Gmelina arborea</i> Roxb.	
Gandhavirojā	Sarala	<i>Pinus roxburghii</i> Sargent.	Vol. III
Gokṣura	Gokṣura	<i>Tribulus terrestris</i> L.	Vol. VI
Gokṣura	Gokṣura	<i>Tribulus terrestris</i> L.	Vol. I
Gopī	Śveta Sārivā	<i>Hemidesmus indicus</i> R.Br.	Vol. I
Guñjā	Guñjā	<i>Abrus precatorius</i> L.	Vol. I, Vol. II
Guḍūcī	Guḍūcī	<i>Tinospora cordifolia</i> (Willd.) Miers ex. Hook.f. & Thoms.	Vol. I
Guḍa	Guḍa	Jaggery	Vol. VI
Guggulu	Guggulu	<i>Commiphora wightii</i> (Arn.) Bhand.	Vol. I
Gulāba	Gulāba	<i>Rosa damascena</i> Mill.	
Haṭhā	Āmalakī	<i>Emblīca officinalis</i> Gaertn.	Vol. I
Harītakī	Harītakī	<i>Terminalia chebula</i> Retz.	Vol. I
Hareṇu	Reṇukā	<i>Vitex agnus-castus</i> L. [<i>Vitex negundo</i> L. (Official substitute)]	Vol. III
Hareṇu	Reṇukā	<i>Vitex agnus-castus</i> L.	Vol. IV

		[<i>Vitex negundo</i> L. (Official substitute)]	
Hareṇu	Reṇukā	<i>Vitex agnus-castus</i> L.	Vol. V
		[<i>Vitex negundo</i> L. (Official substitute)]	
Haridrā	Haridrā	<i>Curcuma longa</i> L.	Vol. I
Hayamāraka	Karavīra	<i>Nerium indicum</i> Mill.	Vol. I, Vol. III
Hayamāraka	Karavīra	<i>Nerium indicum</i> Mill.	Vol. III
Hiṅgu	Hiṅgu	<i>Ferula foetida</i> Regel.	Vol. I
Hima	Śveta Candana	<i>Santalum album</i> L.	Vol. III
Rṣabhaka	Rṣabhaka	<i>Microstylis wallichii</i> Lindle (= <i>Malaxis acuminata</i> D. Don)	Vol. II, V
		[<i>Pueraria tuberosa</i> (Official substitute)]	
Rṣabhaka	Rṣabhaka	<i>Microstylis wallichii</i> Lindle (= <i>Malaxis acuminata</i> D. Don)	Vol. V
		[<i>Pueraria tuberosa</i> (Official substitute)]	
Ibhakeśara	Nāgakeśara	<i>Mesua ferrea</i> L.	Vol. II
Rḍdhi	Rḍdhi	<i>Habenaria intermedia</i> D. Don	Vol. IV
		[<i>Dioscorea bulbifera</i> (Official substitute)]	
Ikṣu Niryāsa	Ikṣu	<i>Saccharum officinarum</i> L.	Vol. IV
Ikṣumūla	Ikṣu	<i>Saccharum officinarum</i> L.	Vol. II
Jātīpatrikā	Jātīphalā	<i>Myristica fragrans</i> Houtt.	Vol. I
Jātīphala	Jātīphala	<i>Myristica fragrans</i> Houtt.	Vol. I
Jātikā	Jātīphala	<i>Myristica fragrans</i> Houtt.	Vol. I
Jātikōśa	Jātīpatrī	<i>Myristica fragrans</i> Houtt.	Vol. I
Jīraka	Śveta Jīraka	<i>Cuminum cyminum</i> L.	Vol. I
Jīvaka	Jīvaka	<i>Microstylis wallichii</i> Lindle (= <i>Malaxis acuminata</i> D. Don)	Vol. II
		[<i>Pueraria tuberosa</i> (Official substitute)]	
Jīvaka	Jīvaka	<i>Microstylis wallichii</i> Lindle (= <i>Malaxis acuminata</i> D. Don)	Vol. V
		[<i>Pueraria tuberosa</i> (Official substitute)]	
Jīvantī	Jīvantī	<i>Leptadenia reticulata</i> W.& A.	Vol. VI

Jaṭāmamsī	Jaṭāmamsī	<i>Nardostachys jatamansi</i> DC.	Vol. I
Jala	Hrīvera	<i>Coleus vettiveroides</i> K.C. Jacob	
Jalada	Mustā	<i>Cyperus rotundus</i> L.	Vol. III
Joṅgaka	Agaru	<i>Aquilaria agallocha</i> Roxb.	Vol. IV
Jyotiṣmatī	Jyotiṣmatī	<i>Celastrus paniculatus</i> Willd.	Vol. II
Jyotiṣmatī Taila	Jyotiṣmatī Taila	<i>Celastrus paniculatus</i> Willd.	
Kāśamūla	Kāśa	<i>Saccharum spontaneum</i> L.	Vol. III
Kāśmīra	Kuṅkuma	<i>Crocus sativus</i> L.	Vol. IV
Kāñjika	Kāñjika	Sour gruel	
Kākamācī	Kākamācī	<i>Solanum nigrum</i> L.	Vol. II
Kākolī	Kākolī	<i>Lilium polyphyllum</i> D. Don [(<i>Withania somnifera</i> Dunal) (Official substitute)]	Vol. III
Kālānusārikā	Methī	<i>Trigonella foenum-graecum</i> L.	Vol. II
Kṛṣṇā	Pippalī	<i>Piper longum</i> L.	Vol. IV
Kṛṣṇāmūla	Pippalī	<i>Piper longum</i> L.	Vol. II
Kṛṣṇa jīraka	Kṛṣṇa jīraka	<i>Carum carvi</i> L.	Vol. I
Kṛimihara	Viḍaṅga	<i>Embelia ribes</i> Burm.f.	Vol. I
Kṛmighna	Viḍaṅga	<i>Embelia ribes</i> Burm.f.	Vol. I
Kaṅkola	Kaṅkola	<i>Piper cubeba</i> L. f.	Vol. I
Kaṅkolikā	Kaṅkola	<i>Piper cubeba</i> L. f.	
Kaṇā	Pippalī	<i>Piper longum</i> L.	Vol. IV
Kaṇṭakārī	Kaṇṭakārī	<i>Solanum xanthocarpum</i> Schrad. & Wendl. (= <i>S. surratense</i> Burm.f.)	Vol. I
Kṣīrakākolī	Kṣīrakākolī	<i>Fritillaria roylei</i> Hook. [(<i>Withania somnifera</i> Dunal (Official substitute)]	Vol. V
Kaṭhillā	Rakta Punarnavā	<i>Boerhaavia diffusa</i> L.	Vol. I, Vol. III
Kaṭphala	Kaṭphala	<i>Myrica esculenta</i> Buch.- Ham. ex D. Don (= <i>M. nagi</i> Hook. f.)	Vol. III
Kaṭurohiṇī	Kaṭukā	<i>Picrorhiza kurroa</i> Royle ex Benth.	Vol. II
Kamalakiñjalka	Kamala	<i>Nelumbo nucifera</i> Gaertn.	Vol. II, Vol. III
Kanyāsāra	Kanyāsāra	<i>Aloe barbadensis</i> - Mill.	Vol. I
Kanyakā	Kumārī - Svarasa	<i>Aloe barbadensis</i> Mill.	Vol. I
Karañjaka	Karañja	<i>Pongamia pinnata</i> Pierre	Vol. II

Karañjaka	Karañja	<i>Pongamia pinnata</i> Pierre	Vol. I
Karpūra	Karpūra	<i>Cinnamomum camphora</i> – (L.) Nees & Eberm.	Vol. VI
Kṣudrā	Kaṅṭakārī	<i>Solanum surattense</i> Burm.f.	Vol. I
Keśara	Nāgakeśara	<i>Mesua ferrea</i> L.	Vol. II
Kejopuṭī Taila	Kejopuṭī Taila	<i>Melaleuca leucodendron</i>	
Khaṇḍa	Śarkarā	Sugar	Vol. VI
Khaṇḍa	Śarkarā	Sugar candy	Vol. VI
Khadira	Khadira	<i>Acacia catechu</i> Willd.	Vol. I
Khadirasāra	Khadira	<i>Acacia catechu</i> - Willd.	Vol. I
Kharjūra	Kharjūra	<i>Phoenix sylvestris</i> Roxb.	
Koraṇṭa	Sahacara	<i>Barleria prionitis</i> L.	Vol. III
Kuñkuma	Kuñkuma	<i>Crocus sativus</i> L.	Vol. IV
Kuśa	Kuśa	<i>Desmostachya bipinnata</i> Stapf	Vol. III
Kuṣṭha	Kuṣṭha	<i>Saussurea lappa</i> C.B. Clarke	Vol. I
Kunduruka	Kunduru	<i>Boswellia serrata</i> Roxb.	Vol. IV
Lāṅgalakī	Lāṅgalī	<i>Gloriosa superba</i> L.	Vol. III
Laśuna	Rasona	<i>Allium sativum</i> L.	Vol. III
Lavaṅga	Lavaṅga	<i>Syzygium aromaticum</i> (L.) Merril & Perry	Vol. I
Lodhra	Lodhra	<i>Symplocos racemosa</i> Roxb.	Vol. I
Māṣaparnī	Māṣaparnī	<i>Teramnus labialis</i> Spreng.	Vol. III
Māṃsī	Jaṭāmāṃsī	<i>Nardostachys jatamansi</i> DC.	Vol. I
Mārga Kṣāra	Apāmārga Kṣāra	<i>Achyranthes aspera</i> L. - Water soluble ash of Pl.	Vol. II
Mārga Kṣāra	Apāmārga Kṣāra	<i>Achyranthes aspera</i> L. - Water soluble ash of Pl.	Vol. II
Mārkava Svarasa	Bhṛṅgarāja	<i>Eclipta alba</i> (L.) Hassk.	Vol. II
Mātuluṅga	Mātuluṅga	<i>Citrus aurantifolia</i> Swingle	
Mṛdvīkā	Drākṣā	<i>Vitis vinifera</i> L.	Vol. III
Mañjiṣṭhā	Mañjiṣṭhā	<i>Rubia cordifolia</i> L.	Vol. III
Madanaka	Madana	<i>Randia dumetorum</i> Poir.	Vol. I
Madhūka	Madhūka	<i>Madhuca indica</i> (L.) J.F. Gmel	Vol. II
Madhuka	Yaṣṭī	<i>Glycyrrhiza glabra</i> L.	Vol. I
Madhusnuhī	Madhusnuhī	<i>Smilax china</i> L. [<i>Smilax glabra</i> Roxb. (Official substitute)]	Vol. I, Vol. V
Madhuyaṣṭīkā	Yaṣṭī	<i>Glycyrrhiza glabra</i> L.	Vol. I

Mahāmedā	Mahāmedā	<i>Polygonatum cirrhifolium</i> Royle. [<i>Asparagus racemosus</i> (Official substitute)]	Vol. IV
Malapū	Phalgu	<i>Ficus hispida</i> L.f.	Vol. III
Marica	Marica	<i>Piper nigrum</i> L.	Vol. III
Mayūrakakṣāra	Apāmārgakṣāra	<i>Achyranthes aspera</i> L.- Water soluble Ash of Pl.	Vol. II
Mayūrakakṣāra	Apāmārgakṣāra	<i>Achyranthes aspera</i> L.- Water soluble Ash of Pl.	Vol. II
Medā	Medā	<i>Polygonatum verticillatum</i> [<i>Asparagus racemosus</i> Willd. (Official substitute)]	Vol. IV
Miṣi	Miśreyā	<i>Foeniculum vulgare</i> Mill.	Vol. I
Muṇḍītikā	Muṇḍītikā	<i>Sphaeranthus indicus</i> L.	Vol. III
Mudga	Mudga	<i>Phaseolus radiatus</i> L. (= <i>Phaseolus</i> <i>mungo</i> L.)	Vol. III
Mudgaparṇī	Mudgaparṇī	<i>Phaseolus trilobus</i> (L.) Schreb.	Vol. III
Mustā	Mustā	<i>Cyperus rotundus</i> L.	Vol. III
Musta	Mustā	<i>Cyperus rotundus</i> L.	Vol. III
Nāgakeśara	Nāgakeśara	<i>Mesua ferra</i> L.	Vol. II
Nāgakesara	Nāgakeśara	<i>Mesua ferrea</i> L.	Vol. II
Nāgara	Śuṅṭhī	<i>Zingiber officinale</i> Rosc.	Vol. II
Nāgara	Śuṅṭhī	<i>Zingiber officinale</i> Rosc.	Vol. I
Nālikera Kṣīra	Nārikela	<i>Cocos nucifera</i> L. - Milk from End.	Vol. III
Nārikela	Nārikela	<i>Cocos nucifera</i> L.	Vol. III
Nārikela paya	Nārikela	<i>Cocos nucifera</i> L.-	Vol. III
Nīlī	Nīlī	<i>Indigofera tinctoria</i> L.	Vol. II, III
Nīlikā	Nīlī	<i>Indigofera tinctoria</i> L.	Vol. III
Nakha	Vyāghranakha	<i>Capparis sepiaria</i> L.	Vol. V
Nimbū	Nimbū	<i>Citrus limon</i> (L.) Burm.f.	Vol. IV
Nimba	Nimba	<i>Azadirachta indica</i> A. Juss.	Vol. V
Nimba patra	Nimba	<i>Azadirachta indica</i> A. Juss.	Vol. II
Nimbu	Nimbū	<i>Citrus limon</i> (L.) Burm.f.	Vol. IV
Nyagrodha	Nyagrodha	<i>Ficus bengalensis</i> L.	Vol. IV
Nyagrodhapraroha	Nyagrodha	<i>Ficus benghalensis</i> L.	Vol. I
Pāṭalā	Pāṭalā	<i>Stereospermum suaveolens</i> DC.	Vol. III
Pāṭalī	Pāṭalā	<i>Stereospermum suaveolens</i> DC.	Vol. III

Pāṭhā	Pāṭhā	<i>Cissampelos pareira</i> L.	Vol. I
Pāribhadra	Pāribhadra	<i>Erythrina indica</i> Lam.	Vol. II
Pārthavāri	Arjuna - Kvātha	<i>Terminalia arjuna</i> (Roxb) W. & A.	Vol. II
Pr̥ṣniparṇī	Pr̥ṣniparṇī	<i>Uraria picta</i> Desv.	Vol. IV
Padmaka	Padmaka	<i>Prunus cerasoides</i> D. Don	Vol. III
Paimannal	Haridrā - Ārdra	<i>Curcuma longa</i> L.	Vol. I
Palāśa	Palāśa	<i>Butea monosperma</i> (Lam.) Kuntze	Vol. II
Palāśa rasa	Palāśa	<i>Butea monosperma</i> (Lam.) Kuntze	Vol. IV
Palāśa rasa	Palāśa	<i>Butea monosperma</i> (Lam.) Kuntze	Vol. V
Parpaṭa	Parpaṭa	<i>Fumaria vaillantii</i> Loisel (= <i>F. parviflora</i> Lam.) (= <i>Fumaria parviflora</i> Lam.)	Vol. IV
Pathyā	Harītakī	<i>Terminalia chebula</i> Retz.	Vol. I
Patra	Tvak patra	<i>Cinnamomum tamala</i> Nees & Eberm.	Vol. I
Pattaṅga	Pattaṅga	<i>Caesalpinia sappan</i> L.	Vol. IV
Pauṣkara	Puṣkara	<i>Inula racemosa</i> Hook. f.	Vol. IV
Payasyā	Ksīrakākolī	<i>Fritillaria roylei</i> Hook	Vol. V
Payoda	Mustā	<i>Cyperus rotundus</i> L.	Vol. III
Pippalī	Pippalī	<i>Piper longum</i> L.	Vol. IV
Pippalījaṭā	Pippalī	<i>Piper longum</i> L.	Vol. II
Pippalīmūla	Pippalī	<i>Piper longum</i> L.	
Plakṣa	Plakṣa	<i>Ficus lacor</i> Buch.-Ham.	Vol. II
Plakṣa	Plakṣa	<i>Ficus lacor</i> Buch.-Ham.	Vol. IV
Poṭagala	Poṭagala	<i>Typha elephantina</i> Roxb.	Vol. V
Prapaṇḍarīka	Prapaṇḍarīka	<i>Nelumbo nucifera</i> Gaertn.	Vol. II
Prapaṇḍarīka	Prapaṇḍarīka	<i>Nelumbo nucifera</i> Gaertn.	Vol. III
Prasāriṇī	Prasāriṇī	<i>Paederia foetida</i> L.	Vol. II
Prasāriṇī	Prasāriṇī	<i>Paederia foetida</i> L.	Vol. III
Priyaṅgu	Priyaṅgu	<i>Callicarpa marcophylla</i> Vahl.	Vol. II, Vol. IV,
Puṇḍrāhva	Prapaṇḍarīka	<i>Nelumbo nucifera</i> Gaertn.	Vol. II
Puṇḍrāhva	Prapaṇḍarīka	<i>Nelumbo nucifera</i> Gaertn.	Vol. III
Puṣkara	Puṣkara	<i>Inula racemosa</i> Hook. f.	Vol. IV
Pudīnā	Pudīnā	<i>Mentha viridis</i> L.	Vol. V
Punarnava	Rakta	<i>Boerhaavia diffusa</i> L.	Vol. III
	Punarnavā		
Punarnava	Rakta	<i>Boerhaavia diffusa</i> L.	Vol. I

	Punarnavā		
Rāla	Śāla	<i>Shorea robusta</i> Gaertn f.	Vol. VI
Rāmaṭha	Hiṅgu - Śuddha	<i>Ferula foetida</i> Regel.	Vol. I
Rāsnā	Rāsnā	<i>Pluchea lanceolata</i> C.B. Clarke [<i>Alpinia galanga</i> Willd. (Official substitute)]	Vol. V
Rakta chandana	Rakta chandana	<i>Pterocarpus santalinus</i> L.f.	Vol. III
Rasāñjana	Rasāñjana	<i>Berberis aristata</i> DC.	Vol. II
Reṇukā	Reṇukā	<i>Vitex negundo</i> L.	Vol. V
Reṇukā	Reṇukā	<i>Vitex negundo</i> L.	Vol. III
Reṇukā	Reṇukā	<i>Vitex negundo</i> L.	Vol. IV
Rodhra	Lodhra	<i>Symplocos racemosa</i> Roxb.	Vol. I
Sārṣapa Taila	Sarṣapa Taila	Mustard oil	Vol. VI
Sārivā	Śveta sārivā	<i>Hemidesmus indicus</i> R.Br.	Vol. I
Sūkṣmailā	Sūkṣmailā	<i>Elettaria cardamomum</i> Maton	Vol. I
Sahāsāra	Kumārī	<i>Aloe barbadensis</i> Mill.	Vol. I
Samaṅgā	Lajjālu	<i>Mimosa pudica</i> L.	Vol. II
Sambhālu patra	Nirguṇḍī	<i>Vitex negundo</i> L.	Vol. III
Sambhālu patra	Nirguṇḍī	<i>Vitex negundo</i> L.	Vol. IV
Saptaparṇa	Saptaparṇa	<i>Alstonia scholaris</i> R.Br.	Vol. I
Saraṇī	Prasāriṇī	<i>Paederia foetida</i> L.	Vol. II
Sarṣapa	Sarṣapa	<i>Brassica campestris</i> L.	Vol. III
Sarala	Sarala	<i>Pinus roxburghii</i> Sargent.	Vol. III
Sevya	Uśīra	<i>Vetiveria zizanioides</i> (L.) Nash	Vol. III
Sitā	Śarkarā	Sugar candy	Vol. VI
Sprkkā	Sprkkā	<i>Anisomeles malabarica</i> R.Br.	Vol. VI
Sthauṇeya	Sthauṇeya	<i>Taxus baccata</i> L.	Vol. III
Sudhā	Snuhī	<i>Euphorbia neriifolia</i> L.	Vol. I

Surabhi	Rāsnā	<i>Pluchea lanceolata</i> C.B. Clarke [<i>Alpinia officinarum</i> Hance (Official substitute)]	Vol. III
Surasa	Tulasī	<i>Ocimum sanctum</i> L.	Vol. II, IV
Surasa	Tulasī	<i>Ocimum sanctum</i> L.	Vol. IV
Tārapīna Taila	Tārapīna Taila	<i>Pinus roxburghii</i>	Vol. III
Tagara	Tagara	<i>Valeriana wallichii</i> DC.	Vol. I
Taila	Tila Taila	Sesame oil	Vol. VI
Tailaparna Taila	Tailaparna Taila	<i>Eucalyptus globulus</i> Labill	Vol. VI
Tila Taila	Tila Taila	Sesame oil	Vol. VI
Tilodbhava	Tila Taila	Sesame oil	Vol. VI
Trivṛt	Trivṛt	<i>Operculina turpethum</i> (L.) Silva mansa (= <i>Ipomoea turpethum</i>) R. Br.	Vol. III
Trivṛtā	Trivṛt	<i>Operculina turpethum</i> (L.) Silva Manso	Vol. III
Truṭi	Sūkṣmailā	<i>Elettaria cardamomum</i> Maton	Vol. I
Tugā	Vaṃśalocana	Bamboo manna	
Tugākṣīrī	Vaṃśalocana	Bamboo manna	
Tumburu	Tejovātī	<i>Zanthoxylum armatum</i> DC.	Vol. IV
Tumburu	Tejovātī	<i>Zanthoxylum armatum</i> DC.	Vol. II
Turuṣka	Turuṣka	<i>Liquidambar orientalis</i> Mill. [<i>Altingia excelsa</i> (Official substitute)]	
Tuvaraka	Tuvaraka	<i>Hydnocarpus laurifolia</i> (Dennst.) Steum.	
Tuvaraka taila	Tuvaraka taila	<i>Hydnocarpus laurifolia</i> (Dennst.) Steum.	
Tvak	Tvak	<i>Cinnamomum zeylanicum</i> Breyn.	Vol. I, Vol. VI
Tvakpatra	Tvakpatra	<i>Cinnamomum tamala</i> Nees & Eberm.	Vol. I
Udumbara	Udumbara	<i>Ficus racemosa</i> L.	Vol. I
Utpala	Utpala	<i>Nymphaea stellata</i> Willd.	Vol. III
Vāṭyālaka	Balā	<i>Sida cordifolia</i> L.	
Vājigandhā	Aśvagandhā	<i>Withania somnifera</i> (L.) Dunal	Vol. I
Vāsaka	Vāsā	<i>Adhatoda zeylanica</i> Medic.	Vol. IV

Vṛścīraka	Śveta Punarnavā	<i>Boerhaavia verticillata</i> Poiret	Vol. V
Vṛddhi	Vṛddhi	<i>Habenaria intermedia</i> D. Don [<i>Dioscorea bulbifera</i> (Official substitute)]	Vol. IV
Vacā	Vacā	<i>Acorus calamus</i> L.	Vol. II
Vari	Śatāvari	<i>Asparagus racemosus</i> <i>Asparagus</i>	Vol. IV
Varuṇa	Varuṇa	<i>Crataeva nurvala</i> Buch.-Ham.	Vol. I
Viḍaṅga	Viḍaṅga	<i>Embelia ribes</i> Burm.f.	Vol. I
Viṣa	Vatsanābha - Śuddha	<i>Aconitum chasmanthum</i> Stapf	Vol. II
Vyāghranakha	Vyāghranakha	<i>Capparis sepiaria</i> L.	Vol. V
Yaṣṭī	Yaṣṭī	<i>Glycyrrhiza glabra</i> L.	Vol. I
Yaṣṭīmadhuka	Yaṣṭī	<i>Glycyrrhiza glabra</i> L.	Vol. I
Yaṣṭi	Yaṣṭī	<i>Glycyrrhiza glabra</i> L.	Vol. I
Yaṣṭyāhva	Yaṣṭī	<i>Glycyrrhiza glabra</i> L.	Vol. I
Yamānikā	Yavānī	<i>Trachyspermum ammi</i> (L.) Sprague	Vol. I
Yavānī	Yavānī	<i>Trachyspermum ammi</i> (L.) Sprague	Vol. I
Yavāsaka	Yavāsaka	<i>Alhagi pseudalhagi</i> (Bieb.) Desv.	Vol. II
Yavakṣāra	Yavakṣāra	Water soluble ash of Pl. of <i>Hordeum vulgare</i> L.	Vol. II
Yavakṣāra	Yavakṣāra	Water soluble ash of Pl. of <i>Hordeum vulgare</i> L.	Vol. IV

APPENDIX-10

BIBLIOGRAPHY

1.	Vāgbhaṭa	Azṛṛāḡgah [®] daya, Ed. By Lāla Chanda Vaidya; Motḡlāla Banārasī Dāsa, Vārānasī 1963
2.	Upādhyāya Mādhava	Āyurveda Prakāśa, Ed. By Gularāja Śarmā Miśra
3.	Govinda Dāsa	BhaiZajyaratnāvalī, Red and Ed. Narendra Nātha Miśra, Jayadeva Vidyālaḡkāra, Haridatta Śāstrī, Lālachandajī Vaidya, Motḡlāla Banārasī Dāsa, Vārānasī, Sarvat 2019
4.		Bhārata BhaiZajya Ratnākara Ed. By Gopinātha Gupta, Śivarāmacandra Bhattācārya, Uējhā Pharmacy, 2 nd edition, 1947, Part - III
5.	Cakrapāṇi Datta	Cakradatta, Ed. Jagadīśvara Prasāda, Chaukhamba Saḡskṛta Series Office, Vārānasī 1961
6.	Yādavajī Trikamajī Ācārya	Rasām [®] ta, Motḡlāla Banārasī Dāsa, Vārānasī 1951
7.	Vāgbhaṭa	Rasaratnasamuccaya, Ed. By Dharmānanda Śarmā, Motḡlāla Banārasī Dāsa, New Delhi 2 nd Edition, 1962
8.	Sadānanda Śarmā	Rasataraḡginī, Ed. By Dharmānanda Śāstrī; Motḡlāla Banārasī Dāsa, Vārānasī Sarvat 2016
9.		Sahasrayoga
10.	Śāraḡgdhara Ācārya	Śāraḡgdhara Saḡhitā, Red. Āḡhamalla, Ed. By Paraśu Rāma Śāstrī, Nirnaya Sāgar Press, Bombay 2 nd Edition 1931
11.	Suśruta	Suśruta Saḡhitā, Ed. By Atri Deva Vidyālaḡkāra, Motḡlāla Banārasī Dāsa, Vārānasī 3 rd Edition 1960